in neighbouring endemic areas. However, substantial work remains to be done to harmonise and validate collected data. Currently, a major concern is the interpretation of molecular data because of the high diversity of the K13 gene: except for four mutations (C580Y, R539T, R543I, and Y493H), no clinical or in-vitro data have been associated with the other K13 mutant alleles.^{1,3} We believe that the scientific community working on artemisinin resistance should now create a consortium around a K13 reference centre that could take over the management and data validation, the harmonisation of molecular biology techniques, the provision of control samples, and the organisation of a quality assurance system, to recommend the best therapeutic options—a prerequisite to move effectively toward malaria elimination.

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- Ariey F, Witkowski B, Amaratunga C, et al. A molecular marker of artemisinin-resistant Plαsmodium falciparum malaria. Nature 2014; **505:** 50–55.
- Ashley EA, Dhorda M, Fairhurst RM, et al. Spread of artemisinin resistance in Plasmodium falciparum malaria. N Engl J Med 2014; 371: 411–23.
- 3 Straimer J, Gnadig NF, Witkowski B, et al. Drug resistance. K13-propeller mutations confer artemisinin resistance in *Plasmodium falciparum* clinical isolates. *Science* 2014; **347**: 428–31.
- 4 Dondorp AM, Nosten F, Yi P, et al. Artemisinin resistance in Plasmodium falciparum malaria. N Engl J Med 2009; **361:** 455–67.
- 5 Roper C, Pearce R, Nair S, Sharp B, Nosten F, Anderson T. Intercontinental spread of pyrimethamine-resistant malaria. *Science* 2004; **305:** 1124.
- 6 Wootton JC, Feng X, Ferdig MT, et al. Genetic diversity and chloroquine selective sweeps in Plasmodium falciparum. Nature 2002; 418: 320–23.
- 7 Trape JF, Pison G, Spiegel A, Enel C, Rogier C. Combating malaria in Africa. Trends Parasitol 2002; 18: 224–30.
- Tun KM, Imwong M, Lwin KM, et al. Spread of artemisinin-resistant Plasmodium falciparum in Myanmar: a cross-sectional survey of the K13 molecular marker. Lancet Infect Dis 2015; published online Feb 20. http:// dx.doi.org/10.1016/S1473-3099(15)70032-0.
- 9 WHO. Emergency response to artemisinin resistance in the Greater Mekong subregion: regional framework for action 2013–2015. Geneva: World Health Organization, 2013.
- 10 Mohon AN, Alam MS, Bayih AG, et al. Mutations in Plasmodium falciparum K13 propeller gene from Bangladesh (2009–2013). Malar J 2014; 13: 431.
- 11 Miotto O, Almagro-Garcia J, Manske M, et al. Multiple populations of artemisinin-resistant Plasmodium falciparum in Cambodia. Nat Genet 2014; 45: 648–55.
- 12 WHO. Status report on artemisinin resistance. September, 2014. World Health Organization: WHO, 2014.

Rotavirus vaccines roll-out in resource-deprived regions

Published Online January 29, 2015 http://dx.doi.org/10.1016/ S1473-3099(14)71089-8 See Articles page 422 Rotaviruses cause 30–50% of severe diarrhoea cases in children younger than 5 years, leading to about 450 000 deaths every year.¹ Infections during the first months of life are protective against symptomatic reinfections later on, setting the stage for vaccine development.²³The existence of four major genotypes— G1[P8], G2[P4], G3[P8], and G4[P8]—created a great challenge because in-vitro studies suggested that antibodies to a specific type neutralised only that type, raising the question of whether it would be necessary for a vaccine to include all common genotypes.

During the 1990s the first licensed vaccine, Rotashield (Wyeth Laboratories, Collegeville, PA, USA), which contained an attenuated simian and three simianhuman reassortant strains of the virus, showed that 70–90% of cases of severe rotavirus disease could potentially be prevented in lower-middle-income and high-income countries with vaccination.⁴ However, intestinal intussusception was induced in about one in 11000 children who received the vaccine, leading to its withdrawal and posing a large challenge for new candidate vaccines because future trials needed to include 60000 children to reasonably assure safety.^{5,6} Post-licensure studies of the second-generation vaccines Rotarix (GlaxoSmithKline, Brentford, UK), which contains a single human attenuated strain, and RotaTeg (Merck, Kenilworth, NJ, USA) based on five human-bovine reassortant strains, suggest an acceptable class effect risk for intestinal intussusception of somewhere between one in 20000 and one in 100 000 individuals.⁷ Importantly, both vaccines showed high efficacy (more than 80%) against severe rotavirus disease in prelicensure studies^{5,6} and against several predominating genotypes. As trials were progressively done in various regions worldwide, it became clear that protective efficacy for both vaccines was lower in resource-deprived countries than in highincome countries⁸ and that efficacy might not be the same among serotypes and genotypes, especially against G2[P4].5.9

First licensed in 2006, these vaccines have been progressively introduced worldwide and dozens of effectiveness trials, done mostly in high or middle-high income countries, have confirmed efficacy rates reported in prelicensure trials. A major unanswered question is how effective these vaccines will be in real-world scenarios in the poorest regions of the world (where diarrhoea mortality is at its highest) and in the presence of varied circulating types. Children might be infected in their first months of life in these regions (where a first infection is not as protective as in higher-income regions) such that children develop several severe episodes of rotavirus disease throughout their first years.^{3,10} Vaccine effectiveness could be substantially lower in these regions, and, thus, meticulous prospective studies are essential for policy decisions and for the potential design and assessment of new vaccine strategies.

In The Lancet Infectious Diseases, Naor Bar-Zeev and colleagues¹¹ report results of the second effectiveness study to be done in Africa (Blantyre, Malawi). In the first study, Michelle Groome and colleagues¹² showed 57% (95% CI 40-68) effectiveness against rotavirus diarrhoea that required a minimum of overnight hospital admission in children in South Africa younger than 2 years who were vaccinated at 6 and 14 weeks of life. Bar-Zeev and colleagues¹¹ report 64% (24-83) effectiveness for reduction of emergency room visits (compared with rotavirus test-negative controls) for rotavirus in children younger than 5 years (94% of samples tested from children younger than 2 years) using an accelerated 6 and 10 week of age schedule with the monovalent human rotavirus vaccine. Early effect was documented with roughly 10% reductions every year in rotavirus detection rates in infants during their first and second years of age, and an overall rate reduction of near 15% after 2 years for all children younger than 5 years. Genotype G2[P4] was the most commonly detected (25% of samples tested), but the vaccine had a lower non-significant effectiveness point estimate of 53% (95% CI -28 to 83) for G2[P4] than it did for G1[P8] (82%, 42-95), strongly suggesting lower effectiveness against this genotype.

That data for vaccine effectiveness in Malawi are similar to, if not better than, those for other efficacy trials is good news and findings can probably be extrapolated to regions with similar socioeconomic conditions. Differential serotype and genotype effectiveness will



have to be continuously monitored and the search for even better vaccines and strategies must continue. Although, the natural history of rotavirus infection and disease in low-resource regions¹⁰ suggests that oral vaccines that mimic protection conferred by natural infections might have reached their maximum effectiveness, this figure is still substantial and vaccines could potentially prevent nearly 300 000 deaths of infants and children every year.

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- Tate JE, Burton AH, Boschi-Pinto C, Steele AD, Duque J, Parashar UD, for the WHO-coordinated Global Rotavirus Surveillance Network. 2008 estimate of worldwide rotavirus-associated mortality in children younger than 5 years before the introduction of universal rotavirus vaccination programmes: a systematic review and meta-analysis. *Lancet Infect Dis* 2012; 12: 136–41.
- 2 Bishop RF, Barnes GL, Cipriani E, Lund JS. Clinical immunity after neonatal rotavirus infection. A prospective longitudinal study in young children. *New Engl J Med* 1983; **309**: 72–76.
- 3 Velazquez FR, Matson DO, Calva JJ, et al. Rotavirus infections in infants as protection against subsequent infections. *New Engl J Med* 1996; 335: 1022–28.
- 4 Perez-Schael I, Guntinas MJ, Perez M, et al. Efficacy of the rhesus rotavirusbased quadrivalent vaccine in infants and young children in Venezuela. New Engl J Med 1997; 337: 1181–87.
- Frank Ruiz-Palacios GM, Perez-Schael I, Velazquez FR, et al. Safety and efficacy of an attenuated vaccine against severe rotavirus gastroenteritis. New Engl J Med 2006; 354: 11–22.

- 6 Vesikari T, Karvonen A, Prymula R, et al. Efficacy of human rotavirus vaccine against rotavirus gastroenteritis during the first 2 years of life in European infants: randomised, double-blind controlled study. *Lancet* 2007; 370: 1757–63.
- 7 Global Advisory Committee on Vaccine Safety, 11–12 December 2013. Wkly Epidemiol Rec 2014; 89: 53–60.
- 8 Soares-Weiser K, Maclehose H, Bergman H, et al. Vaccines for preventing rotavirus diarrhoea: vaccines in use. *Cochrane Database Syst Rev* 2012; 11: cd008521.
- 9 Grant LR, Watt JP, Weatherholtz RC, et al. Efficacy of a pentavalent human-bovine reassortant rotavirus vaccine against rotavirus gastroenteritis among American Indian children. *Pediatr Infect Dis J* 2012; 31: 184–88.
- 10 Gladstone BP, Ramani S, Mukhopadhya I, et al. Protective effect of natural rotavirus infection in an Indian birth cohort. New Engl J Med 2011; 365: 337–46.
- 11 Bar-Zeev N, Kapanda L, Tate JE, et al. Effectiveness of monovalent rotavirus vaccine in infants in Malawi after programmatic roll-out: an observational and case-control study. *Lancet Infect Dis* 2014; published online Jan 29. http://dx.doi.org/10.1016/S1473-3099(14)71060-6.
- 2 Groome MJ, Page N, Cortese MM, et al. Effectiveness of monovalent human rotavirus vaccine against admission to hospital for acute rotavirus diarrhoea in South African children: a case-control study. *Lancet Infect Dis* 2014; **14**: 1096–104.

W Tuberculosis-associated immune reconstitution inflammatory syndrome: a manifestation of adaptive or innate immunity?

Published Online February 9, 2015 http://dx.doi.org/10.1016/ S1473-3099(15)70026-5 See Articles page 429 In *The Lancet Infectious Diseases*, Shruthi Ravimohan and colleagues¹ investigate whether immunological profiles before and after antiretroviral therapy (ART) can distinguish patients co-infected with HIV and tuberculosis who develop tuberculosis-associated immune reconstitution inflammatory syndrome (IRIS) from those who do not develop this disorder and those who have early mortality. The investigators noted decreased pre-ART concentrations of several proinflammatory cytokines in patients with tuberculosisassociated IRIS (eg, interleukin [IL]-6 adjusted odds ratio [OR] per 1 log₁₀ increase 0.40 [95% CI 0.18–0.89]), which resurged during the disease. After ART initiation, the most prominent changes in patients with tuberculosis-



associated IRIS were reported for cytokines related to innate immunity (eg, IL-6: adjusted OR 1.7 [95% CI 1.2-2.5] and tumour necrosis factor [TNF α : 1.5 [1.0-2.2]), whereas recovery of the CD4 T-cell compartment was similar to that shown in control participants who survived without a diagnosis of tuberculosis-associated IRIS.

These findings seem to challenge the fundamental role of T cells in patients with tuberculosis-associated IRIS. Findings from early key reports pointed towards an overproduction of inflammatory T-helper-1 cytokines, such as interferon- γ , as being the driving force behind tuberculosis-associated IRIS.² Lately, however, increasing evidence is emerging implicating the innate immune system in this syndrome.

Why is it that so many studies of tuberculosisassociated IRIS report different or even conflicting observations? In truth, tuberculosis-associated IRIS is a disease with many faces. Even when guidelines proposed by the International Network For The Study of HIV-Associated IRIS are used,³ patients diagnosed with tuberculosis-associated IRIS still present with symptoms that widely vary in type, severity, and timing.⁴ As such, the clinical picture of this disease might not be uniform across different studies, which could lead to observations becoming just as heterogeneous as the disease itself. For example, Ravimohan and colleagues now show the importance of the study of tuberculosis-associated IRIS during the actual IRIS event, when the inflammatory process is peaking. The increase in IL-6 concentrations between baseline and 4 weeks of ART in patients with tuberculosis-associated IRIS was much more pronounced