

Polycystic ovarian morphology in adolescents with regular menstrual cycles is associated with elevated anti-Müllerian hormone

C. Villarroel¹, P.M. Merino^{1,2}, P. López^{1,3}, F.C. Eyzaguirre¹,
A. Van Velzen¹, G. Iñiguez¹, and E. Codner^{1,*}

¹Institute of Maternal and Child Research (I.D.I.M.I.), School of Medicine, University of Chile, Casilla 226-3, Santiago, Chile ²Department of Pediatrics, School of Medicine, University of Chile, Santiago, Chile ³Hospital Clínico San Borja Arriarán, Servicio de Salud Metropolitano Centro, Santiago, Chile

*Correspondence address. Tel: +56-2-977-0862; Fax: +56-2-424-7240; E-mail: ecodner@med.uchile.cl

Submitted on April 4, 2011; resubmitted on June 7, 2011; accepted on June 14, 2011

BACKGROUND: The significance of polycystic ovarian morphology (PCOM) during adolescence is not clear. The aim of this study was to determine the relationship between PCOM and anti-Müllerian hormone (AMH), inhibin B, testosterone and insulin levels in healthy girls during the second decade of life. We also determined whether AMH could be used as a surrogate marker of PCOM during adolescence.

METHODS: Seventy-four non-obese adolescents (age range: 13.5–19.75 years old) with regular menstrual cycles participated in this study. Transabdominal ultrasound and blood samples were obtained during the follicular phase.

RESULTS: PCOM was present in 33.8% of the subjects. Girls with PCOM had higher AMH levels than girls without PCOM (72.5 ± 6.1 versus 33.4 ± 2.6 pmol/l; $P < 0.0001$) and lower FSH levels (5.4 ± 0.3 versus 6.2 ± 0.2 mIU/ml; $P < 0.036$). Similar levels of inhibin B, androgens and LH were observed in girls with and without PCOM. PCOM prevalence and AMH levels were not associated with age ($P = 0.745$ and 0.2 , respectively) or BMI-SDS ($P = 0.951$ and 0.096 , respectively). AMH levels positively correlated with the of 2–5 mm follicle number. AMH levels ≥ 60.15 pmol/l had a sensitivity and specificity of 64.0 and 89.8%, respectively, to diagnose PCOM (area under the curve = 0.873).

CONCLUSIONS: These data confirm that PCOM in healthy non-hyperandrogenic girls with regular menstrual cycles is prevalent and is not associated with hyperandrogenism. The elevated AMH and lower FSH levels observed in healthy girls with regular menses and PCOM suggest that this ovarian pattern is secondary to a larger number of 2–5 mm follicles. An elevated AMH level is suggestive of the presence of PCOM during adolescence.

Key words: polycystic ovarian morphology / hyperandrogenism / polycystic ovarian syndrome diagnosis / adolescence / anti-Müllerian hormone

Introduction

Polycystic ovarian morphology (PCOM) is one of the diagnostic criteria of polycystic ovarian syndrome (PCOS) in adult women according to the Rotterdam and the Androgen and PCOS Society criteria (The Rotterdam ESHRE/ASRM-sponsored PCOS Consensus Workshop Group, 2004; Azziz *et al.*, 2006, 2009). The significance of this ultrasonographic finding during adolescence is controversial, and the International Consensus published by Balen *et al.* (2003) suggested that this diagnostic element should not be employed during adolescence. Even though several recent publications have included PCOM

as a diagnostic criterion for PCOS in adolescents, the presence of this finding has not been validated in this population (Hickey *et al.*, 2009; Nur *et al.*, 2009; Hart *et al.*, 2010a). This lack of consensus regarding the significance of PCOM during this stage of life is explained by the frequent presence of multifollicular ovaries (Adams *et al.*, 1985) that may be confused with polycystic ovaries, and concerns about the diagnostic accuracy of transabdominal ultrasonography (TA-US), the test usually used in adolescents (Balen *et al.*, 2003; Carmina *et al.*, 2010).

Previously, we studied the prevalence of PCOM in a group of 20 normal or overweight girls followed from 2 to 4 years past menarche

and observed that PCOM was observed in one-third of the girls and was not associated with ovulatory dysfunction (Codner et al., 2011). A similar prevalence was reported by Hart et al. (2010a) in girls who were 2 years past menarche. Studies evaluating the prevalence of PCOM in hyperandrogenic or PCOS girls have also shown that one-third of these girls exhibit PCOM (Ibanez et al., 2008; Fruzzetti et al., 2009). However, there is little information available regarding PCOM prevalence in a larger group of girls, during the second decade of life, or the relationship of the ultrasonographic pattern with anti-Müllerian hormone (AMH), inhibin B, androgen and insulin levels.

AMH is a glycoprotein secreted by the granulosa cells of small, growing follicles, and AMH serum levels correlate with the number of small antral follicles (2–5 mm) observed by transvaginal ultrasound in adult women (Pigny et al., 2003, 2006). Whether AMH levels correlate with the number of 2–5 mm follicles observed during adolescence is unknown. AMH levels are slightly elevated in girls with oligomenorrhoea, whereas they have normal or elevated androgen levels (Park et al., 2010a).

Inhibin B is a glycoprotein secreted by larger antral follicles (8–10 mm) (Andersen et al., 2010), and some publications have reported that inhibin B is elevated in adult women with PCOS (Chu et al., 2005; Bayrak et al., 2007). Recently, a positive correlation between inhibin B and antral follicle number (FN) has been described in adolescents (Hart et al., 2010b), but there is no information about the association of this hormone with PCOM during the second decade of life, as well as its use as a diagnostic tool for this condition.

In the current study, we evaluated healthy girls between 1 year after menarche and 19 years old. We postulated that similar to the situation in adults, AMH levels are increased in girls with PCOM, and a relationship between AMH levels and the number of small antral follicles exists, even when measured by TA-US. We also determined whether AMH can be used as a surrogate marker of PCOM during adolescence. In addition, we studied the prevalence of PCOM and its relationship with age, AMH, inhibin B, androgens, insulin and gonadotrophin levels during the second decade of life, performed a cross-sectional study in healthy adolescents who were between 1 year past menarche and 19 years old and had regular menstrual cycles.

Materials and Methods

Subjects

We studied 74 healthy post-menarchal girls between 1 year past menarche and 19 years old (mean 16.3 years old, range: 13.5–19.75 years old) in a cross-sectional study. The subjects were recruited from schools in downtown Santiago, which is a middle-class area. Fifteen girls who had participated in our previous prospective research were included in the present study (Codner et al., 2011); five subjects were excluded due to obesity. Each girl participated in the study only once. The inclusion criteria were as follows: the absence of significant clinical signs of hyperandrogenism, such as moderate-to-severe acne or hirsutism [Ferriman–Gallwey (FG) score ≥ 7], and the presence of regular menstrual cycles, defined as a menstrual cycle length between 21 and 45 days, according to the recent consensus for adolescents developed by the American College of Obstetrics and Gynecology (ACOG) and the American Association of Pediatrics (AAP) (American Academy of Pediatrics et al., 2006). The exclusion

criteria included the following: use of oral contraceptives, steroids or any other type of medication; the presence of other concomitant chronic conditions, such as genetic syndromes, coeliac disease, renal disease, liver disease, cardiac disease or undernourishment; premature pubarche (defined as the appearance of pubic hair in girls younger than 8 years of age); intrauterine growth retardation (neonatal birth/length < 2 SDs for gestational age, according to Chilean standards); and obesity (BMI greater than the 95th percentile). All subjects had normal fasting blood glucose and thyroid hormone levels.

A total of 102 girls from three schools located near our hospital were invited, and 88 girls agreed to participate in this study. In addition, 14 girls were excluded from the study for the following reasons: hirsutism ($n = 8$), hormonal contraception ($n = 2$), obesity ($n = 2$) and cycles longer than 45 days ($n = 2$).

The protocol was approved by the Institutional Review Board of the San Borja Arriarán Hospital. Parents provided informed consent, and volunteers gave their written assent before entering the study. Adolescent girls older than 18 years old signed the consent form.

Study protocol

Girls were evaluated during the follicular phase (Days 1–7 of the cycle). A complete clinical and physical exam was performed. The age at menarche was obtained retrospectively. Cycle length was obtained from the mean length of the last three menstrual cycles. Gynaecological age was defined as the number of years past menarche at the moment of enrolment in the study. The BMI, FG score and waist-to-hip ratio (WHR) were determined. Standard deviation scores (BMI-SDS) were calculated for BMI, using the current NCHS standard curves; these growth curves have been shown to be applicable to contemporary Chilean populations (Youlton and Valenzuela, 1990). Overweight was defined as a BMI between the 85th and 94th percentiles.

An early-morning blood sample was obtained during the follicular phase (Days 1–7 of the cycle) for the measurement of LH, FSH, estradiol, 17-hydroxyprogesterone (17-OHP), dehydroepiandrosterone sulphate (DHEAS), androstenedione, total testosterone, sex hormone-binding globulin (SHBG), AMH and inhibin B. To measure insulin sensitivity, homeostatic model assessment of insulin resistance (HOMA-IR) values were calculated based on the fasting insulin and glucose levels of each subject (Pacini and Mari, 2003).

Ultrasonographic study

PCOM was identified according to the Rotterdam consensus as the presence of either 12 or more follicles measuring 2–9 mm in diameter and/or an ovarian volume (OV) > 10 ml in one or both ovaries (The Rotterdam ESHRE/ASRM-sponsored PCOS Consensus Workshop Group, 2004).

TA-US was performed by a single observer (C.V.) on the same day that the blood sample was obtained. The exam was performed with a 5-MHz transabdominal probe using a Medison SonoAce 6000C (Medison, Seoul, Korea). Measurements were performed in real time, using the highest possible magnification to view the ovaries. The longest medial axis (length) and its corresponding thickness and width were measured to calculate OV. OV was estimated according to the formula for a prolate ellipsoid: $OV = \pi/6 \times \text{length} \times \text{width} \times \text{thickness}$ (Porter, 2008). The FN was established by scanning each ovary from the inner to the outer margins in a longitudinal cross-section. Follicle diameter was obtained from the mean of the maximum and its corresponding perpendicular diameter. All follicles between 2.0 and 9.0 mm were counted. Then, the numbers of follicles measuring between 2–5 mm and 6–9 mm were determined. The intra-observer variation coefficients of the ultrasonographic study were 3.2 and 4.1% for OV and FN, respectively (Codner et al., 2011). In cases where a dominant cyst/

follicle larger than 10 mm was observed, the ultrasonographic exam was repeated in the following menstrual cycle. The ovary with the larger OV and the number of follicles of the ovary with the larger FN was reported.

Laboratory assays

Serum testosterone (sensitivity [S] = 0.0035 nmol/l), androstenedione ([S] = 0.07 nmol/l), 17-OHP ([S] = 0.03 nmol/l), DHEAS ([S] = 0.07 nmol/l) and estradiol ([S] = 18.4 pmol/l) were measured by competitive specific binding radioimmunoassays (Diagnostic Systems Laboratories, Webster, TX, USA), inter-assay coefficients of variation (CVs) were 8.1, 8.9, 7.3, 7.7 and 6.1%, respectively, and intra-assay CVs were 5.3, 4.2, 7.7, 5.3 and 4.1%, respectively.

Serum LH ([S] = 0.10 mIU/ml), FSH ([S] = 0.10 mIU/ml) and SHBG ([S] = 0.5 nmol/l) levels were measured by immunoradiometric assays from Siemens Healthcare Diagnostics (USA). Intra-assay CVs were 6.5% for LH, 3.6% for FSH and 3.9% for SHBG. Inter-assay CVs were 7.6% for LH, 6.2% for FSH and 6.9% for SHBG (Codner *et al.*, 2005). The free androgen index (FAI) was calculated by the formula $[100 \times \text{testosterone (nmol/l)}] / [\text{SHBG (nmol/l)}]$ (Villarreal *et al.*, 2010). The LH/FSH ratio was calculated. Serum AMH was assayed using the AMH/MIS ELISA kit (Immunotech-Beckman, Marseilles, France) as described previously (Codner *et al.*, 2007). The AMH assay had a sensitivity of 0.7 pmol/l and intra- and inter-assay CVs of 5.3 and 8.7%, respectively. Serum inhibin B was measured using a specific two-site ELISA assay (Diagnostic Systems Laboratories). The assay sensitivity was 7 pg/ml, and the intra- and inter-assay CVs were 4.8 and 7.1%, respectively (Rey *et al.*, 2005; Soto *et al.*, 2009).

Statistical analysis

Normal distributions were evaluated using the Kolmogorov–Smirnov test. 17-OHP did not pass the normality test and was transformed to its logarithm; after being logarithmically transformed 17-OHP levels showed a normal distribution. The other hormones measured passed the normality test.

Continuous variables observed in girls who did or did not fulfil PCOM criteria were compared using the Student's *t*-test. Logistic regression analysis was performed to determine the effects of chronological age, age at menarche, gynaecological age and BMI-SDS on the presence of PCOM. Pearson's correlation analysis was performed to evaluate the relationships between continuous variables.

Receiver-operating characteristic (ROC) analysis was employed to determine the diagnostic accuracy of AMH, inhibin B, testosterone, FSH serum levels and FAI for the diagnosis of PCOM. A significance level of 5% was employed. All statistical calculations were performed using SPSS for Windows, version 18.0 and GraphPad Prism version 5.0. The data are shown as arithmetic means \pm SEM, except for the FG score and the data presented in Fig. 1, which are shown as the median, 5th and 95th percentile.

Results

The clinical characteristics of all participants according to the presence of PCOM are shown in Table 1. PCOM was observed in 33.8% of the girls. An OV > 10 ml in at least one ovary was observed in 19 girls (25.7%). The presence of 12 follicles or more between 2 and 9 mm was observed in 13 girls (17.6%). Girls with and without PCOM

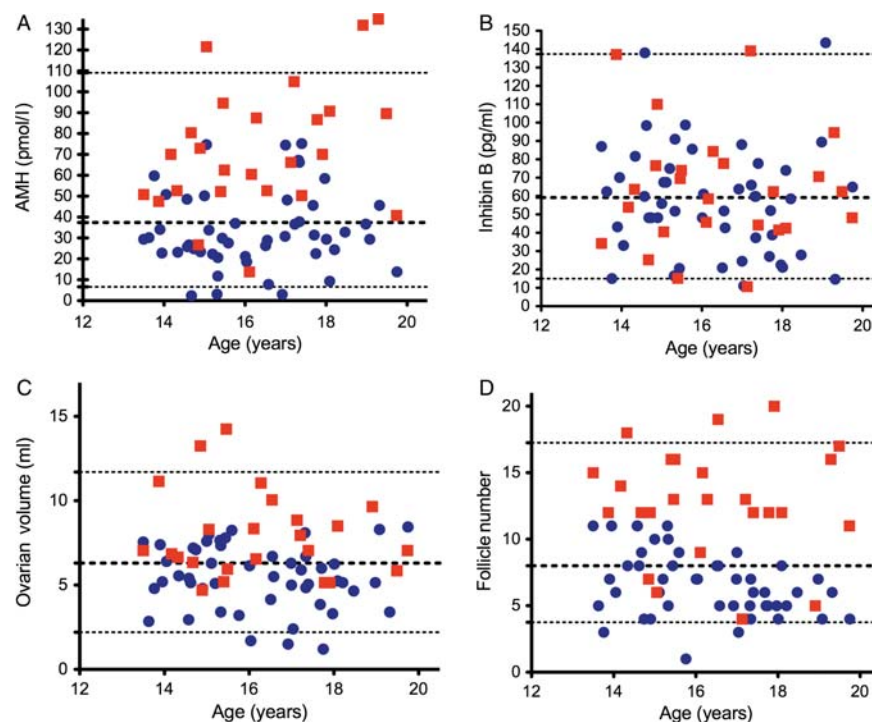


Figure 1 Serum AMH levels, inhibin B levels and ultrasonographic findings according to age and the presence of PCOM in 74 healthy girls during the second decade of life. Black dotted lines represent the median, 5th percentile and 95th percentile. Red squares represent girls with PCOM. Blue circles represent girls without PCOM.

Table I Clinical characteristics according to ovarian morphology.

	PCOM (-)	PCOM (+)
n (%)	49 (66.2)	25 (33.8)
Age (years)	16.2 ± 0.2	16.4 ± 0.4
Age of menarche (years)	12.2 ± 0.1	12.0 ± 0.2
Gynaecological age (years)	3.6 ± 0.3	4.0 ± 0.5
BMI (kg/m ²)	22.7 ± 0.4	22.5 ± 0.5
BMI-SDS	0.5 ± 0.1	0.5 ± 0.1
Menstrual cycle (days)	30.0 ± 0.4	31.4 ± 0.8
Overweight (%)	15 (30.6)	4 (16.0)
Height (cm)	158.6 ± 0.8	160.3 ± 1.0
Waist-to-hip ratio	0.8 ± 0.01	0.8 ± 0.01
Ferriman–Gallwey	2.0 (0.0–6.0)	3.0 (0.0–7.7)

Data are shown as means ± SEM. FG scores are shown as medians (5th to 95th percentile). BMI, body mass index; BMI-SDS, body mass index-standard deviation score.

had a similar chronological age ($P = 0.723$), age at menarche ($P = 0.623$), gynaecological age ($P = 0.391$), BMI-SDS ($P = 0.938$), length of the menstrual cycle ($P = 0.098$), overweight prevalence ($P = 0.174$), WHR ($P = 0.344$) and FG scores ($P = 0.671$). Logistic binary regression showed that gynaecological age ($P = 0.397$), chronological age ($P = 0.745$), age at menarche ($P = 0.617$), length of menstrual cycle ($P = 0.095$) and BMI-SDS ($P = 0.951$) were not associated with PCOM.

The hormonal profiles and ultrasonographic characteristics of girls without PCOM and with PCOM are shown in Table II. Higher AMH levels and lower FSH levels were observed in girls with PCOM ($P < 0.0001$ and 0.036 , respectively). AMH levels were higher in girls who fulfilled the OV criteria of PCOM (> 10 ml) compared with those with $OV < 10$ ml (76.2 ± 11.1 and 40.3 ± 2.9 , respectively; $P = 0.007$), and the same results were observed in girls who met the FN criteria of PCOM compared with those who did not meet the FN criteria (74.3 ± 5.3 and 37.0 ± 3.2 , respectively; $P < 0.0001$). Inhibin B levels ($P = 0.432$), testosterone ($P = 0.674$), androstenedione ($P = 0.696$), LH ($P = 0.651$), the LH/FSH ratio ($P = 0.159$), SHBG ($P = 0.939$), insulin levels ($P = 0.163$), HOMA-IR ($P = 0.127$) and FAI ($P = 0.440$) were similar in girls with and without PCOM and in girls who fulfilled the criteria of PCOM defined by either OV or FN. As expected, the presence of PCOM was associated with a larger OV and FN than in the absence of PCOM. A larger number of follicles with a 2–5-mm diameter ($P < 0.0001$) were observed in girls with PCOM compared with those without PCOM, but there was no difference in the number of follicles with a 6–9-mm diameter ($P = 0.295$).

Figure 1 shows individual AMH and inhibin B levels and ultrasonographic findings according to age and the presence of PCOM. AMH (median: 37.4 pmol/l; p5–p95: 6.6–109.1 pmol/l, Fig. 1A) and inhibin B levels (median: 59.2 pg/ml; p5–p95: 15.0–137.3 pmol/l; Fig. 1B) did not correlate with age or BMI-SDS (Table III). Similarly, no correlation was observed between OV (median: 6.9 ml; p5–p95: 2.3–15.1 ml; Fig. 1C) and FN (median: 8.0; p5–p95: 3.7–17.3;

Table II Hormonal profiles and ultrasonographic characteristics according to ovarian morphology.

	PCOM (-)	PCOM (+)
n (%)	49	25
AMH (pmol/l)	33.4 ± 2.6**	72.5 ± 6.1**
Inhibin B (pg/ml)	57.4 ± 4.2	63.3 ± 6.5
LH (mIU/ml)	3.9 ± 0.3	4.2 ± 0.5
FSH (mIU/ml)	6.2 ± 0.2*	5.4 ± 0.3*
LH/FSH	0.6 ± 0.1	0.9 ± 0.1
Estradiol (pmol/l)	169.2 ± 12.5	167.3 ± 13.6
Testosterone (nmol/l)	1.6 ± 0.1	1.6 ± 0.1
SHBG (nmol/l)	53.3 ± 3.3	52.9 ± 3.4
Free androgen index (%)	3.8 ± 0.4	3.4 ± 0.3
DHEAS (nmol/l)	4485 ± 254	4482 ± 362
Androstenedione (nmol/l)	5.6 ± 0.3	5.9 ± 0.3
17OH-Progesterone (nmol/l)	3.3 ± 0.3	4.5 ± 0.9
Glucose (mmol/l)	4.5 ± 0.1	4.3 ± 0.1
Insulin (mIU/ml)	6.5 ± 0.4	5.7 ± 0.5
HOMA-IR	1.3 ± 0.1	1.1 ± 0.1
Ovarian volume (ml)	6.3 ± 0.3**	9.9 ± 0.7**
Follicle number (n)	6.6 ± 0.4**	12.8 ± 0.8**
FN, follicle 2–5 mm (n)	5.2 ± 0.4**	11.2 ± 0.9**
FN, follicle 6–9 mm (n)	1.9 ± 0.3	2.4 ± 0.5

Results are reported as means ± SEM. The analysis was performed using Student's *t*-test *P*. Conversion to metric units: testosterone, nmol/l × 0.2882 = ng/ml; androstenedione, nmol/l × 0.2865 = ng/ml; DHEAS, nmol/l × 370.37 = ng/ml; estradiol, pmol/l × 0.2725 = pg/ml; 17OH progesterone, nmol/l × 0.3300 = ng/ml; SHBG, nmol/l/ 34.67 = µg/dl; AMH, pmol/l × 0.1400 = µg/l; glucose mmol/l/0.0555 = mg/dl.

*PCOM⁺ versus PCOM⁻: $P < 0.05$.

**PCOM⁺ versus PCOM⁻: $P < 0.0001$.

Table III Relationships between AMH, inhibin B, FSH levels and insulin, androgens and ovarian parameters.

	AMH	Inhibin B	FSH
Age	r 0.195	-0.068	-0.131
BMI-SDS	r -0.151	0.119	-0.100
Ovarian volume	r 0.396**	0.339**	-0.281*
Follicle number	r 0.376**	0.183	-0.141
2–5 mm follicles number	r 0.299**	0.205	-0.170
6–9 mm follicle number	r 0.132	0.004	-0.079
FSH	r -0.193	0.164	-
Testosterone	r 0.124	0.150	-0.179
FAI	r 0.081	0.074	-0.145
Insulin	r -0.311**	0.040	-0.035
HOMA-IR	r -0.306**	0.095	-0.041

The results are shown as the Pearson *r* correlation.

* $P < 0.05$.

** $P < 0.01$.

Table IV Comparison of ROC plot analysis of different ovarian criteria for PCOM.

Hormone	Area under the curve	Cut-off value (pmol/l)	Sensitivity (%)	Specificity (%)	PPV (%)	NPV (%)	P-value
AMH levels	0.873	50.25	84.0	83.7	83.7	84.0	<0.0001
		60.15	64.0	89.8	86.3	71.4	<0.0001
		70.10	48	99.9	99.8	65.8	<0.0001
Inhibin B	0.542	61.75	52	59.2	56.0	55.2	0.560
Free androgen index	0.517	3.15	52	55.2	53.7	53.5	0.815
Testosterone	0.506	0.5	44	61.2	53.1	52.2	0.936

PPV, positive predictive value; NPV, negative predictive value; AUC, area under the curve.

Fig. 1D) with age ($r = -0.113$, $P = 0.341$ and $r = 0.007$, $P = 0.956$, respectively).

The correlations between AMH, inhibin B and FSH levels with the clinical characteristics, ultrasonographic findings and measured hormone levels are shown in Table III. AMH levels exhibited positive correlations with OV, FN and the size of the 2–5-mm follicle pool, but not with the size of the 6–9-mm follicle pool. A negative correlation between AMH with insulin levels and the HOMA-IR index was observed. No correlation was observed between AMH and FSH with androgen levels. Inhibin B and FSH levels showed a positive and negative correlation, respectively, with OV, but showed no correlation with FN. Testosterone and FAI did not exhibit a significant correlation with inhibin B, FSH, OV and FN.

The ROC analysis is shown in Table IV. The area under the curve of the ROC for AMH reached a value of 0.873 [0.782–0.963, 95% confidence interval (CI)]. Several threshold values of serum AMH levels were analysed in terms of specificity and sensitivity from the ROC curve data. The best compromise between specificity (89.2%) and sensitivity (64%) was obtained with a cut-off value of 60.15 pmol/l with a good positive predictive value (86.3%) and negative predictive value (71.4%). The diagnostic accuracy of AMH levels for PCOM defined by OV > 10 ml [area under the curve (AUC) = 0.767, 95% CI = 0.607–0.927] was lower than PCOM defined by FN > 12 (AUC = 0.894 95% CI = 0.824–0.965). ROC analyses showed that testosterone, FAI, FSH and inhibin B had no significant diagnostic utility for detecting PCOM, as shown by a small area under the ROC curve.

Discussion

We report a study of ovarian function in 74 non-obese adolescent girls with regular menstrual cycles and confirmed that PCOM is a prevalent condition in these girls that is not associated with hyperandrogenism or insulin resistance; therefore, this ultrasonographic pattern may correspond to a physiological finding in these girls. In addition, we showed that PCOM is associated with elevated AMH and lower FSH levels, which suggest that this ovarian pattern is secondary to a higher number of 2–5 mm follicles present in these adolescents. Similar elevations in AMH levels were observed in girls who fulfilled PCOM criteria by OV or FN.

We observed that 33.8% of the adolescents in our study exhibited PCOM defined by the Rotterdam criteria, similar to the 35.4% prevalence of PCOM recently reported in healthy girls who were 2 years

post-menarche (Hickey *et al.*, 2009; Hart *et al.*, 2010a) and the 38 and 37% observed in adolescents with hyperandrogenism or PCOS, respectively (Ibanez *et al.*, 2008; Fruzzetti *et al.*, 2009). In contrast, adult women with hyperandrogenism or PCOS exhibit a large difference in the prevalence of PCOM compared with healthy women (80–95 versus 15–25%, respectively) (Polson *et al.*, 1988; Azziz *et al.*, 2006; Codner *et al.*, 2006; Barber *et al.*, 2009; March *et al.*, 2010), suggesting a stronger association of PCOM with hyperandrogenism/PCOS in adult women than in adolescents.

It has been reported that the prevalence of PCOM varies with age (Murphy *et al.*, 2006; Duijkers and Klipping, 2010; Johnstone *et al.*, 2010) and is associated with a physiological decline in FN and OV (Alsamarai *et al.*, 2009). Recent research has shown that PCOM is more prevalent in adult women younger than 30 years old than in older women (Murphy *et al.*, 2006; Duijkers and Klipping, 2010; Johnstone *et al.*, 2010). We did not find an association between OV, follicle count or PCOM prevalence and age, suggesting that the prevalence of PCOM does not change during the second decade of life in girls.

Another important finding was that AMH levels were relatively constant during the second decade of life. These findings are similar to the ones reported by Hagen *et al.* (2010), which recently reported a wide range of normal levels of AMH in healthy girls during adolescence. It is possible that a third of these girls with elevated AMH levels described by Hagen *et al.* (2010) could have PCOM, which would explain the wide range of normal values found in this study.

The negative association between insulin, HOMA-IR and AMH levels has been previously reported in healthy adult women (Chen *et al.*, 2008; Park *et al.*, 2010b). It has been postulated that in women without PCOS, insulin resistance might exert its negative effect on the inhibitory action of AMH on follicular development (Durlinger *et al.*, 2001, 2002a,b; Jonard and Dewailly, 2004) and, therefore, increase the sensitivity of granulosa cells to FSH.

The lower levels of FSH observed in girls with PCOM, in addition to the negative correlation between FSH and OV, suggest that PCOM is not mediated by gonadotrophin action, but rather that this diminished FSH level is associated with an increased follicular mass (Durlinger *et al.*, 2002a). The levels of FSH are regulated by inhibin B serum levels, which depend on follicular mass (Knight and Glistler, 2001; Rosencrantz *et al.*, 2010). We did not find an association between these two hormones that may be explained by the fact that the serum sample was obtained in early follicular phase, a moment of the menstrual cycle that is not the appropriate for the measurement of inhibin B levels (Andersen *et al.*, 2010) or by

the high variability that this hormone shows during the follicular phase (Roberts, 2010).

Previous studies in adolescent girls and adult women with polycystic ovaries and menstrual irregularities have shown higher androgen, insulin and LH levels and lower SHBG levels compared with individuals without polycystic ovaries (Norman et al., 1995; Venturoli et al., 1995; Carmina et al., 1997; van Hooff et al., 2000; Adams et al., 2004; Johnstone et al., 2010). In contrast, our data show that the presence of PCOM in a girl with normal menstrual cycles, defined by a length of 21–45 days (American Academy of Pediatrics et al., 2006), is not associated with hyperandrogenism or elevated insulin levels.

In the current study, we determined through ROC analysis that an AMH cut-off level of 60.15 pmol/l is the best diagnostic accuracy for detecting PCOM, with a sensitivity and specificity for the diagnosis of PCOM of 64.0 and 89.8%, respectively. This cut-off level and sensitivity are similar to the results described by Pigny et al. of 60 pmol/l, using the same assay, in adult women with PCOS (Pigny et al., 2006). In addition, the diagnostic accuracy of AMH levels for the diagnosis of PCOM was somewhat lower when the diagnosis was based on the OV criteria compared with the FN criteria. However, we demonstrated a better diagnostic accuracy of AMH for the detection of PCOM in adolescents compared with the accuracy described by Hart et al. (2010a). The differences in these results may be explained by the fact that in the latter study, the authors evaluated girls with menstrual irregularities, a confounding factor that also leads to elevated AMH levels and, hence, may have decreased the discriminating value of the AMH assay (Park et al., 2010a). The AMH levels found in asymptomatic girls with PCOM are similar to the levels we have recently shown in hyperandrogenic adult women with PCOS using the same assay (Codner et al., 2007). However, the AMH levels we found in healthy adolescents with PCOM in our study are higher than in girls with oligomenorrhea, as previously reported by Park et al. (2010a) (72.5 ± 6.1 versus 37.8 pmol/l, respectively). This finding suggests that though AMH levels may be mildly elevated in girls with oligomenorrhea, the degree of this elevation is not comparable to girls with PCOM.

Previously, we prospectively followed 20 girls from 2 years past menarche and performed annual ultrasonographic studies and assessment of ovulation. We observed a normal ovulatory rate in girls with PCOM and concluded that 'PCOM is an inconstant finding in healthy adolescents and does not appear to be associated with decreased ovulatory rate or metabolic abnormalities in healthy adolescents. This finding suggests that PCOM may correspond to a physiological condition during early adolescence' (Codner et al., 2011). In agreement with our previous publication, the current research, in which a larger number of girls were studied, did not find hyperandrogenism or insulin resistance in girls with PCOM. Because PCOM was an inconstant finding throughout the study period of our previous paper, it is possible that elevated AMH levels may also vary with time. A limitation of the current study is the lack of long-term follow-up study of the girls with PCOM and elevated AMH levels.

In summary, we show that PCOM in healthy adolescents is associated with elevated AMH and lower FSH levels, without biochemical hyperandrogenism. These data suggest that PCOM in a normal weight adolescent with regular menstrual cycles is a physiological ovarian pattern secondary to a larger number of 2–5 mm follicles. In addition, the prevalence of PCOM observed in our study is

similar to previous studies performed in hyperandrogenic adolescents, suggesting that this ultrasonographic finding is not clearly associated with this disorder. These data confirm that PCOM in girls with normal menstrual cycles is a physiological condition that may be linked to a larger follicle mass. An elevated AMH level is suggestive of the presence of PCOM, and this hormone could be used as a surrogate marker of PCOM in adolescents with regular menstrual cycles. Future studies should evaluate the long-term outcome of high AMH levels and PCOM in healthy girls.

Authors' roles

C.V. contributed with the study design, execution, analysis, manuscript drafting and its critical discussion. P.M.M. participated in the execution, manuscript drafting and critical discussion. P.L., F.C.E. and A.V. participated in the execution of the study protocol. G.I. contributed with the execution and manuscript drafting. E.C. contributed to the conception, design and analysis and interpretation of data, manuscript drafting and critical discussion.

Funding

Supported by the Fondo Nacional de Desarrollo Científico y Tecnológico (FONDECYT), grant no. 1100123 to E.C.

References

- Adams J, Franks S, Polson DW, Mason HD, Abdulwahid N, Tucker M, Morris DV, Price J, Jacobs HS. Multifollicular ovaries: clinical and endocrine features and response to pulsatile gonadotropin releasing hormone. *Lancet* 1985;**2**:1375–1379.
- Adams J, Taylor A, Crowley W, Hall JE. Polycystic ovarian morphology with regular ovulatory cycles: insights into the pathophysiology of polycystic ovarian syndrome. *J Clin Endocrinol Metab* 2004;**89**:4343–4350.
- Alsamarai S, Adams JM, Murphy MK, Post MD, Hayden DL, Hall JE, Welt CK. Criteria for polycystic ovarian morphology in polycystic ovary syndrome as a function of age. *J Clin Endocrinol Metab* 2009;**94**:4961–4970.
- American Academy of Pediatrics, Committee on Adolescence, American College of Obstetricians Gynecologists and Committee on Adolescent Health Care. Menstruation in girls and adolescents: using the menstrual cycle as a vital sign. *Pediatrics* 2006;**118**:2245–2250.
- Andersen CY, Schmidt KT, Kristensen SG, Rosendahl M, Byskov AG, Ernst E. Concentrations of AMH and inhibin-B in relation to follicular diameter in normal human small antral follicles. *Hum Reprod* 2010; doi: 10.1093/humrep/deq019.
- Azziz R, Carmina E, Dewailly D, Diamanti-Kandarakis E, Escobar-Morreale HF, Futterweit W, Janssen OE, Legro RS, Norman RJ, Taylor AE et al. Criteria for defining polycystic ovary syndrome as a predominantly hyperandrogenic syndrome: an Androgen Excess Society Guideline. *J Clin Endocrinol Metab* 2006;**91**:4237–4245.
- Azziz R, Carmina E, Dewailly D, Diamanti-Kandarakis E, Escobar-Morreale HF, Futterweit W, Janssen OE, Legro RS, Norman RJ, Taylor AE et al. The Androgen Excess and PCOS Society criteria for the polycystic ovary syndrome: the complete task force report. *Fertil Steril* 2009;**91**:456–488.

- Balen AH, Laven JSE, Tan S-L, Dewailly D. Ultrasound assessment of the polycystic ovary: international consensus definitions. *Hum Reprod Update* 2003;**9**:505–514.
- Barber TM, Alvey C, Greenslade T, Gooding M, Barber D, Smith R, Marland A, Wass JA, Child T, McCarthy MI et al. Patterns of ovarian morphology in polycystic ovary syndrome: a study utilising magnetic resonance imaging. *Eur Radiol* 2009;**20**:1207–1213.
- Bayrak A, Terbell H, Urwitz-Lane R, Mor E, Stanczyk FZ, Paulson RJ. Acute effects of metformin therapy include improvement of insulin resistance and ovarian morphology. *Fertil Steril* 2007;**87**:870–875.
- Carmina E, Wong L, Chang L, Paulson RJ, Sauer MV, Stanczyk FZ, Lobo RA. Endocrine abnormalities in ovulatory women with polycystic ovaries on ultrasound. *Hum Reprod* 1997;**12**:905–909.
- Carmina E, Oberfield SE, Lobo RA. The diagnosis of polycystic ovary syndrome in adolescents. *Am J Obstet Gynecol* 2010;**203**:201.e201–201.e205.
- Chen MJ, Yang WS, Chen CL, Wu MY, Yang YS, Ho HN. The relationship between anti-Müllerian hormone, androgen and insulin resistance on the number of antral follicles in women with polycystic ovary syndrome. *Hum Reprod* 2008;**23**:952–957.
- Chu MC, Carmina E, Wang J, Lobo RA. Müllerian-inhibiting substance reflects ovarian findings in women with polycystic ovary syndrome better than does inhibin B. *Fertil Steril* 2005;**84**:1685–1688.
- Codner E, Mook-Kanamori D, Bazaes RA, Unanue N, Sovino H, Ugarte F, Avila A, Iniguez G, Cassorla F. Ovarian function during puberty in girls with type I diabetes mellitus: response to leuprolide. *J Clin Endocrinol Metab* 2005;**90**:3939–3945.
- Codner E, Soto N, Lopez P, Trejo L, Avila A, Eyzaguirre FC, Iniguez G, Cassorla F. Diagnostic criteria for polycystic ovary syndrome and ovarian morphology in women with type I diabetes mellitus. *J Clin Endocrinol Metab* 2006;**91**:2250–2256.
- Codner E, Iniguez G, Villarreal C, Lopez P, Soto N, Sir-Petermann T, Cassorla F, Rey RA. Hormonal profile in women with polycystic ovarian syndrome with or without type I diabetes mellitus. *J Clin Endocrinol Metab* 2007;**92**:4742–4746.
- Codner E, Villarreal C, Eyzaguirre FC, López P, Merino PM, Pérez-Bravo F, Iniguez G, Cassorla F. Polycystic ovarian morphology in postmenarchal adolescents. *Fertil Steril* 2011;**95**:702–706.
- Duijkers IJ, Klipping C. Polycystic ovaries, as defined by the 2003 Rotterdam consensus criteria, are found to be very common in young healthy women. *Gynecol Endocrinol* 2010;**26**:152–160.
- Durlinger AL, Gruijters MJ, Kramer P, Karels B, Kumar TR, Matzuk MM, Rose UM, de Jong FH, Uilenbroek JT, Grootegoed JA et al. Anti-Müllerian hormone attenuates the effects of FSH on follicle development in the mouse ovary. *Endocrinology* 2001;**142**:4891–4899.
- Durlinger AL, Gruijters MJ, Kramer P, Karels B, Ingraham HA, Nachtigal MW, Uilenbroek JT, Grootegoed JA, Themmen AP. Anti-Müllerian hormone inhibits initiation of primordial follicle growth in the mouse ovary. *Endocrinology* 2002a;**143**:1076–1084.
- Durlinger AL, Visser JA, Themmen AP. Regulation of ovarian function: the role of anti-Müllerian hormone. *Reproduction* 2002b;**124**:601–609.
- Fruzzetti F, Perini D, Lazzarini V, Parrini D, Genazzani AR. Hyperandrogenemia influences the prevalence of the metabolic syndrome abnormalities in adolescents with the polycystic ovary syndrome. *Gynecol Endocrinol* 2009;**25**:335–343.
- Hagen CP, Aksglaede L, Sorensen K, Main KM, Boas M, Cleemann L, Holm K, Gravholt CH, Andersson A-M, Pedersen AT et al. Serum levels of anti-Müllerian hormone as a marker of ovarian function in 926 healthy females from birth to adulthood and in 172 turner syndrome patients. *J Clin Endocrinol Metab* 2010; doi:10.1210/jc.2010-0930.
- Hart R, Doherty DA, Norman RJ, Franks S, Dickinson JE, Hickey M, Sloboda DM. Serum antimüllerian hormone (AMH) levels are elevated in adolescent girls with polycystic ovaries and the polycystic ovarian syndrome (PCOS). *Fertil Steril* 2010a;**94**:1118–1121.
- Hart R, Sloboda DM, Doherty DA, Norman RJ, Atkinson HC, Newnham JP, Dickinson JE, Hickey M. Circulating maternal testosterone concentrations at 18 weeks of gestation predict circulating levels of antimüllerian hormone in adolescence: a prospective cohort study. *Fertil Steril* 2010b;**94**:1544–1547.
- Hickey M, Sloboda DM, Atkinson HC, Doherty DA, Franks S, Norman RJ, Newnham JP, Hart R. The relationship between maternal and umbilical cord androgen levels and polycystic ovary syndrome in adolescence: a prospective cohort study. *J Clin Endocrinol Metab* 2009;**94**:3714–3720.
- Ibanez L, Lopez-Bermejo A, Callejo J, Torres A, Cabre S, Dunger D, de Zegher F. Polycystic ovaries in nonobese adolescents and young women with ovarian androgen excess: relation to prenatal growth. *J Clin Endocrinol Metab* 2008;**93**:196–199.
- Johnstone EB, Rosen MP, Neril R, Trevithick D, Sternfeld B, Murphy R, Addaun-Andersen C, McConnell D, Pera RR, Cedars MI. The polycystic ovary post-rotterdam: a common, age-dependent finding in ovulatory women without metabolic significance. *J Clin Endocrinol Metab* 2010;**95**:4965–4972.
- Jonard S, Dewailly D. The follicular excess in polycystic ovaries, due to intra-ovarian hyperandrogenism, may be the main culprit for the follicular arrest. *Hum Reprod Update* 2004;**10**:107–117.
- Knight P, Glister C. Potential local regulatory functions of inhibins, activins and follistatin in the ovary. *Reproduction* 2001;**121**:503–512.
- March WA, Moore VM, Willson KJ, Phillips DIW, Norman RJ, Davies MJ. The prevalence of polycystic ovary syndrome in a community sample assessed under contrasting diagnostic criteria. *Hum Reprod* 2010;**25**:544–551.
- Murphy MK, Hall JE, Adams JM, Lee H, Welt CK. Polycystic ovarian morphology in normal women does not predict the development of polycystic ovary syndrome. *J Clin Endocrinol Metab* 2006;**91**:3878–3884.
- Norman RJ, Hague WM, Masters SC, Wang XJ. Subjects with polycystic ovaries without hyperandrogenaemia exhibit similar disturbances in insulin and lipid profiles as those with polycystic ovary syndrome. *Hum Reprod* 1995;**10**:2258–2261.
- Nur MM, Newman IM, Siqueira LM. Glucose metabolism in overweight Hispanic adolescents with and without polycystic ovary syndrome. *Pediatrics* 2009;**124**:e496–e502.
- Pacini G, Mari A. Methods for clinical assessment of insulin sensitivity and [beta]-cell function. *Best Pract Res Clin Endocrinol Metab* 2003;**17**:305–322.
- Park AS, Lawson MA, Chuan SS, Oberfield SE, Hoeger KM, Witchel SF, Chang RJ. Serum anti-Müllerian hormone concentrations are elevated in oligomenorrheic girls without evidence of hyperandrogenism. *J Clin Endocrinol Metab* 2010a;**95**:1786–1792.
- Park HT, Cho GJ, Ahn KH, Shin JH, Kim YT, Hur JY, Kim SH, Lee KW, Kim T. Association of insulin resistance with anti-Müllerian hormone levels in women without polycystic ovary syndrome (PCOS). *Clin Endocrinol* 2010b;**72**:26–31.
- Pigny P, Merlen E, Robert Y, Cortet-Rudelli C, Decanter C, Jonard S, Dewailly D. Elevated serum level of anti-müllerian hormone in patients with polycystic ovary syndrome: relationship to the ovarian follicle excess and to the follicular arrest. *J Clin Endocrinol Metab* 2003;**88**:5957–5962.
- Pigny P, Jonard S, Robert Y, Dewailly D. Serum anti-Müllerian hormone as a surrogate for antral follicle count for definition of the polycystic ovary syndrome. *J Clin Endocrinol Metab* 2006;**91**:941–945.

- Polson DW, Adams J, Wadsworth J, Franks S. Polycystic ovaries—a common finding in normal women. *Lancet* 1988;**1**:870–872.
- Porter MB. Polycystic ovary syndrome: the controversy of diagnosis by ultrasound. *Semin Reprod Med* 2008;**26**:241–251.
- Rey RA, Codner E, Iniguez G, Bedecarras P, Trigo R, Okuma C, Gottlieb S, Bergada I, Campo SM, Cassorla FG. Low risk of impaired testicular Sertoli and Leydig cell functions in boys with isolated hypospadias. *J Clin Endocrinol Metab* 2005;**90**:6035–6040.
- Roberts SA. Variability in anti-Müllerian hormone levels: a comment on Sowers et al., Anti-Müllerian hormone and inhibin B variability during normal menstrual cycles. *Fertil Steril* 2010;**94**:1482–1486.
- Rosenkrantz MA, Wachs DS, Coffler MS, Malcom PJ, Donohue M, Chang RJ. Comparison of inhibin B and estradiol responses to intravenous FSH in women with polycystic ovary syndrome and normal women. *Hum Reprod* 2010;**25**:198–203.
- Soto N, Iniguez G, Lopez P, Larenas G, Mujica V, Rey RA, Codner E. Anti-Müllerian hormone and inhibin B levels as markers of premature ovarian aging and transition to menopause in type I diabetes mellitus. *Hum Reprod* 2009;**24**:2838–2844.
- The Rotterdam ESHRE/ASRM-sponsored PCOS Consensus Workshop Group. Revised 2003 consensus on diagnostic criteria and long-term health risks related to polycystic ovary syndrome. *Fertil Steril* 2004;**81**:19–25.
- van Hooff M, Voorhorst FJ, Kaptein MBH, Hirasing RA, Koppenaal C, Schoemaker J. Polycystic ovaries in adolescents and the relationship with menstrual cycle patterns, luteinizing hormone, androgens, and insulin. *Fertil Steril* 2000;**74**:49–58.
- Venturoli S, Porcu E, Fabbri R, Pluchinotta V, Ruggeri S, Macrelli S, Paradisi R, Flamigni C. Longitudinal change of sonographic ovarian aspects and endocrine parameters in irregular cycles of adolescence. *Pediatr Res* 1995;**38**:974–980.
- Villarroel C, Trejo L, Munoz A, Kohen P, Fuentes A, Devoto L. Assessment of diagnostic competence of plasmatic androgens on polycystic ovary syndrome based on receiver operator characteristic curves. *Gynecol Endocrinol* 2010;**26**:600–606.
- Youlton R, Valenzuela C. Growth patterns in height and weight in children aged 0 to 17 years and cranial circumference in children aged 0 to 2 years from medium-high and high socioeconomic status in Santiago. Comparison with growth in children from medium-low and low status in the Northern area of Santiago. *Rev Chil Pediatr* 1990 (Spec No. 1–22).