**IP$_3$ receptors and Ca$^{2+}$ signals in adult skeletal muscle satellite cells *in situ***

**JORDI MOLGÓ$^1$, CESARE COLASANTE$^{1*}$, DANY S. ADAMS$^{2**}$ and ENRIQUE JAIMOVICH$^3$**

$^1$ Laboratoire de Neurobiologie Cellulaire et Moléculaire, Unité Propre de Recherche 9040, Institut Fédératif de Neurobiologie Alfred Fessard, Centre National de la Recherche Scientifique, 1 avenue de la Terrasse, bâtiments 32-33, 91198 Gif-sur-Yvette cedex, France;  
$^2$ Department of Biological Sciences, Smith College, Northampton, MA 01063, USA;  
$^3$ Centro de Estudios Moleculares de la Célula, Instituto de Ciencias Biomédicas, Facultad de Medicina, Universidad de Chile, Casilla 70005, Santiago 6530499, Chile.  
* Present address: The Forsyth Institute, Cytokine Biology, 140 The Fenway, Boston, MA 02115, USA  
** Current address: Laboratorio de Fisiología de La Conducta, Facultad de Medicina, Universidad de Los Andes, Mérida 5101, Venezuela

**ABSTRACT**

In this short article we review muscle satellite cell characteristics and our studies in adult rodent muscle satellite cells *in situ*. Using confocal laser scanning microscopy and immunocytochemistry, a high level of IP$_3$ receptor (IP$_3$R) immunostaining was detected in satellite cells. These cells were identified by their peripheral position, their size, the shape of their nucleus, the paucity of the apparent cytoplasm, and the immunostaining with specific molecular markers such as $\alpha$-actinin, the neural cell adhesion molecule (N-CAM) and desmin. High extracellular K$^+$ (60 mM) induced long-lasting Ca$^{2+}$ signals in satellite cells *in situ*. We suggest that electrical activity stimulates IP$_3$-associated Ca$^{2+}$ signals that could act in concert with signaling pathways triggered by growth factors and/or hormones.

**Key words:** Muscle satellite cells, Inositol 1,4,5-trisphosphate receptors, intracellular Ca$^{2+}$, Signal transduction
INTRODUCTION

In adult skeletal muscles, there is a population of small fusiform mononucleated cells, usually oriented parallel to the axis of the fibers, which may protrude above its surface as seen by electron microscopy. These cells, which are closely associated with myofibers, were first identified under the electron microscope and were called satellite cells (Katz, 1961; Mauro, 1961). A characteristic property of these cells is that they lie between the plasma membrane of the muscle fiber and the basal lamina (Muir, 1970). Muscle satellite cells have been identified in amphibian (Mauro, 1961; Katz, 1961; Popiela, 1976), reptilian (Kahn and Simpson, 1974), avian (Hartley et al, 1992) and mammalian skeletal muscles including muscle spindles (Gamble et al., 1978; Campion et al, 1981). The number of satellite cells in mammalian skeletal muscles depends on the muscle fiber type. Thus, in the adult extensor digitorum longus, which primarily contains glycolytic fibers, the percentage of satellite cells is lower than in the soleus muscle that contains slow oxidative fibers (Gibson and Schultz, 1982; Snow, 1983; Schultz and McCormick, 1994). Age-related differences in absolute numbers of skeletal muscle satellite cells have been reported in various animal species (Gibson and Schultz, 1983; Nnodim, 2000). In general, the overall number of satellite cells appears higher at birth (Bischoff, 1994; Hawke and Garry, 2001), and decreases during aging (Renault et al., 2002). This reduction in satellite cell number contributes to a diminution of the regenerative capacity of skeletal muscle (Grounds, 1998; Jejurikar and Kuzon, 2003). However, the impaired regenerative response observed with aging appears to be much more complex than just satellite cell senescence (see Bortoli et al., 2003; Conboy et al., 2003).

Morphological characteristics of satellite cells in mature muscles include a high nuclear-to-cytoplasmic ratio, a small organelle content, a smaller nuclear size compared with neighboring myonuclei, and an abundant heterochromatin in their nuclei (Muir et al., 1965; Muir, 1970; Cull-Candy et al., 1980) reflecting their mitotic quiescence (Schultz et al., 1978).

For many years the laborious identification of satellite cells in situ by light microscopy limited their study. However, the use of immunohistochemical techniques with antibodies directed to proteins expressed either in the basal lamina, such as laminin, or in the sarcolemma, like dystrophin, greatly facilitated their identification in fixed muscles. Muscle satellite cells are also usually difficult to identify in vivo under the light microscope, due to their close apposition to the muscle fiber sarcolemma (cleft width about 20 nm), and because they are usually covered by connective tissue overlying the muscle. However, the use of mild enzymatic digestion with collagenase (Cull-Candy et al, 1980), to remove the connective tissue, together with improved Normarski’s or Hoffman’s optic-additions to the light microscope, and the current use of confocal laser scanning microscopy greatly refined their visualization in single living muscle fibers, or bundles consisting of a few fibers of skeletal muscle.

The adult skeletal muscle demonstrates a remarkable capacity to adapt to strenuous activity and/or tissue damage, and satellite cells have been implicated as the major source of myogenic cells involved in growth, and repair of myofibers. The processes by which these physiologic adaptations occur are attributed to the activation, multiplication and fusion of satellite cells with myofibers (for a review see Hawke and Garry, 2001). Activation of satellite cells also plays a role in muscle growth following minimal activity (Kadi and Thornell, 2000) probably without muscle damage. The reaction of satellite cells to such muscle activity would probably not involve factors triggered by damage (e.g. growth factors), which are the major substances regulating the activation, proliferation, and differentiation of satellite cells (McFarland, 1999; Hawke and Garry, 2001). Activation of satellite cells probably without muscle damage. The reaction of satellite cells to such muscle activity would probably not involve factors triggered by damage (e.g. growth factors), which are the major substances regulating the activation, proliferation, and differentiation of satellite cells (McFarland, 1999; Hawke and Garry, 2001), but could relate to muscle depolarization. Little is known concerning signal transduction pathways involved in muscle growth in response to activity (however, see Dunn et al., 2000; Pallafacchina et al., 2002). Understanding the signaling systems involved in directing satellite cells between proliferation and differentiation is of great importance. Many factors, including fibroblast growth factors, insulin-like growth factors, and interleukin-6 cytokines, have already been implicated in the control of satellite cell activity (for reviews see Hawke and Garry, 2001; Charge and Rudnicki, 2004). In the present article we review the evidence that inositol 1,4,5-trisphosphate receptors (IP3Rs) are localized in satellite cells of mature muscle, and that depolarization triggers a long-lasting calcium signaling in those cells (Powell et al., 2003).
ADULT SATELLITE CELLS IN SITU EXPRESS IP\textsubscript{3} RECEPTORS

In adult mouse skeletal muscle, satellite cells can be identified in situ by their location on the periphery of myofibers, by their morphology and characteristic nucleus, and because they are covered by laminin of the basal lamina. A positive immunostaining for IP\textsubscript{3}Rs was obtained in the cytoplasm of satellite cells in situ. In the projected confocal image shown in Figure 1A, a high concentration of IP\textsubscript{3}Rs is suggested by the intensity of the immunostaining in specialized regions of the satellite cell cytoplasm. These cells express the neural cell adhesion molecule (N-CAM) that is found in quiescent, activated and proliferating satellite cells (Hawke and Garry, 2001), and at the neuromuscular junction of adult fibers (Covault and Sanes, 1986), but not in fibroblasts or vascular tissue. Some of the satellite cells contain desmin, an intermediate cytoskeletal filament protein, which is considered a molecular marker of activated cells (Bockhold et al., 1998). Furthermore, satellite cells also contain α-actinin (Powell et al., 2003). Proliferating satellite cells in culture have been also shown to express α-actinin, as well as vimentin and desmin (Van der Ven et al., 1992). In double-immunolabeled muscle fibers, IP\textsubscript{3}Rs were also found in cells identified as satellite cells by the presence of N-CAM, or desmin (Powell et al., 2003).

Figure 1. Confocal images of satellite cell immunostained with various protein markers in adult mouse skeletal muscles. A, projected image (12 sections, spaced by 0.12 μm) showing a duet of satellite cells in situ at the periphery of a myofiber (Levator auris longus muscle) immunostained for IP\textsubscript{3}Rs (green), and stained for nuclei with propidium red (orange). Note the distinct intensity of the IP\textsubscript{3}R-immunostaining in the cell’s cytoplasm, and the IP\textsubscript{3}R cross-striation pattern in the myofiber. B, C and D, single optical sections (0.12 μm thickness) of a cryostat section (10 μm) from an Extensor digitorum longus muscle revealing the presence of laminin in the basal lamina surrounding a duet of satellite cells (B), nuclei, stained with TOTO-3, (C) and α-actinin in the cytoplasm of the satellite cells, and in a cross striated pattern in the myofiber (D). N-CAM immunostaining is shown in another satellite cell (E); this marker is found in quiescent, activated and proliferating cells, but not in fibroblasts.
MUSCLE SATELLITE CELLS RESPOND TO K⁺-INDUCED MEMBRANE DEPOLARIZATION

In myofibers loaded with fluo3- AM and stimulated by a high K⁺ (60 mM) isotonic medium, an increase in cytoplasmic Ca²⁺ in satellite cells was observed (Powell et al., 2003). This calcium does not appear to come from the myofiber, but to be confined to each satellite cell. The Ca²⁺ transients in these cells were long lasting (200-300 sec) with respect to the transients observed in the myofiber. These data, and the fact that IP₃Rs are expressed in satellite cells suggest an IP₃-associated Ca²⁺ signaling in those cells.

Although the results with K⁺-induced depolarization do not reveal how satellite cells can be depolarized in situ, it is conceivable that the K⁺ concentration around the satellite cells could increase during trains of action potentials in the muscle fiber. This elevated level of K⁺ could then depolarize the satellite cells. The dihydropyridine receptor (α₁-subunit) voltage sensor has been shown to trigger a depolarization induced IP₃ cascade in cultured muscle (Araya et al., 2003). It is likely that satellite cells may contain low levels of dihydropyridine receptors, since mRNA encoding skeletal muscle isoforms of the dihydropyridine receptor-α₁-subunit have been reported in human myoblasts (Tanaka et al., 2000). If the signal transduction model for muscle in culture (Powell et al., 2001; Araya et al., 2003) holds true for satellite cells, then an IP₃ cascade would end in Ca²⁺-dependent satellite cell's nuclear activation. The more the particular muscle fiber contracts, the stronger the signal to the satellite cell, and the more likely it would be recruited for proliferation and fusion with the muscle fiber. This mechanism of recruitment might be important in mild exercise (Kadi and Thornell, 2000), in muscle growth after atrophy (Mitchell and Pavlath, 2001) and in muscle regeneration. Alternatively, the depolarization-induced Ca²⁺ signal could act in concert with signaling.
pathways triggered by growth factors and/or hormones (see Jaimovich and Espinosa, this issue).

We hope that the data presented here will stimulate further investigation on IP₃ signaling in satellite cells, which may be important for understanding the mechanisms involved in muscle growth and repair.

ACKNOWLEDGEMENTS

This work was made possible in part by an ECOS Sud-CONICYT (Nº C03S02) exchange program, and was supported in part by grants from the Association Française contre les Myopathies (to J.M), the Direction des Systèmes de Forces et de la Prospective (Grant Nº 02 60 65 093 to J.M.), and from FONDAP (# 15010006 to E.J.).

REFERENCES


* This paper is dedicated to the memory of Dr. Jeanne A. Powell.

Corresponding author: Jordi Molgó, Laboratoire de Neurobiologie Cellulaire et Moléculaire, Unité Propre de Recherche 9040, Institut Fédératif de Neurobiologie Alfred Fessard, Centre National de la Recherche Scientifique, 1 avenue de la Terrasse, bâtiments 32-33, 91198 Gif-sur-Yvette cedex, France. Phone: (33) 1 69 82 36 42, FAX: + (33) 169 82 41 41, E-mail: Jordi.Molgo@nbcm.cnrs-gif.fr