Timing Modulates the Effect of Sleep Loss on Glucose Homeostasis

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Context: Chronobiological factors may modulate the impact of sleep loss on glucose homeostasis. However, these interactions have not been systematically assessed in humans.

Objective: To assess the effect of sleep loss during the late vs early night on glucose homeostasis.

Design: Fifteen normal-weight men participated in three conditions of a randomized, balanced crossover study comprising two conditions with shortened sleep (*i.e.*, 4 hours of sleep during the first or the second half of the night) and a control condition with 8 hours of sleep. Glucose, insulin, cortisol, and glucagon were measured. Insulin sensitivity and secretion were assessed with a Botnia clamp.

Results: Compared with regular sleep duration, sleep loss reduced insulin sensitivity (M-value; P = 0.031) irrespective of early- or late-night timing (P = 0.691). The disposition index (*i.e.*, the β -cell response adjusted for insulin sensitivity) also tended to be impaired by short sleep (P = 0.056) but not by sleep timing (P = 0.543). In contrast, sleep loss in the second half but not the first half of the night induced reductions in morning glucagon and cortisol levels (P < 0.031) followed by a transient increase in cortisol (P < 0.044).

Conclusions: Although sleep deprivation acutely reduced insulin sensitivity irrespective of its nocturnal timing, sleep loss in the early morning compromised α -cell and hypothalamic-pituitary-adrenal axis activity to a greater extent than sleep loss in the first half of the night. This pattern suggests that the timing of sleep restriction can partly potentiate its deleterious metabolic effects. (*J Clin Endocrinol Metab* 104: 2801–2808, 2019)

S hort sleep duration and disrupted sleep patterns have become common in modern societies (1). In parallel, obesity and metabolic comorbidities such as type 2 diabetes mellitus (T2DM) have risen on a worldwide scale (2). Large epidemiological studies show that impaired

ISSN Print 0021-972X ISSN Online 1945-7197 Printed in USA Copyright © 2019 Endocrine Society Received 7 December 2018. Accepted 20 February 2019

Received 7 December 2018. Accepted 20 February 2019. First Published Online 26 February 2019 quantity and quality of sleep are associated with adverse metabolic conditions such as obesity and impaired glucose tolerance [for review see (3)]. A meta-analysis including more than 600,000 adults and 30,000 children clearly revealed a strong association between reduced

Abbreviations: DI, disposition index; FPIR, first-phase insulin response; HEC, hyperinsulinemiceuglycemic clamp; HPA, hypothalamic-pituitary-adrenal; ivGTT, intravenous glucose tolerance test; REM, rapid eye movement; SWS, slow-wave sleep; T2DM, type 2 diabetes mellitus; TST, total sleep time.

sleep duration (≤ 5 hours) and obesity (4). Longitudinal analyses of large population surveys further indicate that both long (~9 hours) and short (~5 hours) sleep durations elevate the risk for development of T2DM (5). However, laboratory studies on the impact of insufficient sleep on glucose metabolism are less unequivocal. Although there is solid evidence that sleep curtailment is associated with reduced peripheral insulin sensitivity of muscles and liver (6–8), results regarding islet function (*i.e.*, first-phase insulin response, basal insulin secretion, and α -cell function) are inconsistent. Some studies have shown impaired islet function (9), whereas others have reported unchanged (6) or even enhanced secretory function (10) after two nights of sleep restricted to 4 hours.

The mechanisms that might mediate detrimental effects of short sleep on energy balance and, in the long run, might favor obesity and T2DM are under intense investigation. Pathophysiological factors such as altered activity of neuroendocrine pathways [e.g., the hypothalamicpituitary-adrenal (HPA) axis (6, 9, 11)], elevated levels of free fatty acids (10), and increased sympathetic nerve activity (6, 9, 11) all appear to contribute to impaired glucose tolerance and positive energy balance after impaired sleep. First evidence exists that chronobiological aspects of sleep restriction affect metabolic conditions (12), but still little is known about whether the timing of sleep loss-in addition to sleep's shortened duration-affects glucose homeostasis. This question was addressed in the present experiment. We subjected healthy young men to three conditions: short sleep (4 hours) during the first half of the night, short sleep (4 hours) during the second half of the night, and regular sleep of 8 hours. We hypothesized that not only sleep curtailment *per se* but also timing of sleep loss and, in particular, waking up early after short sleep aggravates the adverse effects of sleep loss on glucose metabolism.

Material and Methods

Participants

Fifteen healthy normal-weight males aged 20 to 30 years (mean \pm SEM: 24.6 \pm 0.7 years) and with a body mass index between 20.0 and 24.9 kg/m² (23.3 \pm 0.4 kg/m²) were enrolled in this study. Exclusion criteria were chronic or acute illness, current medication of any kind, smoking, elevated alcohol (>50 g per day) and caffeine (>300 mg per day) consumption, use of prescription drugs, diabetes in first-degree relatives, shift work, travel across time zones during the past 4 weeks, and short habitual sleep duration (<6 hours per day). Furthermore, abnormal findings on physical examination or routine laboratory testing (complete blood counts, comprehensive metabolic panel, thyroid function, lipid composition) were also exclusion criteria. Physical activity, sleep behavior, nutrition, and subjective feelings during the day before experimental visits were assessed by a structured interview at each in-laboratory visit to ensure that participants adhered to the above-listed criteria. The study protocol was approved by the ethics committee on research involving humans at the University of Lübeck, and all participants gave written informed consent before participation.

Study design and procedure

Participants underwent an adaptation night in the laboratory that included a standard overnight polysomnographic recording to exclude sleep disorders and for familiarization with the experimental setting. Participants were tested in a randomized, counterbalanced, crossover design on three conditions spaced at least 3 weeks apart. In the week preceding the experimental nights, subjects were asked to maintain a regular sleep schedule with a minimum of 7 hours of sleep per night. Glucose homeostasis was assessed by the Botnia clamp technique (13) on the next morning after (i) one night with 4 hours of sleep during the first half of the night ("late-night sleep loss"; bedtime: 2230 to 0300 hours), (ii) one night with 4 hours of sleep during the second half of the night ("early-night sleep loss"; bedtime: 0215 to 0645 hours), and (iii) one night with regular 8 hours of sleep ("regular sleep"; bedtime: 2215 to 0645 hours).

Participants attended the research unit at 1915 hours for each laboratory night. An IV catheter was inserted into a vein of the participant's nondominant distal forearm. After a standardized light dinner (380 kcal) at 2015 hours, participants were prepared for polysomnographic recording. Thereafter, they were allowed to drink only water (max. 250 mL) until the next morning. Participants went to bed and lights were turned off at 2215 and 2230 hours, respectively, in the regular sleep and late-night sleep loss conditions, whereas they remained awake in a sitting position until 0300 hours in the early-night sleep loss condition. During sleep restriction conditions, participants were allowed to read and to watch nonarousing movies while being monitored constantly by the experimenters. Light intensity in the laboratory was 300 lux.

In the morning after each experimental condition, a second IV catheter was inserted into a vein of the subject's contralateral distal forearm, and a Botnia clamp was performed starting at 0800 hours (13). In brief, the Botnia clamp combines a frequently sampled IV glucose tolerance test (ivGTT) over 60 minutes (i.e., 0800 to 0900 hours) with a hyperinsulinemiceuglycemic clamp (HEC) during the following 120 minutes (i.e., 0900 to 1100 hours) to test for β -cell capacity and peripheral insulin sensitivity. For the ivGTT, a glucose bolus of 0.3 g/kg of body weight of a 20% glucose solution was injected IV directly before time point 0, and subsequent blood samples were collected at 2, 4, 6, 8, 10, 20, 30, 40, 50, and 60 minutes after the glucose load. At minute 60, the HEC was started with continuous fixed infusion rates (1.0 IU/kg of body weight) of shortacting human insulin (Insuman Rapid 40 I.E.; Sanofi-Aventis Deutschland GmbH, Frankfurt am Main, Germany) for 120 minutes. At the same time, plasma glucose concentration was kept stable at 5.5 mmol/L via adjustable infusion of 20% glucose solution. The glucose infusion rate was calculated from dynamics of measured (2- to 3-minute intervals throughout the clamp) plasma glucose by clamp software (EKF Diagnostics GmbH, Barleben, Germany). Approximately 60 minutes after starting the HEC, hepatic glucose production was halted and

blood glucose reached a stable euglycemic plateau, which was maintained over an additional 60 minutes ("euglycemic steady state") until the end of the clamp.

Sleep recordings

For standard polysomnography, electrodes were attached to the scalp (C3, C4) for electroencephalographic recordings above, below, and beside the eyes for horizontal and vertical electrooculograms, and two electrodes were placed on the chin for electromyogram signals. Recordings were performed using a Nihon Kohden amplifier (EEG 4400 series; Nihon Kohden, Rosbach, Germany). Data were scored offline according to standard criteria (14). Because of technical issues, a full sleep data set was available in nine participants.

The following sleep parameters were determined: total sleep time (TST), time spent in non–rapid eye movement (REM) sleep stages 1, 2, 3, and 4 and in slow-wave sleep (SWS; *i.e.*, stages 3 and 4 combined), and REM sleep (all as absolute duration and as percentage of TST).

Blood samples and assays

Blood glucose level was determined immediately by EKF Diagnostics' Biosen C-Line (EKF Diagnostics GmbH, Barleben, Germany). The within-assay variation according to the manufacturer was below 1.5% (mean: 12 mmol/L). Concentrations of insulin and cortisol were assessed by immunoassays (Immulite 2000; Siemens Healthcare Diagnostics, UK) with detection limits and average within-assay coefficients of variation of 2.0 μ UI/mL and \leq 5.3% (insulin) and 5.5 nmol/L and $\leq 7.4\%$ (cortisol). Glucagon concentrations were measured by RIAs (Glucagon RIA; IBL International, Hamburg, Germany) with a within-assay coefficient of variation of <2.5%. All values were determined from stored (-80° C) serum or plasma samples, respectively. Based on the results of the Botnia clamp data, the first-phase insulin response (FPIR) was calculated as the sum of insulin concentrations 2, 4, and 6 minutes after an IV glucose load of mIU/10 min (13). Insulin sensitivity was defined as the ratio of glucose infusion rate and steady-state mean insulin concentrations during the last 60 minutes of the clamp, and the M-value [in mg/(kg·min)] was calculated in addition. Higher M-values reflect better insulin sensitivity and *vice versa*. The disposition index (DI) as a measure of insulin secretion adjusted for insulin sensitivity was calculated as the product of the FPIR and the M-value. The homeostatic model assessment index was calculated with fasting values (15).

Statistical analyses

SPSS 22 (IBM Inc., Chicago, IL) for Mac was used for all analyses, and values are expressed as mean \pm SEM. Analyses of sleep data were based on one-way ANOVA, including the factor "condition" (for late-night sleep loss vs early-night sleep loss vs regular sleep). Analyses of hormonal and neuropsychological data were based on ANOVA for repeated measures, including the factors condition (see preceding text) and "time" (for repeated measurements during the experiment). Helmert contrast tests were used to explore first- vs second-level differences (*e.g.*, "regular sleep" vs "short sleep"). For pairwise comparisons of single time points, the Student *t* test was applied. In all analyses, a *P*-value <0.05 was considered significant.

Results

Sleep

Participants slept 426 \pm 43 minutes in the regular sleep condition, whereas TST was reduced to 249 \pm 18 minutes and 263 \pm 10 minutes in the late-night sleep loss and early-night sleep loss conditions, respectively (*P* = 0.041; Table 1). There was no statistical difference in TST between the two short sleep conditions (*P* = 0.771). The longer sleep duration in the regular sleep condition was primarily due to longer absolute and relative durations of REM sleep (both *P* \leq 0.028), whereas absolute and relative durations of SWS as well as shallow non-REM sleep stages 1 and 2 did not differ between all conditions (all *P* \geq 0.134).

Timing-independent effects

Morning fasting concentrations of glucose and insulin were not different between regular sleep, late-night sleep loss, and early-night sleep loss (glucose: 4.2 ± 0.1 vs

| Table 1. Sleep Parameters | | | | |
|---------------------------|----------------|-----------------------|------------------------|---------|
| | Regular Sleep | Late-Night Sleep Loss | Early-Night Sleep Loss | P Value |
| TST, min | 425.7 ± 43.6 | 249.4 ± 18.6 | 263.3 ± 10.4 | 0.041 |
| Wake, min | 3.9 ± 3.3 | 0.2 ± 0.2 | 0.3 ± 0.3 | 0.374 |
| Wake, % | 0.7 ± 0.6 | 0.08 ± 0.08 | 0.1 ± 0.1 | 0.374 |
| S1 sleep, min | 24.7 ± 8.3 | 12.2 ± 6.8 | 12.5 ± 3.4 | 0.470 |
| S1 sleep, % | 5.6 ± 2.1 | 4.9 ± 2.8 | 4.7 ± 1.2 | 0.717 |
| S2 sleep, min | 232.2 ± 44.0 | 114.0 ± 5.0 | 118.8 ± 7.1 | 0.134 |
| S2 sleep, % | 50.9 ± 18.1 | 46.6 ± 3.8 | 45.1 ± 2.1 | 0.782 |
| SWS, min | 78.1 ± 13.1 | 74.6 ± 16.2 | 88.6 ± 7.1 | 0.930 |
| SWS, % | 21.2 ± 6.0 | 35.4 ± 3.6 | 33.3 ± 1.9 | 0.390 |
| REM sleep, min | 80.6 ± 15.3 | 31.5 ± 8.2 | 38.9 ± 6.3 | 0.006 |
| REM sleep, % | 19.2 ± 3.1 | 11.9 ± 2.8 | 15.0 ± 2.4 | 0.028 |
| MA, min | 6.2 ± 1.1 | 2.1 ± 0.5 | 4.2 ± 1.0 | 0.043 |
| MA, % | 1.4 ± 0.1 | 0.8 ± 0.1 | 1.6 ± 0.4 | 0.370 |

Data are presented as mean \pm SEM.

Abbreviations: MA, movement arousal; S1, stage 1; S2, stage 2; REM, rapid eye movement; TST, total sleep time; SWS, slow-wave sleep.

4.2 ± 0.2 vs 4.2 ± 0.1 mmol/L, P = 0.981; insulin: 4.9 ± 0.7 vs 4.4 ± 0.6 vs 3.6 ± 0.6 μ IU/mL, respectively, P = 0.201).

Glucose concentrations during the ivGTT were markedly affected by sleep restriction (P = 0.016 for the main effect of condition). Thus, blood glucose concentrations after the glucose load decreased later in the earlynight sleep loss condition compared with the regular sleep condition (all P < 0.01 for pairwise comparisons from t = 30 minutes until t = 60 minutes after glucose load), whereas glucose concentrations after late-night sleep loss were intermediate (Fig. 1A). During the subsequent HEC procedure, glucose concentrations remained stable (P = 0.126 for the main effect of time) without any difference between conditions (P = 0.809for the main effect of condition) as intended by study protocol.

Short sleep duration compared with regular sleep duration tended to dampen FPIR after glucose load (P =

0.089 for ANOVA Helmert contrast of regular sleep vs short sleep) (Fig. 1B). Of note, the reduction in FPIR seemed to be more pronounced after late-night sleep loss (*i.e.*, 18.5%; P = 0.053 for pairwise comparison with regular sleep) than after early-night sleep loss (*i.e.*, 9%; P = 0.286 for pairwise comparison). The amount of glucose needed to maintain euglycemia during the hyperinsulinemic clamp procedure was significantly decreased to the same extent after both short sleep conditions compared with the regular sleep condition (P =0.010 for ANOVA Helmert contrast of regular sleep vs short sleep; P = 0.580 for pairwise comparison of earlynight sleep loss vs late-night sleep loss) (Fig. 1C). In accordance, insulin sensitivity as displayed by M-values was markedly reduced-again to same extent-after both late-night sleep loss and early-night sleep loss compared with regular sleep $(7.0 \pm 0.7 \text{ mg/(kg \cdot min) vs})$ $6.8 \pm 2.0 \text{ mg/(kg \cdot min)}$ vs $8.0 \pm 0.9 \text{ mg/(kg \cdot min)}$, respectively; P = 0.031 for ANOVA Helmert contrast of



Figure 1. Glucose and insulin homeostasis after sleep restriction. (A) Glucose concentrations during the ivGTT are shown. ^(a)0.05 < P < 0.01 for regular vs late-night sleep loss; ^{bb}P < 0.01; ^{bbb}P < 0.001 for regular sleep vs early-night sleep loss; ^cP < 0.05 for late-night sleep loss vs early-night sleep loss. (B) FPIR during the ivGTT is shown. *0.05 < P < 0.1. (C) Total amount of glucose infused during the hyperinsulinemic clamp to maintain euglycemia is shown. (D) M-values after regular sleep, early-night sleep loss, and late-night sleep loss is shown. *P < 0.05. Error bars, SEM.

regular sleep vs short sleep; P = 0.691 for pairwise comparison of early-night sleep loss vs late-night sleep loss) (Fig. 1D). Analysis of DI showed a trend for impaired secretory β -cell response after both late-night sleep loss and early-night sleep loss compared with regular sleep (P = 0.056 for ANOVA Helmert contrast of regular sleep vs short sleep). There was likewise no difference between both conditions of reduced sleep duration (P = 0.543 for pairwise comparison).

Timing-dependent effects

Morning fasting concentrations of glucagon were slightly different between conditions (P = 0.072 for the ANOVA main effect of condition). Although glucagon concentrations did not differ after short sleep compared with regular sleep (P = 0.326 for Helmert contrast of short sleep vs regular sleep), glucagon concentration was significantly lower after late-night sleep loss than after early-night sleep loss and regular sleep (P = 0.030 for Helmert contrast of late-night sleep loss vs early-night sleep loss and regular sleep) (Fig. 2A). A similar pattern was found for morning cortisol concentrations (P = 0.015 for Helmert contrast of late-night sleep loss vs early-night sleep loss and regular sleep) (Fig. 2B).

Although starting from different baseline levels, glucagon and cortisol decreased in all conditions (both P < 0.001 for ANOVA main effect of time) to reach comparable concentrations at the end of the Botnia clamp at 1100 hours (P > 0.373 for all respective pairwise comparisons). Because of lower morning concentrations, the decrease in glucagon after late-night sleep loss was less steep than after early-night sleep loss and regular sleep (both P < 0.020 for ANOVA condition × time interaction). Cortisol levels during the complete Botnia clamp between 0800 and 1100 hours differed between conditions (P = 0.044 for ANOVA of condition × time interaction). There was a slight increase in cortisol levels upon starting the experimental procedure at 0800 hours after late-night sleep loss, reaching levels at the end of the ivGTT at 0900 hours that are similar to those observed during baseline after regular sleep and early-night sleep loss (P = 0.942).

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Discussion

In this balanced crossover trial, we examined the effects of the timing of acute sleep loss on glucose homeostasis. Irrespective of its timing, sleep curtailment to 4 hours reduced insulin sensitivity as measured by lower glucose infusion rates during an HEC (*i.e.*, lower M-values). Furthermore, β -cell function as reflected by the DI tended to be impaired by short sleep in general. In contrast, the impact of sleep loss on basal α -cell glucagon secretion depended on its timing, in that sleep loss during the second half but not the first half of the night reduced glucagon concentrations. Likewise, an impairing influence on cortisol concentrations emerged only after sleep deprivation in the second half of the night.

The dramatic decrease in insulin sensitivity by ~16% after both short-sleep conditions is in line with previous studies (8, 11), indicating that the detrimental effects of sleep restriction on insulin sensitivity are not modulated by timing. Although findings on the impairing effect of sleep restriction on insulin sensitivity are relatively consistent (8, 11), data on β -cell capacity have been more inconsistent (6, 9, 10). β -cell capacity has been unchanged after an oral glucose load (6) or even increased



Figure 2. Glucagon and cortisol response after sleep restriction. (A) Glucagon (B) and cortisol concentration under fasting conditions (before glucose application) and during the ivGTT and hyperinsulinemic clamp after regular sleep (open circles), early-night sleep loss (black triangles), and late night sleep loss (black squares) are shown. *P < 0.05; **P < 0.01 for Helmert contrast of late-night sleep loss vs early-night sleep loss and regular sleep. HEC, hyperinsulinemic-euglycemic clamp.

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after moderate sleep restriction (10), whereas Spiegel *et al.* (9) reported on 30% lower β -cell capacity after total sleep deprivation. In the current study, β -cell capacity (*i.e.*, FPIR) was slightly lowered by short sleep, and there was a trend toward more pronounced effect after sleep deprivation in the late night. Therefore, an effect on β -cell capacity from prolonged wakefulness under fasting conditions cannot be completely ruled out.

As in our study, studies that reported no changes in β -cell capacity were designed with identical wake-up times in both control and short-sleep conditions (10, 16). However, a study showing impaired β -cell capacity included different wake-up times (0500 hours in the sleep-restriction condition vs 0900 hours in the recovery condition; *i.e.*, sleep restriction was accompanied by a prolonged awake wake time of 4 hours) (9). In contrast to our study, participants of the aforementioned study experienced sleep restriction over six consecutive nights, which may explain the more pronounced effect on β -cell capacity. However, our finding of unchanged fasting glucose and insulin concentrations is in line with previous data from our group (10) that likewise showed no changes after two nights with 4 hours of sleep each. In contrast, other studies showed that three nights with 4 hours of sleep caused an increase in insulin concentrations while glucose concentrations remained unchanged (16) and that 24 hours of total sleep deprivation impaired insulin secretion (17). Overall, a dose-response relationship between the "intensity" of sleep deprivation and impairment in β -cell activity can be assumed.

Of interest, we found no differences in peripheral insulin sensitivity after late-night sleep loss vs early-night sleep loss, indicating that sleep restriction *per se* impaired glucose tolerance independently of the timing of sleep loss. One explanation may be that the amount of SWS was identical in both conditions with restricted sleep duration. In a previous study (18), selective suppression of SWS during unchanged total sleep duration over three consecutive nights decreased insulin sensitivity. However, here we report comparable SWS in both short-sleep conditions as well as the regular sleep condition. Furthermore, it has been shown that two nights with only 4 hours of sleep and a comparable amount of SWS but reduced non-REM stage 2 and REM sleep dramatically impaired glucose tolerance and insulin sensitivity compared with regular sleep (10). Thus, not only SWS but also stage 2 sleep and total sleep duration may be essential for glucose homeostasis.

Reductions in α -cell activity, as measured by morning glucagon concentrations, depended to a greater extent on the timing of short sleep than on sleep duration *per se*. The morning plasma glucagon level was significantly lower when sleep restriction took place in the second half

of the night than after early-night sleep loss or regular sleep. The impairing effect of sleep loss on glucagon concentration is in line with previous reports by our group (17, 19) and others (16). However, the respective interaction of sleep loss and timing identified in the present experiments allows the tempting assumption that impaired α -cell activity after sleep loss in the late night was due to prolonged preceding wakefulness rather than to the shortening of sleep time. Underlying mechanisms might involve specific inhibition of either α -cell functioning or innervation by the autonomic nervous system (20, 21), as autonomic nervous system activity is critically modulated by prolonged wakefulness.

Differential activity patterns of endogenous stress axes may have contributed to the observed results because increased stress axis activity induces distinct effects on β -cell activity, including reduced insulin secretion and decreased peripheral insulin sensitivity. Cortisol concentrations are highest after awakening and then decline continuously (22). In the current study, morning cortisol concentrations were markedly lower after late-night sleep loss than after early-night sleep loss and regular sleep, but they were relatively elevated during the ivGTT of the Botnia clamp. The sleep-wake cycle modulates the primarily circadian rhythm of cortisol release (23), so the observed reduction and subsequent increase in morning cortisol levels may be attributed to the timing of sleep restriction and moreover to differences in wake-up time. HPA axis activity physiologically peaks during awakening, presumably to prepare the organism for subsequent energy requirements of the waking state (24). The transient increase in cortisol during ivGTT after latenight sleep loss may reflect greater stress susceptibility to the experimental procedures, whereas stress axis activity after regular sleep and early-night sleep loss is already on a level to cope with respective stressors. However, during the subsequent HEC, cortisol concentrations were not different between conditions. Because insulin sensitivity was also decreased after early-night sleep loss, alterations in HPA axis activity are probably not a key mechanism. Although Spiegel et al. (9, 25) reported on dampened amplitude of 24-hour HPA axis rhythm (23) with increased cortisol in the evening after short sleep, impairment in glucose metabolism was already evident in the morning after sleep loss. As supported by earlier studies published by our group (10, 17, 19), this contradicts-at least in part-the hypothesis that alterations in glucose homoeostasis after short sleep are due to increased HPA axis activity.

GH, which was not measured in the current study, may be another factor linking sleep loss and impairments in glucose homeostasis. However, the impact of sleep deprivation on GH secretion appears to depend on the specific sleep restriction protocol. Prolonged GH pulses in the absence of effects on total secretion have been observed during four nights of sleep restriction to 0100 to 0530 hours (26), and acute sleep restriction to 2230 to 0300 hours did not alter morning GH concentrations (19). Mean 24-hour GH concentrations did not differ between bedtime schedules of 5.5 and 8.5 hours imposed over 14 days, although a subgroup of males displayed lower GH concentrations during the first 4 hours of bedtime when sleep restricted (6). Against this backdrop, the influence of (subtle) sleep loss-induced changes in GH secretion in response to the Botnia clamp should be clarified via further studies, which should also control for changes in melatonin arising from differences in the duration of light exposure (27, 28). In the current study, all subjects were exposed to light for at least 1 hour before the clamps so that differences in melatonin presumably could be abated before the start of the clamp protocol.

Several limitations need to be discussed. First, we assessed acute changes in glucose homeostasis after only one night of sleep restriction. Therefore, we cannot make conclusions about the effects of chronic sleep loss. However, 1 day of short sleep evoked detrimental effects on insulin sensitivity and glucose homeostasis, and it is tempting to speculate that short sleep for a longer period would result in even stronger effects. Second, in contrast to the early-night sleep loss condition, the late-night sleep loss condition implied 4 hours of additional wakefulness between awakening and the assessment of glucose homeostasis. Although this schedule allowed us to increase external validity by scheduling the glucoregulatory challenge in the morning hours in all conditions (rather than at night), future studies should control for any effects from additional preclamp wakefulness. Third, our study population comprised young, healthy, normal-weight men. Thus, results cannot be transferred to other groups (e.g., women and subjects with obesity). However, epidemiological studies have shown similar associations between short sleep and glucose homeostasis in women and children over a wide range of body weights (29, 30).

Taken together, sleep loss *per se* impairs glucose homeostasis. However, the timing of sleep restriction can potentiate its deleterious endocrine effects, as α -cell and HPA axis activity are even more affected when waking up early after short sleep. These findings highlight the importance of sleep hygiene and appropriate recommendations in our 24-hour societies to prevent impairment of human glucose homeostasis.

Acknowledgments

Financial Support: This work was supported by grants from the Deutsche Forschungsgemeinschaft (TR-SFB 654 and TR-

SFB 134; to S.M.S.) and from the German Federal Ministry of Education and Research to the German Center for Diabetes Research (DZD e.V.; 01GI0925; to S.M.S. and H.L.).

Author Contributions: S.M.S., M.H., and B.S. designed the study. D.T. and N.F. performed the experiments and collected the data. S.M.S., B.W., M.H., and B.S. analyzed the data. B.W. and S.M.S. wrote the first draft of the manuscript. M.H., D.T., N.F., M.M., R.C., F.S., and H.L. contributed to writing the paper. All authors read and approved the final manuscript. S.M.S. has primary responsibility for the final content.

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Disclosure Summary: The authors have nothing to disclose.

References

- Ford ES, Cunningham TJ, Croft JB. Trends in self-reported sleep duration among US adults from 1985 to 2012. *Sleep (Basel)*. 2015; 38(5):829–832.
- 2. Ng M, Fleming T, Robinson M, Thomson B, Graetz N, Margono C, Mullany EC, Biryukov S, Abbafati C, Abera SF, Abraham JP, Abu-Rmeileh NME, Achoki T, AlBuhairan FS, Alemu ZA, Alfonso R, Ali MK, Ali R, Guzman NA, Ammar W, Anwari P, Banerjee A, Barquera S, Basu S, Bennett DA, Bhutta Z, Blore J, Cabral N, Nonato IC, Chang J-C, Chowdhury R, Courville KJ, Criqui MH, Cundiff DK, Dabhadkar KC, Dandona L, Davis A, Dayama A, Dharmaratne SD, Ding EL, Durrani AM, Esteghamati A, Farzadfar F, Fay DFJ, Feigin VL, Flaxman A, Forouzanfar MH, Goto A, Green MA, Gupta R, Hafezi-Nejad N, Hankey GJ, Harewood HC, Havmoeller R, Hay S, Hernandez L, Husseini A, Idrisov BT, Ikeda N, Islami F, Jahangir E, Jassal SK, Jee SH, Jeffreys M, Jonas JB, Kabagambe EK, Khalifa SEAH, Kengne AP, Khader YS, Khang Y-H, Kim D, Kimokoti RW, Kinge JM, Kokubo Y, Kosen S, Kwan G, Lai T, Leinsalu M, Li Y, Liang X, Liu S, Logroscino G, Lotufo PA, Lu Y, Ma J, Mainoo NK, Mensah GA, Merriman TR, Mokdad AH, Moschandreas J, Naghavi M, Naheed A, Nand D, Narayan KMV, Nelson EL, Neuhouser ML, Nisar MI, Ohkubo T, Oti SO, Pedroza A, Prabhakaran D, Roy N, Sampson U, Seo H, Sepanlou SG, Shibuya K, Shiri R, Shiue I, Singh GM, Singh JA, Skirbekk V, Stapelberg NJ, Sturua L, Sykes BL, Tobias M, Tran BX, Trasande L, Toyoshima H, van de Vijver S, Vasankari TJ, Veerman JL, Velasquez-Melendez G, Vlassov VV, Vollset SE, Vos T, Wang C, Wang X, Weiderpass E, Werdecker A, Wright JL, Yang YC, Yatsuya H, Yoon J, Yoon SJ, Zhao Y, Zhou M, Zhu S, Lopez AD, Murray CJ, Gakidou E. Global, regional, and national prevalence of overweight and obesity in children and adults during 1980-2013: a systematic analysis for the Global Burden of Disease Study 2013. Lancet. 2014;384(9945):766-781.
- 3. Schmid SM, Hallschmid M, Schultes B. The metabolic burden of sleep loss. *Lancet Diabetes Endocrinol*. 2015;3(1):52–62.
- Cappuccio FP, Taggart FM, Kandala N-B, Currie A, Peile E, Stranges S, Miller MA. Meta-analysis of short sleep duration and obesity in children and adults. *Sleep*. 2008;31(5):619–626.
- 5. Gangwisch JE, Heymsfield SB, Boden-Albala B, Buijs RM, Kreier F, Pickering TG, Rundle AG, Zammit GK, Malaspina D. Sleep duration as a risk factor for diabetes incidence in a large U.S. sample. *Sleep*. 2007;**30**(12):1667–1673.
- Nedeltcheva AV, Kessler L, Imperial J, Penev PD. Exposure to recurrent sleep restriction in the setting of high caloric intake and physical inactivity results in increased insulin resistance and reduced glucose tolerance. *J Clin Endocrinol Metab.* 2009;94(9): 3242–3250.

- 7. Leproult R, Van Cauter E. Role of sleep and sleep loss in hormonal release and metabolism. *Endocr Dev.* 2010;17:11–21.
- Donga E, van Dijk M, van Dijk JG, Biermasz NR, Lammers G-J, van Kralingen KW, Corssmit EPM, Romijn JA. A single night of partial sleep deprivation induces insulin resistance in multiple metabolic pathways in healthy subjects. *J Clin Endocrinol Metab.* 2010;95(6):2963–2968.
- 9. Spiegel K, Leproult R, Van Cauter E. Impact of sleep debt on metabolic and endocrine function. *Lancet*. 1999;**354**(9188):1435–1439.
- Schmid SM, Hallschmid M, Jauch-Chara K, Wilms B, Lehnert H, Born J, Schultes B. Disturbed glucoregulatory response to food intake after moderate sleep restriction. *Sleep (Basel)*. 2011;34(3): 371–377.
- Buxton OM, Pavlova M, Reid EW, Wang W, Simonson DC, Adler GK. Sleep restriction for 1 week reduces insulin sensitivity in healthy men. *Diabetes*. 2010;59(9):2126–2133.
- Schmid SM, Hallschmid M, Jauch-Chara K, Wilms B, Benedict C, Lehnert H, Born J, Schultes B. Short-term sleep loss decreases physical activity under free-living conditions but does not increase food intake under time-deprived laboratory conditions in healthy men. *Am J Clin Nutr.* 2009;90(6):1476–1482.
- 13. Tripathy D, Wessman Y, Gullström M, Tuomi T, Groop L. Importance of obtaining independent measures of insulin secretion and insulin sensitivity during the same test: results with the Botnia clamp. *Diabetes Care*. 2003;26(5):1395–1401.
- Rechtschaffen A, Kales A, eds. A Manual of Standardized Terminology, Techniques and Scoring System for Sleep Stages of Human Subjects. Bethesda, MD: US National Institute of Neurological Diseases and Blindness, Neurological Information Network; 1968.
- Matthews DR, Hosker JP, Rudenski AS, Naylor BA, Treacher DF, Turner RC. Homeostasis model assessment: insulin resistance and β-cell function from fasting plasma glucose and insulin concentrations in man. *Diabetologia*. 1985;28(7):412–419.
- Klingenberg L, Chaput J-P, Holmbäck U, Visby T, Jennum P, Nikolic M, Astrup A, Sjödin A. Acute sleep restriction reduces insulin sensitivity in adolescent boys. *Sleep (Basel)*. 2013;36(7): 1085–1090.
- Schmid SM, Hallschmid M, Jauch-Chara K, Bandorf N, Born J, Schultes B. Sleep loss alters basal metabolic hormone secretion and modulates the dynamic counterregulatory response to hypoglycemia. J Clin Endocrinol Metab. 2007;92(8):3044–3051.
- Tasali E, Leproult R, Ehrmann DA, Van Cauter E. Slow-wave sleep and the risk of type 2 diabetes in humans. *Proc Natl Acad Sci USA*. 2008;105(3):1044–1049.

- Schmid SM, Jauch-Chara K, Hallschmid M, Schultes B. Mild sleep restriction acutely reduces plasma glucagon levels in healthy men. *J Clin Endocrinol Metab.* 2009;94(12):5169–5173.
- Taborsky GJ Jr, Ahren B, Mundinger TO, Mei Q, Havel PJ. Autonomic mechanism and defects in the glucagon response to insulin-induced hypoglycaemia. *Diabetes Nutr Metab.* 2002;15(5): 318–322, discussion 322–323.
- Havel PJ, Taborsky GJ Jr. The contribution of the autonomic nervous system to changes of glucagon and insulin secretion during hypoglycemic stress. *Endocr Rev.* 1989;10(3): 332–350.
- Späth-Schwalbe E, Gofferje M, Kern W, Born J, Fehm HL. Sleep disruption alters nocturnal ACTH and cortisol secretory patterns. *Biol Psychiatry*. 1991;29(6):575–584.
- 23. Guyon A, Balbo M, Morselli LL, Tasali E, Leproult R, L'Hermite-Balériaux M, Van Cauter E, Spiegel K. Adverse effects of two nights of sleep restriction on the hypothalamicpituitary-adrenal axis in healthy men. J Clin Endocrinol Metab. 2014;99(8):2861–2868.
- 24. Born J, Hansen K, Marshall L, Mölle M, Fehm HL. Timing the end of nocturnal sleep. *Nature*. 1999;**397**(6714):29–30.
- Spiegel K, Knutson K, Leproult R, Tasali E, Van Cauter E. Sleep loss: a novel risk factor for insulin resistance and type 2 diabetes. *J Appl Physiol (1985)*. 2005;99(5):2008–2019.
- Broussard JL, Chapotot F, Abraham V, Day A, Delebecque F, Whitmore HR, Tasali E. Sleep restriction increases free fatty acids in healthy men. *Diabetologia*. 2015;58(4):791–798.
- 27. Gooley JJ, Chamberlain K, Smith KA, Khalsa SBS, Rajaratnam SMW, Van Reen E, Zeitzer JM, Czeisler CA, Lockley SW. Exposure to room light before bedtime suppresses melatonin onset and shortens melatonin duration in humans. J Clin Endocrinol Metab. 2011;96(3):E463–E472.
- Leproult R, Colecchia EF, L'Hermite-Balériaux M, Van Cauter E. Transition from dim to bright light in the morning induces an immediate elevation of cortisol levels. J Clin Endocrinol Metab. 2001;86(1):151–157.
- Ayas NT, White DP, Al-Delaimy WK, Manson JE, Stampfer MJ, Speizer FE, Patel S, Hu FB. A prospective study of self-reported sleep duration and incident diabetes in women. *Diabetes Care*. 2003;26(2):380–384.
- Hall MH, Okun ML, Sowers M, Matthews KA, Kravitz HM, Hardin K, Buysse DJ, Bromberger JT, Owens JF, Karpov I, Sanders MH. Sleep is associated with the metabolic syndrome in a multiethnic cohort of midlife women: the SWAN Sleep Study. *Sleep* (*Basel*). 2012;35(6):783–790.