## Linkage Disequilibrium Between *IRF6* Variants and Nonsyndromic Cleft Lip/Palate in the Chilean Population

José Suazo,<sup>1</sup> José Luis Santos,<sup>2</sup> Lilian Jara,<sup>1</sup> and Rafael Blanco<sup>1</sup>\*

<sup>1</sup>Programa de Genética Humana, Instituto de Ciencias Biomédicas, Facultad de Medicina, Universidad de Chile, Santiago, Chile

<sup>2</sup>Departamento de Nutrición, Diabetes y Metabolismo, Facultad de Medicina, Pontificia Universidad Católica de Chile, Santiago, Chile

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#### To the Editor:

The genetics of nonsyndromic cleft lip and palate (NSCLP) is complex. Several candidate genes have been associated with this phenotype, but replication studies showed conflicting results [Carinci et al., 2000; Murray, 2002]. Candidate gene analyses indicate that variants in a handful of genes taken together may account for approximately 15–20% of the genetic contribution to NSCLP [Zucchero et al., 2004; Vieira et al., 2005; Riley et al., 2007].

Association of NSCLP and *IRF6* SNPs has been reported [Zucchero et al., 2004]. Strong evidence of over transmission of the ancestral allele of the p.V274I polymorphism to patients with NSCLP was observed in Asians but not in groups of European descent. However, analysis of additional SNPs showed that linkage disequilibrium with NSCLP was also detectable in Iowa and Danish populations. Subsequently, several recent studies have reported genetic variants associated with NSCLP in numerous groups [Blanton et al., 2005; Scapoli et al., 2005; Srichomthong et al., 2005; Morküniené et al., 2006; Park et al., 2007; Vieira et al., 2007].

The aim of this study was to evaluate the role of *IRF6* in NSCLP in the Chilean population, which represents an admixture of Amerindians and Spaniards [Rothhammer et al., 1968]. To accomplish this we performed an association study of *IRF6* SNPs and NSCLP using a case-parents trio design.

The study group comprised 150 unrelated caseparent trios ascertained through unrelated probands affected with NSCLP. Among the cases, 94 were sporadic (17 CL and 77 CL/P) and 56 had a positive family history (9 CL and 47 CL/P). No isolated cleft palate cases were included. Families were recruited from the School of Dentistry, University of Chile, the Dr. Alfredo Gantz Foundation, the Cleft Lip/Palate Center, Luis Calvo Mackenna Hospital and the Cleft Lip/Palate Center, Exequiel Gonzalez Cortes Hospital, located in the Santiago, Chile. Interviews of at least three family members were conducted to provide detailed information for pedigree construction. Each proband and their relatives were evaluated to exclude syndromic CLP. All the families enrolled were of admixed Chilean ancestry and included subjects presenting NSCLP as the unique familial disease. To exclude potential teratogenic influences, a history was taken to evaluate the presence of any neurological disorder in the family or the use of teratogens, such as phenytoin, warfarian, and ethanol.

The Chilean population shows a gradient of ethnicity, Amerindian admixture, genetic markers, socioeconomic strata, and prevalence of NSCLP [Valenzuela, 1988; Palomino et al., 1997]. The families included in this study belong to low to middle-low socioeconomic strata which present the

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<sup>\*</sup>Correspondence to: Dr. Rafael Blanco, Av. Independencia 1027, P.O. Box 70061, Santiago, Chile. E-mail: rblanco@med.uchile.cl

TABLE I. IRF6 SNPs Information and Allele and Haplotype TDT R	lesults
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					<i>P</i> -values for		
SNP	Alleles <sup>a</sup>	MAF <sup>b</sup>	T/NT <sup>c</sup>	P-value <sup>d</sup>	2 SNPs	3 SNPs	4 SNPs
rs764093	T/C	0.363	47/44	0.753	0.094	0.022	0.068
rs2236909	G/A	0.442	73/58	0.190	0.013	0.139	_
rs2235375	C/G	0.448	56/70	0.212	0.307	_	_
rs2235371	C/T	0.262	46/41	0.592	_	_	_

<sup>a</sup>Mayor allele listed first.

<sup>b</sup>MAF: minor allele frequency was estimated using 160 control individuals.

<sup>c</sup>T/NT: transmission/nontransmission count from heterozygous parents are given for the minor allele.

<sup>d</sup>*P*-value for TDT.

<sup>e</sup>Global *P*-value for haplotype analysis. In each case, the *P*-value is place on the line of the first SNP of the haplotype.

highest degree of Amerindian admixture. The study was approved by the Institutional Review Board of the School of Medicine of the University of Chile.

Genomic DNA was extracted from peripheral blood lymphocytes according to the method described by Lahiri and Nurnberger [1991]. Four *IRF6* SNPs (supporting information Table SI may be found in the online version of this article) were chosen based on the HapMap database (http:// www.hapmap.org/cgi-perl/gbrowse/hapmap\_b35/ ) [Carlson et al., 2004] with a selection criterion of  $r^2 > 0.8$  and minor allele frequency (MAF) > 0.2. We used PCR-RFLP for genotyping *IRF6* SNPs. Primers were designed based on the SNP flanking sequence (http://www.ncbi. nlm.nih.gov) and using Primer3 (http://frodo.wi.mit. edu/cgi-bin/primer3/primer3\_www.cgi). Adequate restriction endonucleases for each SNP were selected using the software DNA for Windows version 2.2 (http://www.dna-software.co.uk/).

Simple proportions were used to estimate allele frequencies for the four SNPs (Table I). All the SNPs were in Hardy–Weinberg equilibrium (data not shown) according to an exact test [Schneider et al., 2000]. Lewontin's standardized disequilibrium coefficient (D') [Devlin and Risch, 1995] was calculated using STATA 8 (http://www.gene.cimr.cam.ac.uk/ clayton/software/stata). Strong linkage disequilibrium between pairs of SNPs was observed (Table II).

The Transmission Disequilibrium Test (TDT) [Spielman et al., 1993] was used to assess the differential pattern of excess allele over transmission. TDT analysis of the four individual SNPs showed that none of them presented a significant transmission distortion from heterozygous parents to affected progeny (Table I). Additionally, HBAT, the haplo-type extension of FBAT program [Laird et al., 2000], was used to examine in extended haplotypes possible transmission distortions. The results of the Global *P*-value for TDT haplotype analysis showed significant results for rs764093-rs2235375 (P=0.022) and rs2236909-rs2235375 (P=0.013) (Table I). Table III presents haplotypes that showed significant transmission distortion in two-, three-, and four-

TABLE II.	Linkage Disequilibrium Between IRF6 SNPs Estimated in				
160 Control Individuals					

1	s764093	rs2236909	rs2235375	rs2235371
rs764093 rs2236909 rs2235375 rs2235371	<0.001 <0.001 <0.001	0.882 <0.001 <0.001	0.855 0.943 <0.001	0.853 0.952 0.953

D' are located above the diagonal; P-values are below the diagonal.

TABLE III. Two, Three, and Four SNPs Haplotypes Demonstrating Significantly Altered Transmission

Haplotype	Observed <sup>a</sup>	Expected <sup>b</sup>	P-value
Two markers			
X-A-C-X	149	128	0.039
X-G-X-C	89	100	0.050
Three markers			
C-A-C-X	115	78	0.009
C-A-X-C	90	79	0.046
C-X-C-C	95	84	0.039
T-X-C-C	14	19	0.027
Four markers			
C-A-C-C	110	71	0.008
C-A-G-C	8	12	0.038

Order of SNPs: rs764093-rs2236909-rs2235375-rs2235371. An X indicates that the SNP was not included in creating the haplotype.

<sup>a</sup>Number of individuals that presented the haplotype.

<sup>b</sup>Expected number of individuals that should have presented the haplotype.

marker haplotype analyses. The haplotypes which showed the highest significance shared the allele combination C-A-C for the first three SNPs rs764093rs2236909-rs2235375 (C-A-C-X, P = 0.009 and C-A-C-C P = 0.008). Haplotypes X-A-C-X (P = 0.039), C-A-X-C (P = 0.046), C-X-C-C (P = 0.039) were also over transmitted and partially share the allele combination C-A-C. It is likely that the significant haplotypes are not the risk factors, but rather in linkage disequilibrium with the susceptibility changes in *IRF6*.

Our results differ from previous studies that reported altered allele and haplotype transmission in Asians and Caucasians, which tested SNPs rs2235371 (p.V274I) and rs2235375 [Zucchero et al., 2004; Blanton et al., 2005; Scapoli et al., 2005; Srichomthong et al., 2005; Morküniené et al., 2006; Park et al., 2007; Vieira et al., 2007]. The present findings can be partially explained given that the patients are from an admixed Caucasian-Amerindian population. For example, the p.V274 allele frequency of p.V274I in our sample was approximately 74%, greater than in Asians but lower than in Caucasians [Zucchero et al., 2004; Scapoli et al., 2005; Blanton et al., 2005; Srichomthong et al., 2005; Park et al., 2007]. Zucchero et al. [2004] proposed that p.274V allele shows a three-fold increased risk of recurrence of NSCLP in families with one affected child in Caucasian, Asian, and South American populations. Nevertheless, Hering and Grundmann [2005] do not consider p.274V as a risk allele given that its frequency in Asians, Europeans, Africans, and Pakistani is close to 100%.

Our findings are consistent with the association of *IRF6* gene variations in NSCLP. Functional further studies are needed to establish the role of *IRF6* in the etiology of NSCLP.

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#### REFERENCES

- Blanton SH, Cortez A, Stal S, Mulliken JB, Finnell RH, Hecht JT. 2005. Variation in IRF6 contributes to nonsyndromic cleft lip and palate. Am J Med Genet Part A 137A:259–262.
- Carinci F, Pezzetti F, Scapoli L, Martinelli M, Carinci P, Tognon M. 2000. Genetics of nonsyndromic cleft lip and palate: A review of international studies and data regarding the Italian population. Cleft Palate Craniofac J 37:33–40.
- Carlson CS, Eberle MA, Rieder MJ, Yi Q, Kruglyak L, Nickerson DA. 2004. Selecting a maximally informative set of single-nucleotide polymorphisms for association analyses using linkage disequilibrium. Am J Hum Genet 74:106–120.
- Devlin B, Risch N. 1995. A comparison of linkage disequilibrium measures for fine-scale mapping. Genomics 29:311–322.
- Hering R, Grundmann K. 2005. The IR F6 p.274V polymorphism is not a risk factor for isolated cleft lip. Genet Med 7:209–210.
- Lahiri DK, Nurnberger JI Jr. 1991. A rapid non-enzymatic method for the preparation of HMW DNA from blood for RFLP studies. Nucleic Acids Res 19:5444.
- Laird N, Horvath S, Xu X. 2000. Implementing unified approach to family based tests of association. Genet Epidemiol 19:S36–S42.

- Morküniené A, Steponaviciüté D, Kasnauskienè J, Kucinskas V. 2006. Nucleotide sequence changes in the MSX1 and IRF6 genes in Lithuanian patients with nonsyndromic orofacial clefting. Acta Médica Lituanica 13:219–225.
- Murray JC. 2002. Gene/environment causes of cleft lip and/or palate. Clin Genet 61:248–256.
- Palomino HM, Palomino H, Cauvi D, Barton SA, Chackraborty R. 1997. Facial clefting and Amerindian admixture in populations of Santiago, Chile. Am J Hum Biol 9:225–232.
- Park JW, McIntosh I, Hetmanski JB, Jabs EW, Vander Kolk CA, Wu-Chou YH, Chen PK, Chong SS, Yeow V, Jee SH, Park BY, Fallin MD, Ingersoll R, Scott AF, Beaty TH. 2007. Association between IRF6 and nonsyndromic cleft lip with or without cleft palate in four populations. Genet Med 9:219–227.
- Riley BM, Mansilla MA, Ma J, Daack-Hirsch S, Maher BS, Raffensperger LM, Russo ET, Vieira AR, Dodé C, Mahammadi M, Marazita ML, Murray JC. 2007. Impaired FGF signaling contributes to cleft lip and palate. Proc Natl Acad Sci USA 104:4512–4517.
- Rothhammer F, Lasserre E, Blanco R, Covarrubias E, Dixon M. 1968. Microevolution in human Chilean populations. IV. Shovel shape, mesial-palatal version and other dental traits in Pewenche Indians. Z Morphol Anthropol 60:162–169.
- Scapoli L, Palmieri A, Martinelli M, Pezzetti F, Carinci P, Tognon M, Carinci F. 2005. Strong evidence of linkage disequilibrium between polymorphisms at the IRF6 locus and nonsyndromic cleft lip with or without cleft palate, in an Italian population. Am J Hum Genet 76:180–183.
- Schneider S, Roessli D, Excoffier L. 2000. Arlequin: A software for population genetics data analysis. Geneva: University of Geneva, Genetics and Biometry Laboratory, Department of Anthropology.
- Spielman RS, McGinnis RE, Ewens WJ. 1993. Transmission test for linkage disequilibrium: The insulin gene region and insulindependent diabetes mellitus. Am J Hum Genet 52:506– 516.
- Srichomthong C, Siriwan P, Shotelersuk V. 2005. Significant association between IRF6  $820G \rightarrow A$  and nonsyndromic cleft lip with or without cleft palate in the Tahi population. J Med Genet 42:e46.
- Valenzuela CY. 1988. On sociogenetic clines. Ethol Sociobiol 9:259–268.
- Vieira AR, Avila JR, Daack-Hirsch S, Dragan E, Felix TM, Rahimov F, Harrington J, Schultz RR, Watanabe Y, Johnson M, Fang J, O'Brien SE, Orioli IM, Castilla EE, Fitzpatrick DR, Jiang R, Marazita ML, Murray JC. 2005. Medical sequencing of candidate genes for nonsyndromic cleft lip and palate. PLoS Genet 1:e65.
- Vieira AR, Cooper ME, Marazita ML, Orioli IM, Castilla EE. 2007. Interferon regulatory factor 6 (IRF6) is associated with oralfacial cleft in individuals that originate in South America. Am J Med Genet Part A 143A:2075–2078.
- Zucchero TM, Cooper ME, Maher BS, Daack-Hirsch S, Nepomuceno B, Ribeiro L, Caprau D, Christensen K, Suzuki Y, Machida J, Natsume N, Yoshiura K, Vieira AR, Orioli IM, Castilla EE, Moreno L, Arcos-Burgos M, Lidral AC, Field LL, Liu YE, Ray A, Goldstein TH, Schultz RE, Shi M, Johnson MK, Kondo S, Schutte BC, Marazita ML, Murray JC. 2004. Interferon regulatory factor 6 (IRF6) gene variants and the risk of isolated cleft lip or palate. N Engl J Med 3518:769– 780.