We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists

6,300 Open access books available 171,000

190M Downloads



Our authors are among the

TOP 1%





WEB OF SCIENCE

Selection of our books indexed in the Book Citation Index in Web of Science™ Core Collection (BKCI)

Interested in publishing with us? Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected. For more information visit www.intechopen.com



Chapter

Perspective Chapter: Technological Strategies to Increase Insect Consumption – Transformation of Commodities Meal and Oil into Food/Functional Ingredients

Valeria Villanueva, Yanelis Ruiz, Fabrizzio Valdés, Marcela Sepúlveda and Carolina Valenzuela

Abstract

Insects have been proposed as an alternative source of nutrients to conventional foods, mainly protein sources because they have excellent nutritional quality and are sustainable. However, there are multiple barriers to mass consumption of insects, primarily the rejection and neophobia they provoke in individuals from Western cultures. Several studies have indicated that the acceptance of insects as food ingredients could be improved "if insects did not look like insects." Therefore, the focus of current research is to transform commodity-type ingredients such as insect flour and oil through various technologies applied in the food industry such as protein concentration, encapsulation, hydrolysis, fermentation, deodorization, to develop food ingredients with better sensory and technological properties are better accepted by people as a part of their diet. Interestingly, some food ingredients obtained from insects also have functional properties that could increase interest in consumption. These aspects will be reviewed in this chapter for further consideration of insects as food ingredients of the future.

Keywords: insect, functional ingredients, sensorial properties, food, feed

1. Introduction

The size of the world population and its accelerated growth are the greatest threat to humanity in terms of sustainability. The world population is expected to increase to 9.8 billion people by 2050 [1], requiring a 70–100% increase in food production to feed the world. Population growth could soon outstrip food production [2, 3]. Among the foods produced to feed humans and animals, those of animal origin are recognized as the least sustainable. For example, the production of 1 kg of beef protein has a carbon footprint between 45 and 640 kg CO₂ equivalents and a land use of 37–2100 m² [4, 5]. Enteric-derived methane from ruminant livestock accounts

for 17–37% of the methane emitted to the atmosphere from human activities [6–8]. Ingredients of animal origin are the most complex to replace in animal and human diets in terms of nutritional needs because: i) They have high crude protein content (20–23% for meat and fish and 40–70% for animal meals); ii) have highly digestible amino acids (close to 85–90% for meals and even higher for meat) [9–13]; iii) have a high content of essential amino acids [14, 15], iv) have highly bioavailable organic minerals, such as heme iron and zinc [14, 16], and v) have a high concentration of vitamins. Vitamin B12 is only found in foods of animal origin [17]. Several of these characteristics are not present in plant sources [18–21]. In addition, projections indicate that the price of animal-derived meat and meals will increase steadily [22].

For these reasons, there is an urgent search for new sustainable and moderate-cost protein ingredients with nutritional properties similar to those from an animal origin. Among the available alternatives are protein ingredients obtained from non-conventional raw vegetable materials such as chickpeas, lentils, beans, peas, broad beans, and others [23–25]. However, they do not always meet the demanding amino acid requirements (in terms of digestibility and essential amino acid supply) of animals and humans [26, 27]. Other alternatives are the development of protein ingredients from microalgae, algae, yeasts, fungi, microorganisms, and the re-processing of animal or marine waste [28–33]. The drawback of these alternatives is their low productive volume, which is extremely variable, and their high cost. Technological strategies have also been applied to protein ingredients, such as fermentation [34] and hydrolysis [35, 36], which increase protein digestibility, but do not modify the amino acid profile [26, 37].

The Food and Agriculture Organization of the United Nations (FAO) has proposed insects as food ingredients of the future to feed humans and animals [38]. Their use is based on the fact that insects have similar nutritional characteristics to ingredients of animal origin, in terms of protein contribution, amino acid profile, amino acid digestibility, and the presence of minerals and vitamins [39–44]. The most widely used insects worldwide for the development of food ingredients for humans and animals are black soldier fly larvae (BSFL, Hermetia illucens), mealworm larvae (ML, Tenebrio molitor), and adult house crickets (Acheta domesticus) [45-48], because they are produced industrially in mini-farms. From these, basic ingredients, commodities such as whole meal, defatted meal, and insect oil are obtained using simple technologies commonly used in the food industry [48–50]. However, there is great potential for obtaining other food ingredients from insects, with better sensory, and technological and even functional properties that have only been scarcely studied and have few existing industrial applications. The objective of this chapter is to analyze the potential of transforming insect flour and oil commodities into food/functional ingredients with improved sensory, technological and functional properties for massification and use as food ingredients for the future.

2. Development

2.1 Sensory barriers to the massification of insect consumption

There are several insect-based foods on the market such as cereal bars, drinks, pastas, candies, snacks, hamburgers, for human consumption [51, 52] and made with different concentrations of insect flour and oil [53–55]. The main problem lies with the acceptance of this type of food by people who are not familiar with entomophagy

(insect consumption), such as people from Western cultures, who often feel disgust, perceive insects as unpleasant, and reject their consumption [56]. In several survey-type studies, it was found that insect consumption could be better accepted if "insects did not look like insects" [57–59].

People who would be willing to consume insects describe some unpleasant sensory characteristics, such as: i) unpleasant odors [60, 61], identified as smelling of fungi, algae, fishy, and earthy [62–64], ii) unpleasant tastes of fish, fungi, bitterness [65, 66], iii) dark brown color of flour causing rejection [67], iv) grainy and rough texture of flour and whole insects [62, 63], and soft and oily texture in larvae [68] (Figure 1A and B), and v) unpalatable appearance of whole insects (Figure 1C) and meals, such as BSFL meal (Figure 1E) [69]. Additionally, processing to convert whole larvae/insects into meal (Figure 1D) can worsen these sensory perceptions, as Maillard reactions occur during the thermal process [70]. These reactions alter the color, odor, and flavor of insect flours in a negative way [71] and reduce the availability of some nutrients such as vitamin B12, potassium, phosphorus, sodium, some amino acids such as lysine [66]. There is also generation of unpleasant volatiles such as aldehydes, ketones, alcohols, esters, hydrocarbons, sulfur compounds and phenols, which generate unpleasant aromas [65, 72]. Thermal processing also generates darkening; for example, fly and mealworm larvae are cream-colored with yellow and orange shades (Figure 1A) and the meal obtained from these is dark brown (Figure 1E), due to the generation of brown and black coloring pigments such as melanoidins [66, 73]. The chitinous exoskeleton of insects is resistant to crushing [74]; therefore, the flour obtained after the milling process has granular texture, due to the large particle size (1.0–1.4 mm) of insect flours [53, 75], compared with flours of plant origin for example, wheat flour, which has a small particle size of about 100–150 µm [76].

The color of insect oil varies in yellow shades, and their melting point is variable depending on the profile and fatty acid content. For example, oil from BSFL contains lauric acid as the main fatty acid (21–29% of the total fatty acids, depending on the larval diet) [77]. Lauric acid is a saturated fatty acid, which gives the oil a high melting temperature ($\approx 43^{\circ}$ C). The oil is solid at room temperature, limiting its use as a food ingredient and making the incorporation into feed and/or diet formulations complex [78]. During oil processing, negative sensory changes also occur, mainly in oils with higher polyunsaturated fatty acid content, which tend to oxidation, producing odors and flavors described as "rancid and unpleasant" [60, 61]. Crude oil contains various components such as gums, free fatty acids, aromatic residues, and pigments, which negatively affect flavor, nutritional value, appearance, and stability [79].



Figure 1.

Appearance of whole insects, A: BSFL, B: mealworm larvae, C: adult house crickets, D: processing of insects into food ingredients, such as flour (E) and oil (F).

The addition of whole or processed insects to a food negatively affects its sensory quality, even if added in small amounts (<5% for flour), because they contribute to characteristic flavors and aromas, considered unpalatable to people, affect the appearance and texture, and darken the product [54, 80–82]. Therefore, the addition of insect-based ingredients to foods remains a major challenge.

2.2 Common food ingredients from insects: meal and oil

The main insect-based ingredients produced in the world have been whole meal and defatted meal and oil, which are obtained by relatively simple processing and are widely used in the food industry [50]. The following processes are used to obtain flour: blanching, drying, grinding, and addition of additives [83, 84]. Blanching is the process where whole insects (larvae and/or adults) are placed in boiling water, and then removed and immersed in ice water to stop the thermal process. Blanching is used as a pretreatment to reduce the microbial load of bacteria and fungi and inactivate the degradative enzymes responsible for spoilage, but does not affect bacterial spores [85–88]. Blanching time can be from seconds to 16 minutes, with a 5-minute average, and this process can be repeated several times for differing periods of time. The ratio of insects/water used has been 1/10-1/12. The time of immersion in ice water is from 30 seconds to 5 minutes. Between the blanching and cooling processes in water, the insects can be drained and crushed. Sterilizing solutions of 5% NaCl can also be used in this process [50]. The second process the insects receive is drying to reduce total water content and water activity, decreasing degradation reactions, including enzymatic reactions and those produced by microorganisms [89, 90]. The drying methods used include air convention dryer, solar drying, oven drying, smoke drying, frying pan, freeze drying, microwave-assisted drying, fluidized bed drying, oven drying with air circulation, and ultrasound-assisted aqueous extraction. Of all these methods, the most widely used for the industrial production of insect meal is oven drying in conventional hot air drying, using temperature ranges between 40 and 80°C for 8 to 48 hours until the sample reaches constant weight [50]. The last process is milling, which mechanically reduces the whole insect to the consistency of powder or flour [91]. For grinding, the use of a roller mill [75], blade mill [92, 93], colloid mill [94], or mechanical disruptor [60] has been described with times varying between 2 and 10 minutes, depending on the method chosen [60, 93].

To obtain defatted meal, it is necessary to extract the oil. Oil extraction is commonly performed with organic solvents, such as hexane, ethanol, isopropanol, methanol, petroleum ether, acetone, diethyl ether, and their mixtures [94–102]. Solvent extraction techniques involve partitioning between two immiscible liquids, continuous extractions, or batch extraction of solids. This process consists of three stages: pretreatment, desolventization, and solvent refining [103]. Although extraction with organic solvents has been the most widely used for oil extraction, other methods recognized as "green" for being more innocuous have also been studied, such as extraction with supercritical CO₂ [104–106]. The latter is a promising process, with a good percentage of defatting; however, it is more expensive. High hydrostatic pressure extraction has also been investigated [107, 108]. Insect oil has been used in the formulation of human food [109, 110], salmonid diets [111, 112], complete feeds, and pet snacks [48].

The two most important nutritional components in insect meal are protein and fat. The ranges of crude protein and crude fat content of whole and defatted meal of the insects most commonly used in human food development and most consumed by animals are presented in **Table 1** [14, 94, 99, 113–120]. Of the three insects

analyzed, crickets have the highest protein content in the complete meal, followed by mealworm and BSFL. The insects with the least amount of fat content in the complete meal are mealworm followed by BSFL and then crickets. Complete meal has been widely used to formulate diets for productive animals, mainly in aquaculture [121–123], pet diets and snacks [48], and human food [44, 124]. Defatted meal has a significant increase in total protein content (20–23%) and a reduction in fat content [99]. Defatted meal has been used to develop new ingredients that concentrate insect protein (hydrolysates, isolates, protein concentrates) for humans [44], specialized pet foods, such as hypoallergenic foods [48], and bioactive extracts with potential nutraceutical use [125, 126].

The protein and fat content is variable for each insect, so **Table 1** presents ranges. The primary factors influencing fat content are intrinsic variability of each insect species, developmental stage (larvae, pupae or adults), the diets used to feed the insects during the rearing and fattening period, and environmental conditions [127, 128].

2.3 Transformation of insect meal and oil into new food/functional ingredients

For mass consumption of insects to become the food of the future, it is necessary to transform insects into food ingredients of greater acceptability for the human population, using various technologies used by the food industry [129]. For animals, this is not necessary as insect meals and oil have high acceptability by aquaculture species [130], productive animals (pigs, hens, and chickens) [131–133], and domestic pets such as dogs and exotic animals [48]. **Table 2** and **Figure 2** present the new food and/or functional ingredients based on insect oil (**Figure 2A**) and meal (**Figure 1B**) commodities developed for humans. The main ingredients used as a base have been whole meal, defatted meal, and oil [172, 173]. More insect meal-based ingredients have been developed than insect oil. The developed oil-based ingredients are refined and deodorized oils, with better sensory properties (better odor and lighter yellow color). Emulsion technology has been applied to change the physical appearance of some insect oils, primarily BSFL, which as indicated above is solid at room temperature (**Figure 2A**), making it difficult to use in

Ingredients	BSFL	Mealworm	Cricket
	AN SE		
Whole meal			
Protein (%)	40-43 [14]	48–57 [14]	58–69 [14]
Fat (%)	17–34 [14]	32–40 [14]	11–23 [14]
Defatted meal			
Protein (%)	46–60 [113–116]	62–71 [94, 117–119]	79–81 [99, 107]
Fat (%)	5–11 [113–116]	1–14 [94, 117–119]	1–5 [99, 107]
RSEL · Black soldier flui	larvae (Hermetia Illucens) mea	worm (T molitor) cricket	(A domesticus)

Table 1.

Protein and fat ranges in dry basis of whole and defatted meal, of common insects used in human and animal feed.

Insect	Technology	New ingredient	Properties
From insect oil			
BSFL[79, 134]	Oil purification	Refined oil	Reduction of oil viscosity, turbidity and density Oil with high oxidative stability and better quality
ML[110]	Oil deodorization	Deodorized oil	Improvement of the organoleptic properties of the oil, such as appearance, color, and odor
ML[135]	Oleogelation with waxes	Oleogel as solid fat replacer in cookies	The replacement with carnauba wax/insect oleogel showed a desirable cookie quality in terms of spreadability and texture properties
BSFL[136]	Homogenization	Fat emulsions	Emulsions showed twofold lower consistency compared to the lecithin solutions of the same concentration
BSFL[137, 138]	Pre-homogenization and ultrasonication	Nanoemulsions	Nanoemulsions with high value- added for several applications in food industry Applications as drug delivery vehicles
BSFL, BM[139]	Solvent extraction	Oils with antimicrobial activity	Antimicrobial activity against <i>B. subtilis</i> and <i>S. aureus</i>
BSFL[140–145]	Solvent extraction	Lauric acid	Reduction of <i>E. coli</i> , <i>Streptococcus</i> spp., <i>Yersinia enterocolitica</i> and <i>Enterobacteriaceae</i> Increased number of <i>Lactobacillus</i> and <i>Bifidobacteriu</i> . Higher concentrations of total volatile fatty acids. Reduction of cytokine (IL-10 and 6) Positive effect on the gut microbiota composition and intestinal morphology
From insect meal	991 L		
AD, BSFL, ML, SG, AM[92, 95, 101, 146]	Alkaline solubilization and isoelectric precipitation	Protein concentrates	Lighter colored powders (beige- brown) were obtained High protein content (62–85%) Better aroma and appearance
ML, AL[147, 148]	Alkaline extraction, acid precipitation and salting out procedures	Protein isolates	Higher protein content (65–87%). Similar total amino acid content Better technological properties
AD, ML[64, 149, 150]	Biological and enzymatic hydrolysis and high hydrostatic pressures	Protein hydrolysates	Enhanced flavor (by hydrolysis and Maillard reaction) Lower chitin content obtaining antimicrobial substances Good sensory properties

Insect	Technology	New ingredient	Properties
MB, SB[151]	Acid/alkaline/water extraction	Gelatin	Very good gelling ability
ML, ADI, PB[152]	Homogenization of larvae with pork fat and freezing	Emulsions as meat replacement	ML was the most suitable candidate for use as a meat replacement due to its physicochemical and rheological properties
ML[153]	Extrusion	Snack	Extrusion improved the digestibility of ML proteins and starch Snacks with 10% YLM meal showed good expansion properties and pore structure, obtaining acceptable textural qualities; at 20%, the snacks showed poor expansion properties due to the higher fat content
BSFL[154]	Extrusion	Pellets	Extrudates of pure insect meal had the lowest water absorption index and the highest water solubility index. Insect meal in the corn blends negatively affected the pasting properties of the extrudates
HFL[155]	Spray drying	Micro-powder	Better appearance, similar to meals of vegetable origin Reduced emission of volatile compounds. Smaller particle size (9 μm) than other insect meals (355–1400 μm) [53, 75] Low protein content (5.1% db)
HFL[156]	Ionic gelation	Beads	Black "caviar" looking beads, darkening the color of HFL meal. Low protein content (27%) compared with HFL meal (54%) High antioxidant capacity of 1235–6903 µmol TE/100 g
AD, ML[157–159]	Yeast/lactic acid fermentation	Fermented powder	Flavor was improved The intensity of indole, pyrazines, 1-octen-3-ol and 3-octanol, which have unpleasant odors, was reduced. Pleasant volatiles, such as ethyl acetate, isopentyl acetate and 2-butanone, were increased Lactic fermentation resulted in successful acidification and increased shelf-life and safety by the control of Enterobacteria and bacterial spores

Insect	Technology	New ingredient	Properties
AD, ML, TE, OC, PB[125, 160–163]	Ultrasound-assisted extraction Pressurized liquid extraction	Functional extract	All extracts exhibited antioxidant activity and showed lipase inhibitory activity Potent hemolytic activity and anticoagulation activities
BSFL[164-166]	Solvent extraction, RNA isolation, cDNA cloning, solid-phase extraction and reverse-phase chromatography	Antimicrobial peptides (AMP)	The highest levels of AMP were induced by larvae diets supplemented with protein or sunflower oil. AMPs demonstrated activities against a spectrum of bacteria AMP exhibited antibacterial activity against both <i>Escherichia</i> <i>coli</i> and methicillin-resistant <i>Staphylococcus aureus</i>

BSFL: black soldier fly larvae, HFL: house fly larvae, ML: mealworm larvae, ADL: Allomyrina dichotoma larvae, AD: Acheta domesticus, BM: Bombyx mori; SG: Schistocerca gregaria; AM: Apis mellifera; AL: Anastrepha ludens; ADI: Allomyrina dichotoma; PB: Protaetia brevitarsis; TE: Teleogryllus emma; OC: Oxya chinensis; MB: Melon bug; SB: Sorghum bug.

Table 2.

New food and/or functional ingredients developed from insect meal and oil.

the formulation of diets, since it is complex to homogenize with the other ingredients and tends to form aggregates when combined with ingredients in powder form. After the emulsion process, liquid formulations are obtained (**Figure 2Ai**), with a milky appearance (**Figure 2Ai-iv**), which could be converted to powder by spray drying, to facilitate their use as a food ingredient and increase shelf life. These emulsions have been proposed as value-added ingredients for the food industry and as potential nutrient and drug vehicles for the pharmaceutical industry (in the case of nanoemulsions). Some nanoemulsions retain a milky appearance, while others tend to be transparent (**Figure 2Av**). BSFL fat has a similar fatty acid composition as coconut and palm oil, making it one of the most promising alternative fat sources for the food industry, where these lipid sources are used in a large number of processed foods [136].

The antimicrobial capacity of BSFL oil has been studied and demonstrated in *in vitro* studies against Gram-positive and Gram-negative bacteria [139, 174]. The antimicrobial capacity of BSFL is due to its high concentration of lauric acid [14]. The mechanisms of lauric acid antimicrobial processes are still being studied, but three have been described: 1) destruction of the cell membrane of gram-positive bacteria and lipid-coated viruses by physicochemical processes, 2) interference with cellular processes, such as signal transduction and transcription, and 3) destabilization of cell membranes [175], through inhibition of the enzyme MurA [176]. Very few *in vivo* investigations have been performed in animals to study the antimicrobial property of BSFL. The inclusion of BSFL oil in the diet does not affect the microbiota, improves intestinal morphology, and increases beneficial microorganism populations [140–142].

BSFL oil has the ability to regulate blood cholesterol levels due to its lauric acid content. In an *in silico* study animals fed lauric acid had increased cholesterol metabolism due to reduced HMG-CoA enzyme activity [177]. BSFL oil may also affect markers and coagulation factors, inhibiting platelet aggregation, prolonging the activated partial thromboplastin time. In *ex vivo* and *in vivo* studies, the compounds extracted



Figure 2.

Appearance of traditional insect ingredients, BSFL oil (A) and BSFL meal (B) and new insect-based ingredients, such as emulsions (Ai-Aiv [167, 168]), nanoemulsions (Av, [137]), protein extracts (Bi, [169]), protein concentrates (Bii, [170]), alginate-insect meal beads (Biii, [156]), micro-powders (Biv, [155]), and aqueous extracts (Bv, [171]).

from three insects, *Protaetia brevitarsis* seulensis, *Tenebrio mollitor* and *Oxya chinensis* sinuosa, succeeded in reducing platelet aggregation and the rate and size of arterial and pulmonary thrombus formation in mice [160–162].

Studies on new ingredients based on whole and defatted meals have focused on concentrating protein by developing protein concentrates, protein isolates, protein hydrolysates, and protein fermentates, using methods such as alkaline extraction and isoelectric precipitation (**Table 2**). The development of protein concentrates and protein isolates is focused on because i) protein is one of the most expensive nutrients in human and animal diets; and projections indicate that the price of protein ingredients of animal and plant origin will increase steadily [22]. ii) Proteins of animal origin are complex to replace in human and animal diets and are not very sustainable [178, 179]. Insect proteins represent a sustainable replacement alternative to animal proteins [180, 181]. iii) The protein content of insect meals, especially defatted meal, is very high (**Table 1**) and similar to meals of vegetable

origin (40–55%), meat/bone meal (40–50%), and offal (40–60%) [182]. The protein content of defatted cricket, mealworm, and BSFL meal is similar to meals of marine origin such as fish meal (60–75%) [182]. iv) The protein quality of insects, in terms of essential amino acid content (good source of lysine, methionine, threonine, leucine, alanine, valine) and amino acid digestibility (80–93%), is excellent. v) Insect proteins tend to be high in glutamic acid, which is the main amino acid in BSFL and mealworm meal [48], and is related to umami taste, highly preferred by animals and humans [183, 184]. vi) Insect proteins have technological properties suitable for the processing of certain foods such as meat substitutes [185], jerky meat analog [186], extruded cereals [153], rusks [187], and pastas [188]. vii) Products that concentrate insect proteins as concentrates and isolates have better sensory properties than insect meals, such as lighter colors [65], better taste [64], better volatile profile [149], and better emulsifying and foaming properties [98, 102]. Insect protein concentrates and isolates have been used in human food and are commercially available. Some examples are Becrit® and Trillions®, under the concept of protein shakes; Isaac nutrition®, protein powder; AdalbaPro IPC®, protein concentrate; and AdalbaPro FTIP®, protein concentrate powder with fiber texture. Figure 2 shows that the main change in appearance of protein concentrates (Figure 2Bi-ii), made from insect meals (Figure 2B), is a lighter coloration of the brown shades of the meals.

Hydrolysates and fermentates have been developed for the purpose of reducing some antinutritional factors of insect meals such as chitin [149], improving organoleptic properties, increasing shelf life [189, 190], providing antioxidant properties [149, 191, 192], increased nutrient digestibility, production of antimicrobial substances, and health-promoting molecules [149]. Enzymatic hydrolysis using a variety of enzymes, such as alkalase, papain, peptidase, protease; and alcalase, papain, peptidase, protease; and alcalase, papain, peptidase, protease; and biological hydrolysis using yeasts (*Yarrowia lipolytica* and *Debaryomyces hansenii*) have been studied to obtain peptides with bioactive properties and to improve protein digestibility, mainly. Fermentation of insect meal with lactic acid bacteria or yeast (*Saccharomyces cerevisiae*) has been used to improve sensory properties, mainly by modifying the volatile profiles, decreasing indole, pyrazines, 1-octen-3-ol, and 3-octanol, and increasing propanol, ethanol, acetone and 2-butanone, reducing fecal, toasted, earthy, mushroom, and bitter taste [157, 158].

Other technologies applied to develop new ingredients have been extrusion to produce pellets and snacks based mainly on insect meal mixed with cereals. The main result has been the increase in the digestibility of some nutrients such as protein and starch. In these studies, it was indicated that the insect meal content used alters some important properties of the extruded products (**Table 2**). This technology has been widely used for the development of complete foods and snacks for dogs and cats [48]. Some examples are Eat Small Mindfulness®, Brit Care Immunity®, CircularPet®, Yora®, Insecta®, buggybigs®.

Encapsulation is a technology widely used in the food industry, especially spray drying [193], because it improves the sensory characteristics of the compounds to be encapsulated [194]. In addition, controlled release formulations can be developed to add the encapsulated compounds in complex foods such as yogurt, beverages, dairy, and others [195–197]. Although, this technology is widely used to improve the sensory properties of ingredients, it has not been thoroughly studied for encapsulating insect meal and oil. Among the existing works, spray drying has been used to develop insect meal micro-powders (**Figure 2Biv**), which presented better appearance and color (similar to wheat flour), better texture (with smaller particle size), and better aroma than unencapsulated house fly larvae meal (**Table 2**). However, the protein content of

the micro-powders was low at about 5.1 g per 100 g of powder, whereas the meal that gave rise to the micro-powders contained 54 g of protein per 100 g of flour. Even so, micro-powders are considered a "source of protein" according to the Codex Alimentarius [198]. The challenge for this technology is to concentrate on the nutrients, especially protein, from insect meals, being able to use previously described technologies such as protein isolates and concentrates. In another study, house fly meal was encapsulated by ionic gelation, obtaining alginate-insect meal beads with an appearance similar to black "caviar" (**Figure 2Biii**), with better aroma than the unencapsulated meal (**Table 2**). The application of this type of product in human food is complex, due to the rejection that its appearance could cause, but it is possible to incorporate it as food for exotic pets (such as water turtles, fish, ferrets, hedgehogs), which consume live and dehydrated insect larvae [48]. The pet industry has a high level of innovation in food products and consumption of innovative foods and snacks is on the rise [199, 200].

In **Table 2**, the development of oil nanoemulsions is described. The technique is considered an encapsulation process, since the oil is protected and separated from the water by a dynamic surfactant layer formed by emulsifying agents. This technique has been widely used in the food industry for the following reasons: 1) to improve the stability of some lipophilic active compounds such as vitamin D3 [201], carotenoids [202], and α -tocopherol [203], 2) to improve the absorption, bioavailability, and bioactivity of lipophilic bioactive compounds with low absorption such as curcumin [204] and astaxanthin [205], and 3) to provide the ability to release encapsulated actives in a controlled manner [206, 207].

The functional properties of insect-based food ingredients, such as antioxidant capacity, antimicrobial activity, inhibition of platelet aggregation, enzymatic inhibition, and antidiabetic potential, have been less studied than their nutritional properties as food ingredients [208, 209]. The literature shows that ingredients obtained from insects such as aqueous extracts [125], meals [191, 210, 211], and proteins and peptides [126, 212, 213], exhibit high antioxidant capacity, so they could have potential use in health disorders associated with oxidative stress [214]. The antioxidant capacity is due to the presence of phenolic compounds, proteins, peptides, chitin, fatty acids, and others [215, 216].

A great diversity of bioactive compounds has been isolated from insects, such as free fatty acids, amino acids, organic acids, carbohydrates, hydrocarbons, sterols, and others [125]. The methodologies for their extraction have been ultrasound-assisted extraction (UAE) and pressurized liquid extraction (PLE), using ethanol or a mixture of ethanol and water [125] (**Table 2**). The appearance of these extracts is presented in **Figure 2Bv**, observing different colorations that depend on several factors, such as concentration of the extracts and extraction technique. Extracts have anti-inflammatory, antimicrobial, antiangiogenic, antiproliferative, and antioxidant properties. The ability to inhibit the activity of certain enzymes has also been studied. In in vitro studies, extracts of A. domesticus and T. molitor were able to inhibit pancreatic lipase [125]. These extracts could have an application in the treatment and prevention of obesity [217]. Proteins from the insects such as B. mori, T. molitor, Alphitobius diaperinus, and Gryllus bimaculatus [218–221] were able to inhibit angiotensin-converting enzyme (ACE), dipeptidyl peptidase-4 (DPP-IV) [220], and α -glucosidase activity [218, 221], with antidiabetic potential. In animal studies, supplementation with ethanolic extract of B. mori improved glycemic status in obese mice with type 2 diabetes, reducing glycemia and restoring pancreatic functionality [222, 223]. Soluble extracts obtained from 12 insect species showed antioxidant activity, the highest in extracts from grasshoppers, silkworms, and crickets [216]. Antioxidant activity was also found in aqueous extract of Vespa affinis L. [214]. Antibacterial substances

such as N-beta-alanyl-5-S-glutathionyl-5-S-glutathionyl-3,4-dihydroxyphenylalanine from *Sarcophaga peregrina* and p-hydroxycinnamaldehyde from *Acantholyda parki* larvae were isolated from extracts of immunized insects [224, 225].

Other insect-based functional compounds extensively studied in the recent years are antimicrobial peptides (AMPs), which are extracted and purified by different technologies, such as reverse phase high-performance liquid chromatography (RP-HPLC), DNA extraction, RNA extraction, fast performance liquid chromatography (FPLC), and gel filtration chromatography [226] (**Table 2**). AMPs are peptides with low molecular weight, high thermal stability, and a broad antimicrobial spectrum [227, 228]. A large number of AMPs derived from *Acalolepta luxuriosa*, *A. mellifera*, *B. mori*, *Galleria mellonella*, *Heterometrus spinifer*, *Holotrichia diomphalia*, *Hyalophora cecropia*, *Oxysternon conspicillatum*, *Pandinus imperator*, and *Sarcophaga peregrine* have been investigated and are effective against a wide range of Gram-negative and Gram-positive bacteria [229, 230]. Their mechanism of action depends on the type of AMP and the target pathogen. AMPs can interact with the microbial membrane surface, alter permeability and induce cell lysis, enter the cell, and damage bacterial components such as DNA and RNA, and promote the bacteriostatic effects [226, 228]. The use of AMPs as an alternative to antimicrobials in human and animal health could help reduce antimicrobial resistance [228].

The challenge for the future of insects as food for humans and animals is to increase research on technologies that can be used to transform common insect-based ingredients such as meal and oil into ingredients with higher added value, functional properties, and optimal sensory properties for greater acceptance and consumption of insects by humans, so these ingredients can be included in a greater number of foods.

Acknowledgements

The authors acknowledge the support of FONDEF IDea I + D ID22I10030 and the valuable help of Susan Cleveland, who proofread the chapter.

Conflict of interest

The authors declare no conflict of interest.

Author details

Valeria Villanueva, Yanelis Ruiz, Fabrizzio Valdés, Marcela Sepúlveda and Carolina Valenzuela^{*} Facultad de Ciencias Veterinarias y Pecuarias, Universidad de Chile, Santiago, Chile

*Address all correspondence to: cvalenzuelav@u.uchile.cl

IntechOpen

© 2022 The Author(s). Licensee IntechOpen. This chapter is distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/3.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

References

[1] Food and Agriculture Organization of the United Nations (FAO). FAOSTAT Food and agriculture data [Internet].
2022. Available from: https://www.fao. org/faostat/es/#home. [Accessed: August 4, 2022]

[2] Bruinsma J. The resource outlook to 2050: by how much do land, water and crop yields need to increase by 2050? [Internet]. 2009. Available from: https://www.fao.org/publications/ card/es/c/f70d2098-c9ad-5b05-8972cf987e74d681/. [Accessed: August 4, 2022]

[3] Myers SS, Smith MR, Guth S, Golden CD, Vaitla B, Mueller ND, et al. Climate change and global food systems: Potential impacts on food security and undernutrition. Annual Review of Public Health. 2017;**38**(1):259-277. DOI: 10.1146/ annurev-publhealth-031816-044356

[4] Nijdam D, Rood T, Westhoek H. The price of protein: Review of land use and carbon footprints from life cycle assessments of animal food products and their substitutes. Food Policy. 2012;**37**(6):760-770. DOI: 10.1016/j. foodpol.2012.08.002

[5] Röös E, Ekelund L, Tjärnemo H. Communicating the environmental impact of meat production: Challenges in the development of a Swedish meat guide. Journal of Cleaner Production. 2014;**73**(6):154-164. DOI: 10.1016/j. jclepro.2013.10.037

[6] Steinfeld H, Wassenaar T. The role of livestock production in carbon and nitrogen cycles. Annual Review of Environment and Resources. 2007;**32**(1):271-294. DOI: 10.1146/ annurev.energy.32.041806.143508 [7] Lassey KR. Livestock methane emission and its perspective in the global methane cycle. Australian Journal of Experimental Agriculture. 2008;**48**(2):114-118. DOI: 10.1071/ EA07220

[8] Cai Y, Tang R, Tian L, Chang SX. Environmental impacts of livestock excreta under increasing livestock production and management considerations: Implications for developing countries. Current Opinion in Environmental Science & Health. 2021;24(3):100300. DOI: 10.1016/j. coesh.2021.100300

[9] Murray SM, Patil AR, Fahey GC, Merchen NR, Hughes DM. Raw and rendered animal by-products as ingredients in dog diets. Journal of Animal Science. 1997;75(9):2497-2505. DOI: 10.2527/1997.7592497x

[10] Johnson ML, Parsons CM, Fahey GC, Merchen NR, Aldrich CG. Effects of species raw material source, ash content, and processing temperature on amino acid digestibility of animal by-product meals by cecectomized roosters and ileally cannulated dogs. Journal of Animal Science. 1998;**76**(4):1112-1122. DOI: 10.2527/ 1998.7641112x

[11] Wang X, Parsons CM. Effect of raw material source, processing systems, and processing temperatures on amino acid digestibility of meat and bone meals. Poultry Science. 1998;77(6):834-841. DOI: 10.1093/ps/77.6.834

[12] Shirley RB, Parsons CM. Effect of ash content on protein quality of meat and bone meal. Poultry Science.
2001;80(5):626-632. DOI: 10.1093/ ps/80.5.626 [13] Dozier WA, Dale NM, Dove CR.
Nutrient composition of feed-grade and pet-food-grade poultry by-product meal.
Journal of Applied Poultry Research.
2003;12(4):526-530. DOI: 10.1093/
japr/12.4.526

[14] Makkar HPS, Tran G, Heuzé V, Ankers P. State-of-the-art on use of insects as animal feed. Animal Feed Science and Technology.
2014;197(11):1-33. DOI: 10.1016/j. anifeedsci.2014.07.008

[15] Bosch G, Zhang S, Oonincx DGAB, Hendriks WH. Protein quality of insects as potential ingredients for dog and cat foods. Journal of Nutritional Science. 2014;**3**(3):e29. DOI: 10.1017/jns.2014.23

[16] Ordoñez-Araque R, Egas-Montenegro E. Edible insects: A food alternative for the sustainable development of the planet. International Journal of Gastronomy and Food Science. 2021;**23**(6):100304. DOI: 10.1016/j. ijgfs.2021.100304

[17] Schmidt A, Call L-M, Macheiner L, Mayer HK. Determination of vitamin B12 in four edible insect species by immunoaffinity and ultra-high performance liquid chromatography. Food Chemistry. 2019;**281**(3):124-129. DOI: 10.1016/j.foodchem.2018.12.039

[18] Bednar GE, Murray SM, Patil AR,
Flickinger EA, Merchen NR, Fahey GC.
Selected animal and plant protein sources affect nutrient digestibility and fecal characteristics of ileally cannulated dogs. Arch für Tierernaehrung.
2000;53(2):127-140. DOI: 10.1080/
17450390009381942

[19] Donadelli RA, Jones CK, Beyer RS. The amino acid composition and protein quality of various egg, poultry meal by-products, and vegetable proteins used in the production of dog and cat diets. Poultry Science. 2019;**98**(3):1371-1378. DOI: 10.3382/ps/pey462

[20] Félix AP, Menezes Souza CM, Bastos TS, Kaelle GCB, Oliveira SG, Maiorka A. Digestibility of raw soybeans in extruded diets for dogs determined by different methods. Italian Journal of Animal Science. 2020;**19**(1):95-102. DOI: 10.1080/1828051X.2019.1698324

[21] Acuff HL, Dainton AN, Dhakal J, Kiprotich S, Aldrich G. Sustainability and pet food: Is there a role for veterinarians? The Veterinary Clinics of North America. Small Animal Practice. 2021;**51**(3):563-581. DOI: 10.1016/j.cvsm.2021.01.010

[22] OECD/FAO. OECD-FAO
Agricultural outlook (Edition
2021), OECD Agriculture Statistics
(database) [Internet]. 2021. DOI:
10.1787/4bde2d83-en. [Accessed: March
26, 2022]

[23] Øverland M, Sørensen M, Storebakken T, Penn M, Krogdahl Å, Skrede A. Pea protein concentrate substituting fish meal or soybean meal in diets for Atlantic salmon (Salmo salar)— Effect on growth performance, nutrient digestibility, carcass composition, gut health, and physical feed quality. Aquaculture. 2009;**288**(3-4):305-311. DOI: 10.1016/j.aquaculture.2008.12.012

[24] Freeman LM, Stern JA, Fries R, Adin DB, Rush JE. Diet-associated dilated cardiomyopathy in dogs: What do we know? Journal of the American Veterinary Medical Association. 2018;**253**(11):1390-1394. DOI: 10.2460/ javma.253.11.1390

[25] ReillyLM,vonSchaumburgPC,HokeJM, DavenportGM,UtterbackPL,ParsonsCM, et al. Macronutrient composition, true metabolizable energy and amino acid digestibility, and indispensable amino acid scoring of pulse ingredients for

use in canine and feline diets. Journal of Animal Science. 2020;**98**(6):skaa149. DOI: 10.1093/jas/skaa149

[26] Dodd SAS, Adolphe JL,
Verbrugghe A. Plant-based diets
for dogs. Journal of the American
Veterinary Medical Association.
2018;253(11):1425-1432. DOI: 10.2460/
javma.253.11.1425

[27] Dodd SAS, Shoveller AK, Fascetti AJ, Yu ZZ, Ma DWL, Verbrugghe A. A comparison of key essential nutrients in commercial plant-based pet foods sold in Canada to American and European canine and feline dietary recommendations. Animals. 2021;**11**(8):2348. DOI: 10.3390/ ani11082348

[28] Brain-Isasi S, Carú C, Lienqueo ME. Valorization of the green seaweed *Ulva rigida* for production of fungal biomass protein using a hypercellulolytic terrestrial fungus. Algal Research. 2021;**59**(3):102457. DOI: 10.1016/j. algal.2021.102457

[29] Dale N, Valenzuela C. Nutritional properties of dried salmon silage for broiler feeding. Animal Science Journal. 2016;**87**(6):791-795. DOI: 10.1111/ asj.12480

[30] Luigia DS. New protein sources: Novel foods. In: Ferranti P, Berry EM, Anderson JR, editors. Encyclopedia of Food Security and Sustainability. Amsterdam: Elsevier; 2019. pp. 276-279. DOI: 10.1016/B978-0-08-100596-5. 22129-X

[31] Fasolin LH, Pereira RN, Pinheiro AC, Martins JT, Andrade CCP, Ramos OL, et al. Emergent food proteins – Towards sustainability, health and innovation. Food Research International. 2019;**125**(5):108586. DOI: 10.1016/j. foodres.2019.108586 [32] Food and Agriculture Organization of the United Nations (FAO). The state of world fisheries and aquaculture sustainability in action [Internet]. 2020. Available from: https://www.fao.org/3/ ca9229en/ca9229en.pdf. [Accessed: November 16, 2021]

[33] Mosna D, Bottani E, Vignali G, Montanari R. Environmental benefits of pet food obtained as a result of the valorisation of meat fraction derived from packaged food waste. Waste Management. 2021;**125**(3):132-144. DOI: 10.1016/j.wasman.2021.02.035

[34] Thirunathan P, Manickavasagan A.
Processing methods for reducing alpha-galactosides in pulses. Critical Reviews in Food Science and Nutrition.
2019;59(20):3334-3348. DOI: 10.1080/
10408398.2018.1490886

[35] Tavano OL. Protein hydrolysis using proteases: An important tool for food biotechnology. Journal of Molecular Catalysis B: Enzymatic. 2013;**90**(1):1-11. DOI: 10.1016/j.molcatb.2013.01.011

[36] Marciniak A, Suwal S, Naderi N, Pouliot Y, Doyen A. Enhancing enzymatic hydrolysis of food proteins and production of bioactive peptides using high hydrostatic pressure technology. Trends in Food Science and Technology. 2018;**80**(11):187-198. DOI: 10.1016/j.tifs.2018.08.013

[37] Kanakubo K, Fascetti AJ, Larsen JA. Assessment of protein and amino acid concentrations and labeling adequacy of commercial vegetarian diets formulated for dogs and cats. Journal of the American Veterinary Medical Association. 2015;**247**(4):385-392. DOI: 10.2460/javma.247.4.385

[38] Food and Agriculture Organization of the United Nations (FAO). Edible Insects. Future Prospects for Food and Feed Security. Rome, Italy: Wageningen UR; 2013. p. 201

[39] Rumpold BA, Schlüter OK. Nutritional composition and safety aspects of edible insects. Molecular Nutrition & Food Research. 2013;57(5):802-823. DOI: 10.1002/ mnfr.201200735

[40] Belluco S, Losasso C, Maggioletti M, Alonzi CC, Paoletti MG, Ricci A. Edible insects in a food safety and nutritional perspective: A critical review. Comprehensive Reviews in Food Science and Food Safety. 2013;**12**(3):296-313. DOI: 10.1111/1541-4337.12014

[41] Zielińska E, Baraniak B, Karaś M, Rybczyńska K, Jakubczyk A. Selected species of edible insects as a source of nutrient composition. Food Research International. 2015;77:460-466. DOI: 10.1016/j.foodres.2015.09.008

[42] Williams JP, Williams JR, Kirabo A, Chester D, Peterson M. Chapter 3 -Nutrient content and health benefits of insects. In: Dossey AT, Morales-Ramos JA, Rojas G, editors. Insects as Sustainable Food Ingredients. USA: Academic Press; 2016. pp. 61-84. DOI: 10.1016/B978-0-12-802856-8/ 00003-X

[43] Mishyna M, Glumac M. So different, yet so alike Pancrustacea: Health benefits of insects and shrimps. Journal of Functional Foods. 2021;**76**:104316. DOI: 10.1016/j.jff.2020.104316

[44] Borges MM, da Costa DV, Trombete FM, Câmara AKFI. Edible insects as a sustainable alternative to food products: An insight into quality aspects of reformulated bakery and meat products. Current Opinion in Food Science. 2022;**46**(3):100864. DOI: 10.1016/j.cofs.2022.100864 [45] van Huis A. Environmental sustainability of insects as human food. In: Reference Module in Food Science. Ámsterdam: Elsevier; 2019. pp. 1-5. DOI: 10.1016/ B978-0-08-100596-5.22589-4

[46] van Huis A. Insects as food and feed, a new emerging agricultural sector: A review. Journal of Insects as Food and Feed. 2020;**6**(1):27-44. DOI: 10.3920/ JIFF2019.0017

[47] van Huis A. Nutrition and health of edible insects. Current Opinion in Clinical Nutrition and Metabolic Care. 2020;**23**(3):228-231. DOI: 10.1097/ MCO.000000000000641

[48] Valdés F, Villanueva V, Durán E, Campos F, Avendaño C, Sánchez M, et al. Insects as feed for companion and exotic pets: A current trend. Animals. 2022;**12**(11):1450. DOI: 10.3390/ ani12111450

[49] Tzompa-Sosa DA, Yi L, van Valenberg HJF, Lakemond CMM. Four insect oils as food ingredient: Physical and chemical characterisation of insect oils obtained by an aqueous oil extraction. Journal of Insects as Food and Feed. 2019;5(4):279-292. DOI: 10.3920/ JIFF2018.0020

[50] Melgar-Lalanne G, Hernández-Álvarez A, Salinas-Castro A. Edible insects processing: Traditional and innovative technologies. Comprehensive Reviews in Food Science and Food Safety. 2019;**18**(4):1166-1191. DOI: 10.1111/ 1541-4337.12463

[51] Caparros Megido R, Gierts C, Blecker C, Brostaux Y, Haubruge É, Alabi T, et al. Consumer acceptance of insect-based alternative meat products in Western countries. Food Quality and Preference. 2016;**52**(3):237-243. DOI: 10.1016/j.foodqual.2016.05.004

[52] Motoki K, Ishikawa S, Spence C, Velasco C. Contextual acceptance of insect-based foods. Food Quality and Preference. 2020;**85**(3):103982. DOI: 10.1016/j.foodqual.2020.103982

[53] de Oliveira LM, da Silva Lucas AJ, Cadaval CL, Mellado MS. Bread enriched with flour from cinereous cockroach (*Nauphoeta cinerea*). Innovative Food Science and Emerging Technologies. 2017;44(6):30-35. DOI: 10.1016/j. ifset.2017.08.015

[54] Osimani A, Milanović V, Cardinali F, Roncolini A, Garofalo C, Clementi F, et al. Bread enriched with cricket powder (*Acheta domesticus*): A technological, microbiological and nutritional evaluation. Innovative Food Science and Emerging Technologies. 2018;**48**(6):150-163. DOI: 10.1016/j.ifset.2018.06.007

[55] Cicatiello C, Vitali A, Lacetera N. How does it taste? Appreciation of insectbased snacks and its determinants. International Journal of Gastronomy and Food Science. 2020;**21**(5):100211. DOI: 10.1016/j.ijgfs.2020.100211

[56] Jensen NH, Lieberoth A. We will eat disgusting foods together – Evidence of the normative basis of Western entomophagy-disgust from an insect tasting. Food Quality and Preference. 2019;**72**(4):109-115. DOI: 10.1016/j. foodqual.2018.08.012

[57] Orsi L, Voege LL, Stranieri S. Eating edible insects as sustainable food? Exploring the determinants of consumer acceptance in Germany. Food Research International. 2019;**125**:108573. DOI: 10.1016/j.foodres.2019.108573

[58] Tuccillo F, Marino MG, Torri L. Italian consumers' attitudes towards entomophagy: Influence of human factors and properties of insects and insect-based food. Food Research International. 2020;**137**(20):109619. DOI: 10.1016/j.foodres.2020.109619

[59] Higa JE, Ruby MB, Rozin P. Americans' acceptance of black soldier fly larvae as food for themselves, their dogs, and farmed animals. Food Quality and Preference. 2021;**90**(4):104119. DOI: 10.1016/j.foodqual.2020.104119

[60] Larouche J, Deschamps M-H, Saucier L, Lebeuf Y, Doyen A, Vandenberg GW. Effects of killing methods on lipid oxidation, colour and microbial load of black soldier fly (*Hermetia illucens*) larvae. Animals. 2019;**9**(4):182. DOI: 10.3390/ani9040182

[61] Singh Y, Cullere M, Kovitvadhi A, Chundang P, Dalle ZA. Effect of different killing methods on physicochemical traits, nutritional characteristics, in vitro human digestibility and oxidative stability during storage of the house cricket (*Acheta domesticus* L.). Innovative Food Science and Emerging Technologies. 2020;**65**(5):102444. DOI: 10.1016/j.ifset.2020.102444

[62] Dion-Poulin A, Turcotte M, Lee-Blouin S, Perreault V, Provencher V, Doyen A, et al. Acceptability of insect ingredients by innovative student chefs: An exploratory study. International Journal of Gastronomy and Food Science. 2021;**24**(6):100362. DOI: 10.1016/j. ijgfs.2021.100362

[63] Sogari G, Menozzi D, Mora C. Sensory-liking expectations and perceptions of processed and unprocessed insect products. International Journal of Food System Dynamics. 2018;**9**(4):314-320. DOI: 10.18461/ijfsd.v9i4.942

[64] Grossmann KK, Merz M, Appel D, De Araujo MM, Fischer L. New insights into the flavoring potential of cricket (*Acheta domesticus*) and mealworm (*Tenebrio molitor*) protein hydrolysates and their Maillard products. Food Chemistry. 2021;**364**(3):130336. DOI: 10.1016/j.foodchem.2021.130336

[65] Lee JH, Cha J-Y, Kim T-K, Choi Y-S, Jang HW. Effects of a defatting process on the thermal stabilities and volatile compound profiles of proteins isolated from *Protaetia brevitarsis* larvae. LWT. 2021;**151**(1):112095. DOI: 10.1016/j. lwt.2021.112095

[66] Ssepuuya G, Nakimbugwe D, De Winne A, Smets R, Claes J, Van Der Borght M. Effect of heat processing on the nutrient composition, colour, and volatile odour compounds of the longhorned grasshopper *Ruspolia differens* serville. Food Research International. 2020;**129**(4):108831. DOI: 10.1016/j. foodres.2019.108831

[67] Kim H-W, Setyabrata D, Lee YJ, Jones OG, Kim YHB. Pre-treated mealworm larvae and silkworm pupae as a novel protein ingredient in emulsion sausages. Innovative Food Science and Emerging Technologies. 2016;**38**(5):116-123. DOI: 10.1016/j.ifset.2016.09.023

[68] Wang Y-S, Shelomi M. Review of black soldier fly (*Hermetia illucens*) as animal feed and human food. Food. 2017;**6**(10):91. DOI: 10.3390/ foods6100091

[69] Mishyna M, Chen J, Benjamin O. Sensory attributes of edible insects and insect-based foods – Future outlooks for enhancing consumer appeal. Trends in Food Science and Technology. 2020;**95**(4):141-148. DOI: 10.1016/j. tifs.2019.11.016

[70] David-Birman T, Raften G, Lesmes U. Effects of thermal treatments on the colloidal properties, antioxidant capacity and *in-vitro* proteolytic degradation of cricket flour. Food Hydrocolloids. 2018;**79**(6):48-54. DOI: 10.1016/j. foodhyd.2017.11.044

[71] Mishyna M, Haber M, Benjamin O, Martinez JJI, Chen J. Drying methods differentially alter volatile profiles of edible locusts and silkworms. Journal of Insects as Food and Feed. 2020;**6**(4):405-415. DOI: 10.3920/JIFF2019.0046

[72] Khatun H, Claes J, Smets R, De Winne A, Akhtaruzzaman M, Van Der Borght M. Characterization of freezedried, oven-dried and blanched house crickets (*Acheta domesticus*) and Jamaican field crickets (*Gryllus assimilis*) by means of their physicochemical properties and volatile compounds. European Food Research and Technology. 2021;**247**(5):1291-1305. DOI: 10.1007/ s00217-021-03709-x

[73] Kouřimská L, Adámková A. Nutritional and sensory quality of edible insects. NFS Journal. 2016;**4**(5):22-26. DOI: 10.1016/j.nfs.2016.07.001

[74] Bassett FS, Dunn ML, Pike OA, Jefferies LK. Physical, nutritional, and sensory properties of spray-dried and oven-roasted cricket (*Acheta domesticus*) powders. Journal of Insects as Food and Feed. 2021;7(6):987-1000. DOI: 10.3920/ JIFF2020.0107

[75] Purschke B, Brüggen H, Scheibelberger R, Jäger H. Effect of pre-treatment and drying method on physico-chemical properties and dry fractionation behaviour of mealworm larvae (*Tenebrio molitor* L.). European Food Research and Technology. 2018;**244**(2):269-280. DOI: 10.1007/ s00217-017-2953-8

[76] Liu T, Hou GG, Lee B, Marquart L, Dubat A. Effects of particle size on the quality attributes of reconstituted whole-wheat flour and tortillas made from it. Journal of Cereal Science.

2016;**71**(4):145-152. DOI: 10.1016/j. jcs.2016.08.013

[77] Matsue M, Mori Y, Nagase S, Sugiyama Y, Hirano R, Ogai K, et al. Measuring the antimicrobial activity of lauric acid against various bacteria in human gut microbiota using a new method. Cell Transplantation. 2019;**28**(12):1528-1541. DOI: 10.1177/ 0963689719881366

[78] Zeng C, Liu Y, Ding Z, Xia H, Guo S. Physicochemical properties and antibacterial activity of hydrophobic deep eutectic solvent-in-water nanoemulsion. Journal of Molecular Liquids. 2021;**338**(11):116950. DOI: 10.1016/j.molliq.2021.116950

[79] Mai HC, Dao ND, Lam TD, Nguyen BV, Nguyen DC, Bach LG. Purification process, physicochemical properties, and fatty acid composition of black soldier fly (*Hermetia illucens* Linnaeus) larvae oil. Journal of the American Oil Chemists' Society. 2019;**96**(11):1303-1311. DOI: 10.1002/ aocs.12263

[80] Biró B, Fodor R, Szedljak I, Pásztor-Huszár K, Gere A. Buckwheatpasta enriched with silkworm powder: Technological analysis and sensory evaluation. LWT. 2019;**116**(3):108542. DOI: 10.1016/j.lwt.2019.108542

[81] Choi Y-S, Kim T-K, Choi H-D, Park J-D, Sung J-M, Jeon K-H, et al. Optimization of replacing pork meat with yellow worm (*Tenebrio molitor* L.) for frankfurters. Korean Journal for Food Science of Animal Resources. 2017;**37**(5):617-625. DOI: 10.5851/ kosfa.2017.37.5.617

[82] Haber M, Mishyna M, Martinez JJI, Benjamin O. The influence of grasshopper (*Schistocerca gregaria*) powder enrichment on bread nutritional and sensorial properties. LWT. 2019;**115**(3):108395. DOI: 10.1016/j. lwt.2019.108395

[83] Mishyna M, Keppler JK, Chen J. Techno-functional properties of edible insect proteins and effects of processing. Current Opinion in Colloid & Interface Science. 2021;**56**:101508. DOI: 10.1016/j. cocis.2021.101508

[84] Meshulam-Pascoviche D, David-Birman T, Refael G, Lesmes U. Big opportunities for tiny bugs: Processing effects on the techno-functionality and digestibility of edible insects. Trends in Food Science and Technology. 2022;**122**(4):265-274. DOI: 10.1016/j. tifs.2022.02.012

[85] Vandeweyer D, Lenaerts S, Callens A, Van Campenhout L. Effect of blanching followed by refrigerated storage or industrial microwave drying on the microbial load of yellow mealworm larvae (*Tenebrio molitor*). Food Control. 2017;**71**:311-314. DOI: 10.1016/j. foodcont.2016.07.011

[86] Wynants E, Crauwels S, Verreth C, Gianotten N, Lievens B, Claes J, et al.
Microbial dynamics during production of lesser mealworms (*Alphitobius diaperinus*) for human consumption at industrial scale. Food Microbiology. 2018;**70**:181-191. DOI: 10.1016/j.fm. 2017.09.012

[87] Xiao H-W, Bai J-W, Sun D-W, Gao Z-J. The application of superheated steam impingement blanching (SSIB) in agricultural products processing – A review. Journal of Food Engineering. 2014;132:39-47. DOI: 10.1016/j.jfoodeng. 2014.01.032

[88] Severini C, Baiano A, De Pilli T, Carbone BF, Derossi A. Combined treatments of blanching and dehydration: Study on potato cubes. Journal of Food Engineering. 2005;**68**(3):289-296. DOI: 10.1016/j.jfoodeng.2004.05.045

[89] Grabowski NT, Klein G. Microbiology of processed edible insect products – Results of a preliminary survey. International Journal of Food Microbiology. 2017;**243**:103-107. DOI: 10.1016/j.ijfoodmicro.2016.11.005

[90] Grabowski NT, Klein G. Microbiology of cooked and dried edible Mediterranean field crickets (*Gryllus bimaculatus*) and superworms (*Zophobas atratus*) submitted to four different heating treatments. Food Science and Technology International. 2017;**23**(1):17-23. DOI: 10.1177/1082013216652994

[91] Liceaga AM. Processing insects for use in the food and feed industry. Current Opinion in Insect Science. 2021;**48**:32-36. DOI: 10.1016/j. cois.2021.08.002

[92] Caligiani A, Marseglia A, Leni G, Baldassarre S, Maistrello L, Dossena A, et al. Composition of black soldier fly prepupae and systematic approaches for extraction and fractionation of proteins, lipids and chitin. Food Research International. 2018;**105**:812-820. DOI: 10.1016/j.foodres.2017.12.012

[93] Scholliers J, Steen L, Glorieux S, Van de Walle D, Dewettinck K, Fraeye I. The effect of temperature on structure formation in three insect batters. Food Research International. 2019;**122**:411-418. DOI: 10.1016/j.foodres.2019.04.033

[94] Son Y-J, Lee J-C, Hwang I-K, Nho CW, Kim S-H. Physicochemical properties of mealworm (*Tenebrio molitor*) powders manufactured by different industrial processes. LWT. 2019;**116**:108514. DOI: 10.1016/j. lwt.2019.108514

[95] Azagoh C, Ducept F, Garcia R, Rakotozafy L, Cuvelier M-E, Keller S, et al. Extraction and physicochemical characterization of *Tenebrio molitor* proteins. Food Research International. 2016;**88**:24-31. DOI: 10.1016/j.foodres. 2016.06.010

[96] Bußler S, Rumpold BA, Jander E, Rawel HM, Schlüter OK. Recovery and techno-functionality of flours and proteins from two edible insect species: Meal worm (*Tenebrio molitor*) and black soldier fly (*Hermetia illucens*) larvae. Heliyon. 2016;**2**(12):e00218. DOI: 10.1016/j.heliyon.2016.e00218

[97] Choi BD, Wong NAK, Auh JH. Defatting and sonication enhances protein extraction from edible insects. Korean Journal for Food Science of Animal Resources. 2017;**37**(6):955-961. DOI: 10.5851/kosfa.2017.37.6.955

[98] Mishyna M, Martinez J-JI, Chen J, Benjamin O. Extraction, characterization and functional properties of soluble proteins from edible grasshopper (*Schistocerca gregaria*) and honey bee (*Apis mellifera*). Food Research International. 2019;**116**:697-706. DOI: 10.1016/j.foodres.2018.08.098

[99] Ribeiro JC, Lima RC, Maia MRG, Almeida AA, Fonseca AJM, Cabrita ARJ, et al. Impact of defatting freeze-dried edible crickets (*Acheta domesticus* and *Gryllodes sigillatus*) on the nutritive value, overall liking and sensory profile of cereal bars. LWT. 2019;**113**(5):108335. DOI: 10.1016/j.lwt.2019.108335

[100] Zhao X, Vázquez-Gutiérrez JL, Johansson DP, Landberg R, Langton M. Yellow mealworm protein for food purposes - extraction and functional properties. PLoS One. 2016;**11**(2):e0147791. DOI: 10.1371/ journal.pone.0147791

[101] Amarender RV, Bhargava K, Dossey AT, Gamagedara S. Lipid and

protein extraction from edible insects – Crickets (Gryllidae). LWT. 2020;**125**:109222. DOI: 10.1016/j. lwt.2020.109222

[102] Kim T-K, Yong HI, Kim Y-B, Jung S, Kim H-W, Choi Y-S. Effects of organic solvent on functional properties of defatted proteins extracted from *Protaetia brevitarsis* larvae. Food Chemistry. 2021;**336**:127679. DOI: 10.1016/j.foodchem.2020.127679

[103] Birch EJ. FATS | extraction by solvent based methods. In: Wilson ID, editor. Encyclopedia of Separation Science. London: Academic Press; 2000. pp. 2794-2801. DOI: 10.1016/ B0-12-226770-2/06661-8

[104] Pan W-J, Liao A-M, Zhang J-G, Dong Z, Wei Z-J. Supercritical carbon dioxide extraction of the oak silkworm (*Antheraea pernyi*) pupal oil: Process optimization and composition determination. International Journal of Molecular Sciences. 2012;**13**(2):2354-2367. DOI: 10.3390/ijms13022354

[105] Kim SW, Jung TS, Ha YJ, Gal SW, Noh CW, Kim IS, et al. Removal of fat from crushed black soldier fly larvae by carbon dioxide supercritical extraction. Journal of Animal and Feed Sciences. 2019;**28**(1):83-88. DOI: 10.22358/ jafs/105132/2019

[106] Purschke B, Stegmann T, Schreiner M, Jäger H. Pilot-scale supercritical CO2 extraction of edible insect oil from *Tenebrio molitor* L. larvae – Influence of extraction conditions on kinetics, defatting performance and compositional properties. European Journal of Lipid Science and Technology. 2017;**119**(2):1600134. DOI: 10.1002/ ejlt.201600134

[107] Bolat B, Ugur AE, Oztop MH, Alpas H. Effects of high hydrostatic pressure assisted degreasing on the technological properties of insect powders obtained from *Acheta domesticus* & *Tenebrio molitor*. Journal of Food Engineering. 2021;**292**:110359. DOI: 10.1016/j.jfoodeng.2020.110359

[108] Laroche M, Perreault V, Marciniak A, Gravel A, Chamberland J, Doyen A. Comparison of conventional and sustainable lipid extraction methods for the production of oil and protein isolate from edible insect meal. Food. 2019;8(11):572. DOI: 10.3390/ foods8110572

[109] Delicato C, Schouteten JJ, Dewettinck K, Gellynck X, Tzompa-Sosa DA. Consumers' perception of bakery products with insect fat as partial butter replacement. Food Quality and Preference. 2020;**79**:103755. DOI: 10.1016/j.foodqual.2019.103755

[110] Tzompa-Sosa DA, Dewettinck K, Gellynck X, Schouteten JJ. Replacing vegetable oil by insect oil in food products: Effect of deodorization on the sensory evaluation. Food Research International. 2021;**141**:110140. DOI: 10.1016/j.foodres.2021.110140

[111] Belghit I, Waagbø R, Lock E-J, Liland NS. Insect-based diets high in lauric acid reduce liver lipids in freshwater Atlantic salmon. Aquaculture Nutrition. 2019;**25**(2):343-357. DOI: 10.1111/anu.12860

[112] Kumar V, Fawole FJ, Romano N, Hossain MS, Labh SN, Overturf K, et al. Insect (black soldier fly, *Hermetia illucens*) meal supplementation prevents the soybean meal-induced intestinal enteritis in rainbow trout and health benefits of using insect oil. Fish & Shellfish Immunology. 2021;**109**:116-124. DOI: 10.1016/j.fsi.2020.12.008

[113] Kamarudin MS, Rosle S, Md Yasin IS. Performance of defatted black soldier fly pre-pupae meal as fishmeal replacement in the diet of lemon fin barb hybrid fingerlings. Aquaculture Reports. 2021;**21**:100775. DOI: 10.1016/j. aqrep.2021.100775

[114] Li M, Li M, Wang G, Liu C, Shang R, Chen Y, et al. Defatted black soldier fly (*Hermetia illucens*) larvae meal can partially replace fish meal in diets for adult Chinese soft-shelled turtles. Aquaculture. 2021;**541**:736758. DOI: 10.1016/j.aquaculture.2021.736758

[115] Stejskal V, Tran HQ, Prokesová M, Zare M, Gebauer T, Policar T, et al. Defatted black soldier fly (*Hermetia illucens*) in pikeperch (*Sander lucioperca*) diets: Effects on growth performance, nutrient digestibility, fillet quality, economic and environmental sustainability. Animal Nutrition. 2022;**12**:7-19. DOI: 10.1016/j. aninu.2022.06.022

[116] Kishawy ATY, Mohammed HA, Zaglool AW, Attia MS, Hassan FAM, Roushdy EM, et al. Partial defatted black solider larvae meal as a promising strategy to replace fish meal protein in diet for Nile tilapia (*Oreochromis niloticus*): Performance, expression of protein and fat transporters, and cytokines related genes and economic efficiency. Aquaculture. 2022;555:738195. DOI: 10.1016/j.aquaculture.2022.738195

[117] Basto A, Matos E, Valente LMP. Nutritional value of different insect larvae meals as protein sources for European sea bass (*Dicentrarchus labrax*) juveniles. Aquaculture. 2020;**521**:735085. DOI: 10.1016/j. aquaculture.2020.735085

[118] Tran HQ, Van Doan H, Stejskal V. Does dietary *Tenebrio molitor* affect swimming capacity, energy use, and physiological responses of European perch *Perca fluviatilis*? Aquaculture. 2021;**539**:736610. DOI: 10.1016/j. aquaculture.2021.736610

[119] Tran HQ, Prokešová M, Zare M, Matoušek J, Ferrocino I, Gasco L, et al. Production performance, nutrient digestibility, serum biochemistry, fillet composition, intestinal microbiota and environmental impacts of European perch (*Perca fluviatilis*) fed defatted mealworm (*Tenebrio molitor*). Aquaculture. 2022;**547**:737499. DOI: 10.1016/j.aquaculture.2021.737499

[120] Tang C, Yang D, Liao H, Sun H, Liu C, Wei L, et al. Edible insects as a food source: A review. Food Production, Processing and Nutrition. 2019;1(1):8. DOI: 10.1186/s43014-019-0008-1

[121] Ferrer Llagostera P, Kallas Z, Reig L, Amores de Gea D. The use of insect meal as a sustainable feeding alternative in aquaculture: Current situation, Spanish consumers' perceptions and willingness to pay. Journal of Cleaner Production. 2019;**229**:10-21. DOI: 10.1016/j. jclepro.2019.05.012

[122] Hidalgo MC, Morales AE, Pula HJ, Tomás-Almenar C, Sánchez-Muros MJ, Melenchón F, et al. Oxidative metabolism of gut and innate immune status in skin and blood of tench (Tinca tinca) fed with different insect meals (*Hermetia illucens* and *Tenebrio molitor*). Aquaculture. 2022;**558**(1):738384. DOI: 10.1016/j. aquaculture.2022.738384

[123] Mastoraki M, Katsika L, Enes P, Guerreiro I, Kotzamanis YP, Gasco L, et al. Insect meals in feeds for juvenile gilthead seabream (Sparus aurata): Effects on growth, blood chemistry, hepatic metabolic enzymes, body composition and nutrient utilization. Aquaculture. 2022;**561**(1):738674. DOI: 10.1016/j.aquaculture.2022.738674

[124] González CM, Garzón R, Rosell CM. Insects as ingredients for bakery goods.

A comparison study of *H. illucens*, *A. domestica* and *T. molitor* flours. Innovative Food Science and Emerging Technologies. 2019;**51**:205-210. DOI: 10.1016/j.ifset.2018.03.021

[125] Navarro del Hierro J, Gutiérrez-Docio A, Otero P, Reglero G, Martin D. Characterization, antioxidant activity, and inhibitory effect on pancreatic lipase of extracts from the edible insects *Acheta domesticus* and *Tenebrio molitor*. Food Chemistry. 2020;**309**:125742. DOI: 10.1016/j.foodchem.2019.125742

[126] Montiel-Aguilar LJ, Torres-Castillo JA, Rodríguez-Servin R, López-Flores AB, Aguirre-Arzola VE, Méndez-Zamora G, et al. Nutraceutical effects of bioactive peptides obtained from *Pterophylla beltrani* (Bolivar & Bolivar) protein isolates. Journal of Asia-Pacific Entomology. 2020;**23**(3):756-761. DOI: 10.1016/j.aspen.2020.06.006

[127] Weru J, Chege P, Kinyuru J. Nutritional potential of edible insects: A systematic review of published data. International Journal of Tropical Insect Science. 2021;**41**(3):2015-2037. DOI: 10.1007/s42690-021-00464-0

[128] Hernández-Álvarez A-J, Mondor M, Piña-Domínguez I-A, Sánchez-Velázquez O-A, Melgar LG. Drying technologies for edible insects and their derived ingredients. Drying Technology. 2021;**39**(13):1991-2009. DOI: 10.1080/ 07373937.2021.1915796

[129] Menozzi D, Sogari G, Veneziani M, Simoni E, Mora C. Eating novel foods: An application of the theory of planned behaviour to predict the consumption of an insect-based product. Food Quality and Preference. 2017;**59**:27-34. DOI: 10.1016/j.foodqual.2017.02.001

[130] Rawski M, Mazurkiewicz J, Kierończyk B, Józefiak D. Black soldier fly full-fat larvae meal as an alternative to fish meal and fish oil in Siberian sturgeon nutrition: The effects on physical properties of the feed, animal growth performance, and feed acceptance and utilization. Animals. 2020;**10**(11):2119. DOI: 10.3390/ani10112119

[131] de Souza VJ, Andronicos NM, Kolakshyapati M, Hilliar M, Sibanda TZ, Andrew NR, et al. Black soldier fly larvae in broiler diets improve broiler performance and modulate the immune system. Animal Nutrition. 2021;7(3):695-706. DOI: 10.1016/j. aninu.2020.08.014

[132] Håkenåsen IM, Grepperud GH, Hansen JØ, Øverland M, Ånestad RM, Mydland LT. Full-fat insect meal in pelleted diets for weaned piglets: Effects on growth performance, nutrient digestibility, gastrointestinal function, and microbiota. Animal Feed Science and Technology. 2021;**281**:115086. DOI: 10.1016/j.anifeedsci.2021.115086

[133] Zhao Q, Xie T, Hong X, Zhou Y, Fan L, Liu Y, et al. Modification of functional properties of perilla protein isolate by high-intensity ultrasonic treatment and the stability of o/w emulsion. Food Chemistry. 2022;**368**:130848. DOI: 10.1016/j. foodchem.2021.130848

[134] Matthäus B, Piofczyk T, Katz H, Pudel F. Renewable resources from insects: Exploitation, properties, and refining of fat obtained by cold-pressing from *Hermetia illucens* (black soldier fly) larvae. European Journal of Lipid Science and Technology. 2019;**121**(7):1800376. DOI: 10.1002/ejlt.201800376

[135] Kim D, Oh I. The characteristic of insect oil for a potential component of oleogel and its application as a solid fat replacer in cookies. Gels. 2022;8(6):355. DOI: 10.3390/gels8060355 [136] Ruban AA, Novikova (Zakharova) MB, Kostin AA. Effective viscosity of lecithin solutions and fat emulsions of black soldier fly larvae with different lecithin content. Food Systems. 2021;4(3):220-225. DOI: 10/21323/ 2618-9771-2021-4-3-220-225

[137] Chou T-H, Nugroho DS, Cheng Y-S, Chang J-Y. Development and characterization of nano-emulsions based on oil extracted from black soldier fly larvae. Applied Biochemistry and Biotechnology. 2020;**191**(1):331-345. DOI: 10.1007/s12010-019-03210-y

[138] Chou T-H, Nugroho DS, Chang J-Y, Cheng Y-S, Liang C-H, Deng M-J. Encapsulation and characterization of nanoemulsions based on an antioxidative polymeric amphiphile for topical apigenin delivery. Polymers (Basel). 2021;**13**(7):1016. DOI: 10.3390/ polym13071016

[139] Saviane A, Tassoni L, Naviglio D, Lupi D, Savoldelli S, Bianchi G, et al. Mechanical processing of *Hermetia illucens* larvae and *Bombyx mori* pupae produces oils with antimicrobial activity. Animals. 2021;**11**(3):783. DOI: 10.3390/ ani11030783

[140] Spranghers T, Michiels J, Vrancx J, Ovyn A, Eeckhout M, De Clercq P, et al. Gut antimicrobial effects and nutritional value of black soldier fly (*Hermetia illucens* L.) prepupae for weaned piglets. Animal Feed Science and Technology. 2018;**235**:33-42. DOI: 10.1016/j. anifeedsci.2017.08.012

[141] Yu M, Li Z, Chen W, Rong T, Wang G, Ma X. Hermetia illucens larvae as a potential dietary protein source altered the microbiota and modulated mucosal immune status in the colon of finishing pigs. Journal of Animal Science and Biotechnology. 2019;**10**(1):50. DOI: 10.1186/s40104-019-0358-1 [142] Dabbou S, Ferrocino I, Gasco L, Schiavone A, Trocino A, Xiccato G, et al. Antimicrobial effects of black soldier fly and yellow mealworm fats and their impact on gut microbiota of growing rabbits. Animals. 2020;**10**(8):1292. DOI: 10.3390/ ani10081292

[143] Yu M, Li Z, Chen W, Wang G, Rong T, Liu Z, et al. Hermetia illucens larvae as a fishmeal replacement alters intestinal specific bacterial populations and immune homeostasis in weanling piglets. Journal of Animal Science. 2020;**98**(3):1-13. DOI: 10.1093/jas/ skz395/5810268

[144] Józefiak A, Nogales-Mérida S, Rawski M, Kierończyk B, Mazurkiewicz J. Effects of insect diets on the gastrointestinal tract health and growth performance of Siberian sturgeon (Acipenser baerii Brandt, 1869). BMC Veterinary Research. 2019;**15**(1):348. DOI: 10.1186/s12917-019-2070-y

[145] Sypniewski J, Kierończyk B, Benzertiha A, Mikołajczak Z, Pruszyńska-Oszmałek E, Kołodziejski P, et al. Replacement of soybean oil by Hermetia illucens fat in Turkey nutrition: Effect on performance, digestibility, microbial community, immune and physiological status and final product quality. British Poultry Science. 2020;**61**(3):294-302. DOI: 10.1080/ 00071668.2020.1716302

[146] Zielińska E, Karaś M, Baraniak B. Comparison of functional properties of edible insects and protein preparations thereof. LWT. 2018;**91**:168-174. DOI: 10.1016/j.lwt.2018.01.058

[147] Jiang Y, Zhu Y, Zheng Y, Liu Z, Zhong Y, Deng Y, et al. Effects of salting-in/out-assisted extractions on structural, physicochemical and functional properties of *Tenebrio molitor*

larvae protein isolates. Food Chemistry. 2021;**338**:128158. DOI: 10.1016/j. foodchem.2020.128158

[148] Valle FR, Mena MH, Bourges H. An investigation into insect protein. Journal of Food Processing & Preservation. 1982;**6**(2):99-110. DOI: 10.1111/j.1745-4549.1982.tb00645.x

[149] Patrignani F, Parrotta L, Del Duca S, Vannini L, Camprini L, Dalla Rosa M, et al. Potential of *Yarrowia lipolytica* and *Debaryomyces hansenii* strains to produce high quality food ingredients based on cricket powder. LWT. 2020;**119**:108866. DOI: 10.1016/j. lwt.2019.108866

[150] Dion-Poulin A, Laroche M, Doyen A, Turgeon SL. Functionality of cricket and mealworm hydrolysates generated after pretreatment of meals with high hydrostatic pressures. Molecules. 2020;**25**(22):5366. DOI: 10.3390/molecules25225366

[151] Mariod AA, Fadul H. Extraction and characterization of gelatin from two edible Sudanese insects and its applications in ice cream making. Food Science and Technology International. 2015;**21**(5):380-391. DOI: 10.1177/ 1082013214541137

[152] Kim T-K, Yong HI, Jung S, Sung J-M, Jang HW, Choi Y-S. Physicochemical and textural properties of emulsions prepared from the larvae of the edible insects *Tenebrio molitor*, *Allomyrina dichotoma*, and *Protaetia brevitarsis seulensis*. Journal of Animal Science and Technology. 2021;**63**(2):417-425. DOI: 10.5187/jast.2021.e25

[153] Azzollini D, Derossi A, Fogliano V, Lakemond CMM, Severini C. Effects of formulation and process conditions on microstructure, texture and digestibility of extruded insect-riched snacks. Innovative Food Science and Emerging Technologies. 2018;**45**:344-353. DOI: 10.1016/j.ifset.2017.11.017

[154] Alam MR, Scampicchio M, Angeli S, Ferrentino G. Effect of hot melt extrusion on physical and functional properties of insect based extruded products. Journal of Food Engineering. 2019;**259**:44-51. DOI: 10.1016/j. jfoodeng.2019.04.021

[155] Sánchez M, Gómez C, Avendaño C, Harmsen I, Ortiz D, Ceballos R, et al. House fly (*Musca domestica*) larvae meal as an ingredient with high nutritional value: Microencapsulation and improvement of organoleptic characteristics. Food Research International. 2021;**145**(5):110423. DOI: 10.1016/j.foodres.2021.110423

[156] Sánchez M, Villamizar-Sarmiento MG, Harmsen I, Valdés F, Villanueva V, Ceballos R, et al. Encapsulation of house fly larvae (*Musca domestica*) meal by ionic gelation as a strategy to develop a novel nutritive food ingredient with improved aroma and appearance. LWT. 2022;**163**:113597. DOI: 10.1016/j.lwt.2022.113597

[157] Kim J, Lee HE, Kim Y, Yang J, Lee S-J, Jung YH. Development of a postprocessing method to reduce the unique off-flavor of *Allomyrina dichotoma*: Yeast fermentation. LWT. 2021;**150**(5):111940. DOI: 10.1016/j.lwt.2021.111940

[158] Lee HE, Kim J, Kim Y, Bang WY, Yang J, Lee S-J, et al. Identification and improvement of volatile profiles of *Allomyrina dichotoma* larvae by fermentation with lactic acid bacteria. Food Bioscience. 2021;**43**:101257. DOI: 10.1016/j.fbio.2021.101257

[159] Klunder HC, Wolkers-Rooijackers J, Korpela JM, Nout MJR. Microbiological aspects of processing and storage of edible insects. Food Control. 2012;**26**(2):628-631. DOI: 10.1016/j. foodcont.2012.02.013

[160] Lee W, Kim M-A, Park I, Hwang JS, Na M, Bae J-S. Novel direct factor Xa inhibitory compounds from *Tenebrio molitor* with anti-platelet aggregation activity. Food and Chemical Toxicology. 2017;**109**(6):19-27. DOI: 10.1016/j. fct.2017.08.026

[161] Lee W, Lee H, Kim M-A, Choi J, Kim K-M, Hwang JS, et al. Evaluation of novel factor Xa inhibitors from *Oxya chinensis sinuosa* with antiplatelet aggregation activity. Scientific Reports. 2017;7(1):7934. DOI: 10.1038/ s41598-017-08330-1

[162] Lee J, Lee W, Kim M-A, Hwang JS, Na M, Bae J-S. Inhibition of platelet aggregation and thrombosis by indole alkaloids isolated from the edible insect *Protaetia brevitarsis seulensis* (Kolbe). Journal of Cellular and Molecular Medicine. 2017;**21**(6):1217-1227. DOI: 10.1111/jcmm.13055

[163] Pyo S-J, Kang D-G, Jung C, Sohn H-Y. Anti-thrombotic, anti-oxidant and haemolysis activities of six edible insect species. Food. 2020;**9**(4):401. DOI: 10.3390/foods9040401

[164] Vogel H, Müller A, Heckel DG, Gutzeit H, Vilcinskas A. Nutritional immunology: Diversification and dietdependent expression of antimicrobial peptides in the black soldier fly *Hermetia illucens*. Developmental and Comparative Immunology. 2018;**78**:141-148. DOI: 10.1016/j.dci.2017.09.008

[165] Shin HS, Park S-I. Novel attacin from *Hermetia illucens*: cDNA cloning, characterization, and antibacterial properties. Preparative Biochemistry & Biotechnology. 2019;**49**(3):279-285. DOI: 10.1080/10826068.2018.1541807 [166] Park S-I, Kim J-W, Yoe SM. Purification and characterization of a novel antibacterial peptide from black soldier fly (*Hermetia illucens*) larvae. Developmental and Comparative Immunology. 2015;**52**(1):98-106. DOI: 10.1016/j.dci.2015.04.018

[167] Baigts-Allende D, Doost AS, Ramírez-Rodrigues M, Dewettinck K, Van der Meeren P, de Meulenaer B, et al. Insect protein concentrates from Mexican edible insects: Structural and functional characterization. LWT. 2021;**152**:112267. DOI: 10.1016/j.lwt.2021.112267

[168] Tello A, Aganovic K, Parniakov O, Carter A, Heinz V, Smetana S. Product development and environmental impact of an insect-based milk alternative. Future Foods. 2021;**4**:100080. DOI: 10.1016/j.fufo.2021.100080

[169] Zhang L, Xiao Q, Wang Y, Hu J, Xiong H, Zhao Q. Effects of sequential enzymatic hydrolysis and transglutaminase crosslinking on functional, rheological, and structural properties of whey protein isolate. LWT. 2022;**153**:112415. DOI: 10.1016/j. lwt.2021.112415

[170] Ndiritu AK, Kinyuru JN, Kenji GM, Gichuhi PN. Extraction technique influences the physico-chemical characteristics and functional properties of edible crickets (*Acheta domesticus*) protein concentrate. Journal of Food Measurement and Characterization. 2017;**11**(4):2013-2021. DOI: 10.1007/ s11694-017-9584-4

[171] Janssen RH, Canelli G, Sanders MG, Bakx EJ, Lakemond CMM, Fogliano V, et al. Iron-polyphenol complexes cause blackening upon grinding *Hermetia illucens* (black soldier fly) larvae. Scientific Reports. 2019;**9**(1):2967. DOI: 10.1038/s41598-019-38923-x

[172] Akpossan R, Digbeu Y, Koffi M, Kouadio J, Dué E, Kouamé P. Protein fractions and functional properties of dried *Imbrasia oyemensis* larvae fullfat and defatted flours. International Journal of Biochemistry Research & Review. 2015;5(2):116-126. DOI: 10.9734/ IJBCRR/2015/12178

[173] dos Santos OV, Dias PCS, Soares SD, da Conceição LRV, Teixeira-Costa BE. Artisanal oil obtained from insects' larvae (*Speciomerus ruficornis*): Fatty acids composition, physicochemical, nutritional and antioxidant properties for application in food. European Food Research and Technology. 2021;**247**(7):1803-1813. DOI: 10.1007/ s00217-021-03752-8

[174] Borrelli L, Varriale L, Dipineto L, Pace A, Menna LF, Fioretti A. Insect derived lauric acid as promising alternative strategy to antibiotics in the antimicrobial resistance scenario. Frontiers in Microbiology. 2021;**12**(3):620798. DOI: 10.3389/ fmicb.2021.620798

[175] Dayrit FM. The properties of lauric acid and their significance in coconut oil. Journal of the American Oil Chemists' Society. 2015;**92**(1):1-15. DOI: 10.1007/ s11746-014-2562-7

[176] Herdiyati Y, Astrid Y, Shadrina AAN, Wiani I, Satari MH, Kurnia D. Potential fatty acid as antibacterial agent against oral bacteria of *Streptococcus mutans* and *Streptococcus sanguinis* from basil (*Ocimum americanum*): *In vitro* and *in silico* studies. Current Drug Discovery Technologies. 2021;**18**(4):532-541. DOI: 10.2174/157016 3817666200712171652

[177] Lekshmi Sheela D, Nazeem PA, Narayanankutty A, Manalil JJ, Raghavamenon AC. *In silico* and wet lab studies reveal the cholesterol lowering efficacy of lauric acid, a medium chain fat of coconut oil. Plant Foods for Human Nutrition. 2016;**71**(4):410-415. DOI: 10.1007/s11130-016-0577-y

[178] Alexander P, Berri A, Moran D,
Reay D, Rounsevell MDA. The global environmental paw print of pet food. Global Environmental Change.
2020;65:102153. DOI: 10.1016/j.
gloenvcha.2020.102153

[179] Okin GS. Environmental impacts of food consumption by dogs and cats. PLoS One. 2017;**12**(8):e0181301. DOI: 10.1371/ journal.pone.0181301

[180] Swanson KS, Carter RA, Yount TP, Aretz J, Buff PR. Nutritional sustainability of pet foods. Advances in Nutrition. 2013;4(2):141-150. DOI: 10.3945/an.112.003335

[181] Henchion M, Hayes M, Mullen A, Fenelon M, Tiwari B. Future protein supply and demand: Strategies and factors influencing a sustainable equilibrium. Food. 2017;**6**(7):53. DOI: 10.3390/foods6070053

[182] Batal A, Dale N. Feedstuffs
Ingredient Analysis Table: 2016 Edition.
[Internet]. 2016. Available from: https:// feedstuffs.farmcentric.com/mdfm/
Feeess50/author/427/2015/11/Feedstuffs_
RIBG_Ingredient_Analysis_Table_2016.
pdf. [Accessed: March 15, 2021]

[183] Luna D, Carrasco C, Álvarez D, González C, Egaña JI, Figueroa J. Exploring anhedonia in kennelled dogs: Could coping styles affect hedonic preferences for sweet and umami flavours? Animals. 2020;**10**(11):2087. DOI: 10.3390/ ani10112087

[184] Wang W, Zhou X, Liu Y. Characterization and evaluation of umami taste: A review. TrAC Trends in Analytical Chemistry. 2020;**127**:115876. DOI: 10.1016/j.trac.2020.115876

[185] Smetana S, Ashtari Larki N, Pernutz C, Franke K, Bindrich U, Toepfl S, et al. Structure design of insectbased meat analogs with high-moisture extrusion. Journal of Food Engineering. 2018;**229**:83-85. DOI: 10.1016/j. jfoodeng.2017.06.035

[186] Kim T-K, Yong HI, Cha JY, Park S-Y, Jung S, Choi Y-S. Drying-induced restructured jerky analog developed using a combination of edible insect protein and textured vegetable protein. Food Chemistry. 2022;**373**:131519. DOI: 10.1016/j.foodchem.2021.131519

[187] Roncolini A, Milanović V, Aquilanti L, Cardinali F, Garofalo C, Sabbatini R, et al. Lesser mealworm (*Alphitobius diaperinus*) powder as a novel baking ingredient for manufacturing high-protein, mineral-dense snacks. Food Research International. 2020;**131**:109031. DOI: 10.1016/j.foodres.2020.109031

[188] Pasini G, Cullere M, Vegro M, Simonato B, Dalle ZA. Potentiality of protein fractions from the house cricket (*Acheta domesticus*) and yellow mealworm (*Tenebrio molitor*) for pasta formulation. LWT. 2022;**164**:113638. DOI: 10.1016/j.lwt.2022.113638

[189] Borremans A, Lenaerts S, Crauwels S, Lievens B, Van Campenhout L. Marination and fermentation of yellow mealworm larvae (*Tenebrio molitor*). Food Control. 2018;**92**:47-52. DOI: 10.1016/j. foodcont.2018.04.036

[190] Kewuyemi YO, Kesa H, Chinma CE, Adebo OA. Fermented edible insects for promoting food security in Africa. Insects. 2020;**11**(5):283. DOI: 10.3390/ insects11050283 [191] Mintah BK, He R, Dabbour M, Xiang J, Agyekum AA, Ma H. Technofunctional attribute and antioxidative capacity of edible insect protein preparations and hydrolysates thereof: Effect of multiple mode sonochemical action. Ultrasonics Sonochemistry. 2019;**58**(4):104676. DOI: 10.1016/j. ultsonch.2019.104676

[192] Rossi S, Parrotta L, Del Duca S, Rosa MD, Patrignani F, Schluter O, et al. Effect of Yarrowia lipolytica RO25 cricket-based hydrolysates on sourdough quality parameters. LWT. 2021;**148**:111760. DOI: 10.1016/j. lwt.2021.111760

[193] Ray S, Raychaudhuri U, Chakraborty R. An overview of encapsulation of active compounds used in food products by drying technology. Food Bioscience. 2016;**13**:76-83. DOI: 10.1016/j.fbio.2015.12.009

[194] Siccama JW, Pegiou E, Zhang L, Mumm R, Hall RD, Boom RM, et al. Maltodextrin improves physical properties and volatile compound retention of spray-dried asparagus concentrate. LWT. 2021;**142**:111058. DOI: 10.1016/j.lwt.2021.111058

[195] Leylak C, Özdemir KS, Gurakan GC, Ogel ZB. Optimisation of spray drying parameters for Lactobacillus acidophilus encapsulation in whey and gum Arabic: Its application in yoghurt. International Dairy Journal. 2021;**112**:104865. DOI: 10.1016/j. idairyj.2020.104865

[196] Verma K, Tarafdar A, Kumar D, Kumar Y, Rana JS, Badgujar PC. Formulation and characterization of nano-curcumin fortified milk cream powder through microfluidization and spray drying. Food Research International. 2022;**160**:111705. DOI: 10.1016/j.foodres.2022.111705

[197] Obradović N, Volić M, Nedović V, Rakin M, Bugarski B. Microencapsulation of probiotic starter culture in protein– carbohydrate carriers using spray and freeze-drying processes: Implementation in whey-based beverages. Journal of Food Engineering. 2022;**321**:110948. DOI: 10.1016/j.jfoodeng.2022.110948

[198] FAO/WHO. Codex Alimentarius: Food Labelling. 5th ed. Rome: Food and Agriculture Organization of the United Nations; 2007

[199] Radosevich J, McGee N,
Rawson NE. Functional ingredients in the pet food industry. In: Bagchi D, editor. Nutraceutical and Functional Food Regulations in the United States and around the World.
3rd ed. London: Academic Press;
2019. pp. 611-616. DOI: 10.1016/ B978-0-12-816467-9.00038-1

[200] Schleicher M, Cash SB, Freeman LM. Determinants of pet food purchasing decisions. The Canadian Veterinary Journal. 2019;**60**(6):644-650

[201] Dima C, Dima S. Bioaccessibility study of calcium and vitamin D3 co-microencapsulated in water-in-oil-inwater double emulsions. Food Chemistry. 2020;**303**:125416. DOI: 10.1016/j. foodchem.2019.125416

[202] Boonlao N, Ruktanonchai UR, Anal AK. Enhancing bioaccessibility and bioavailability of carotenoids using emulsion-based delivery systems. Colloids Surfaces B Biointerfaces. 2022;**209**:112211. DOI: 10.1016/j. colsurfb.2021.112211

[203] Xu W, Lv K, Mu W, Zhou S, Yang Y. Encapsulation of α -tocopherol in whey protein isolate/chitosan particles using oil-in-water emulsion with optimal stability and bioaccessibility. LWT. 2021;**148**:111724. DOI: 10.1016/j. lwt.2021.111724 [204] Aditya NP, Aditya S, Yang H, Kim HW, Park SO, Ko S. Co-delivery of hydrophobic curcumin and hydrophilic catechin by a water-in-oil-in-water double emulsion. Food Chemistry. 2015;**173**:7-13. DOI: 10.1016/j.foodchem.2014.09.131

[205] Yu H, Wang H, Su W,

Song Y, Zaky AA, Abd El-Aty AM, et al. Co-delivery of hydrophobic astaxanthin and hydrophilic phycocyanin by a pH-sensitive water-in-oil-in-water double emulsion-filled gellan gum hydrogel. Food Hydrocolloids. 2022;**131**:107810. DOI: 10.1016/j.foodhyd.2022.107810

[206] Aditya NP, Macedo AS, Doktorovova S, Souto EB, Kim S, Chang PS, et al. Development and evaluation of lipid nanocarriers for quercetin delivery: A comparative study of solid lipid nanoparticles (SLN), nanostructured lipid carriers (NLC), and lipid nanoemulsions (LNE). LWT. 2014;**59**(1):115-121. DOI: 10.1016/j. lwt.2014.04.058

[207] Muschiolik G, Dickinson E. Double emulsions relevant to food systems: Preparation, stability, and applications. Comprehensive Reviews in Food Science and Food Safety. 2017;**16**(3):532-555. DOI: 10.1111/1541-4337.12261

[208] Gravel A, Doyen A. The use of edible insect proteins in food:
Challenges and issues related to their functional properties. Innovative Food Science and Emerging Technologies.
2020;59(3):102272. DOI: 10.1016/j.
ifset.2019.102272

[209] Hall FG, Jones OG, O'Haire ME, Liceaga AM. Functional properties of tropical banded cricket (*Gryllodes sigillatus*) protein hydrolysates. Food Chemistry. 2017;**224**(11):414-422. DOI: 10.1016/j.foodchem.2016.11.138

[210] Ngo D-H, Kim S-K. Antioxidant effects of chitin, chitosan, and their

derivatives. Advances in Food and Nutrition Research. 2014;**73**:15-31. DOI: 10.1016/B978-0-12-800268-1. 00002-0

[211] Mouithys-Mickalad A, Schmitt E, Dalim M, Franck T, Tome NM, van Spankeren M, et al. Black soldier fly (*Hermetia illucens*) larvae protein derivatives: Potential to promote animal health. Animals. 2020;**10**(6):941. DOI: 10.3390/ani10060941

[212] Liu Y, Wan S, Liu J, Zou Y, Liao S. Antioxidant activity and stability study of peptides from enzymatically hydrolyzed male silkmoth. Journal of Food Processing & Preservation. 2017;**41**(1):e13081. DOI: 10.1111/ jfpp.13081

[213] Ma S, Li X, Sun Y, Mi R, Li Y, Wen Z, et al. Enzymatic hydrolysis of defatted *Antheraea pernyi* (Lepidoptera: Saturniidae) pupa protein by combined neutral protease yield peptides with antioxidant activity. Journal of Insect Science. 2021;**21**(2):276-279. DOI: 10.1093/jisesa/ieab013

[214] Dutta P, Dey T, Manna P, Kalita J. Antioxidant potential of *Vespa affinis* L., a traditional edible insect species of north East India. PLoS One. 2016;**11**(5):e0156107. DOI: 10.1371/ journal.pone.0156107

[215] Ai H, Wang F, Xia Y, Chen X, Lei C. Antioxidant, antifungal and antiviral activities of chitosan from the larvae of housefly, *Musca domestica* L. Food Chemistry. 2012;**132**(1):493-498. DOI: 10.1016/j.foodchem.2011.11.033

[216] Di Mattia C, Battista N, Sacchetti G, Serafini M. Antioxidant activities *in vitro* of water and liposoluble extracts obtained by different species of edible insects and invertebrates. Frontiers in Nutrition. 2019;**6**:1-7. DOI: 10.3389/ fnut.2019.00106/full [217] Dechakhamphu A, Wongchum N. Investigation of the kinetic properties of Phyllanthus chamaepeuce Ridl. Extracts for the inhibition of pancreatic lipase activity. Journal of Herbal Medicine. 2022;**32**:100508. DOI: 10.1016/j. hermed.2021.100508

[218] Yoon S, Wong NAK, Chae M, Auh J-H. Comparative characterization of protein hydrolysates from three edible insects: Mealworm larvae, adult crickets, and silkworm pupae. Food. 2019;8(11):563. DOI: 10.3390/ foods8110563

[219] Sousa P, Borges S, Pintado M. Enzymatic hydrolysis of insect *Alphitobius diaperinus* towards the development of bioactive peptide hydrolysates. Food & Function. 2020;**11**(4):3539-3548. DOI: 10.1039/ D0FO00188K

[220] Hall F, Johnson PE, Liceaga A.
Effect of enzymatic hydrolysis on bioactive properties and allergenicity of cricket (*Gryllodes sigillatus*) protein. Food Chemistry. 2018;262(3):39-47.
DOI: 10.1016/j.foodchem.2018.04.058

[221] Hall F, Reddivari L, Liceaga AM. Identification and characterization of edible cricket peptides on hypertensive and glycemic *in vitro* inhibition and their anti-inflammatory activity on RAW 264.7 macrophage cells. Nutrients. 2020;12(11):3588. DOI: 10.3390/ nu12113588

[222] Ahn MY, Kim BJ, Kim HJ, Jin JM, Yoon HJ, Hwang JS, et al. Anti-diabetic activity of field cricket glycosaminoglycan by ameliorating oxidative stress. BMC Complementary Medicine and Therapies. 2020;**20**(1):232. DOI: 10.1186/s12906-020-03027-x

[223] Zhao J-G, Wang H-Y, Wei Z-G, Zhang Y-Q. Therapeutic effects of

ethanolic extract from the green cocoon shell of silkworm *Bombyx mori* on type 2 diabetic mice and its hypoglycaemic mechanism. Toxicology Research. 2019;**8**(3):407-420. DOI: 10.1039/ c8tx00294k

[224] Leem JY, Nishimura C,
Kurata S, Shimada I, Kobayashi A,
Natori S. Purification and
characterization of N-βAlanyl-5-S-glutathionyl-3,4dihydroxyphenylalanine, a
novel antibacterial substance of *Sarcophaga peregrina* (flesh fly).
The Journal of Biological Chemistry.
1996;271(23):13573-13577. DOI: 10.1074/
jbc.271.23.13573

[225] Leem JY, Jeong IJ, Park KT, Park HY. Isolation of p-hydroxycinnamaldehyde as an antibacterial substance from the saw fly *Acantholyda parki* S. FEBS Letters. 1999;**442**(1):53-56. DOI: 10.1016/ S0014-5793(98)01614-7

[226] Azmiera N, Krasilnikova A, Sahudin S, Al-Talib H, Heo CC. Antimicrobial peptides isolated from insects and their potential applications. Journal of Asia-Pacific Entomology. 2022;**25**(2):101892. DOI: 10.1016/j. aspen.2022.101892

[227] Xia J, Ge C, Yao H. Antimicrobial peptides from black soldier fly (*Hermetia illucens*) as potential antimicrobial factors representing an alternative to antibiotics in livestock farming. Animals. 2021;**11**(7):1937. DOI: 10.3390/ ani11071937

[228] Hadj Saadoun J, Sogari G, Bernini V, Camorali C, Rossi F, Neviani E, et al. A critical review of intrinsic and extrinsic antimicrobial properties of insects. Trends in Food Science and Technology. 2022;**122**:40-48. DOI: 10.1016/j. tifs.2022.02.018 [229] Sahoo A, Swain SS, Behera A, Sahoo G, Mahapatra PK, Panda SK. Antimicrobial peptides derived from insects offer a novel therapeutic option to combat biofilm: A review. Frontiers in Microbiology. 2021;**12**:661195. DOI: 10.3389/fmicb.2021.661195/full

[230] Van Moll L, De Smet J, Paas A, Tegtmeier D, Vilcinskas A, Cos P, et al. *In vitro* evaluation of antimicrobial peptides from the black soldier fly (Hermetia illucens) against a selection of human pathogens. Microbiology Spectrum. 2022;**10**(1):e0166421. DOI: 10.1128/ spectrum.01664-21

