# Acclimation to daily thermal variability drives the metabolic performance curve

Francisco Bozinovic<sup>1</sup>, Tamara P. Catalan<sup>1</sup>, Sergio A. Estay<sup>2</sup> and Pablo Sabat<sup>3</sup>

<sup>1</sup>Departamento de Ecología, LINC-Global, Facultad de Ciencias Biológicas, Universidad Católica de Chile, Santiago, Chile, <sup>2</sup>Instituto de Ciencias Ambientales y Evolutivas, Facultad de Ciencias, Universidad Austral de Chile, Valdivia, Chile and <sup>3</sup>Departamento de Ciencias Ecológicas, Facultad de Ciencias, Universidad de Chile, Santiago, Chile

#### **ABSTRACT**

**Background:** Among the predictions of the effect of future climate change, the impact of thermal conditions at local levels on the physiological performance of individuals and their acclimation capacities is key to understanding animals' responses to global warming.

**Goal:** Test for the effect of acclimation to environmental thermal variability, namely  $24 \pm 0^{\circ}$ C,  $24 \pm 4^{\circ}$ C, and  $24 \pm 8^{\circ}$ C, on the metabolic performance curve in the mealworm beetle *Tenebrio molitor*.

**Results:** Maximum resting metabolism and metabolic breadth were significantly different, but optimal temperature was similar, between treatments. Thus, increases in ambient thermal variability caused a reduction in maximum performance at each level of acclimation, with a decrease of almost 50% between the nil variability and  $\pm$  8°C daily variability treatments.

**Conclusions:** If thermal variability changes in any of the directions forecast by climatologists, ecologists will have to use mechanistic and modelling approaches based on physiological and biophysical traits to predict the biodiversity consequences of climate change.

Keywords: fitness, global warming, physiological responses, thermal variability.

# **INTRODUCTION**

Global climate change poses one of the greatest threats to biodiversity (Dawson et al., 2011). Anthropogenic activities are likely to lead to increased frequency of climatic extremes (e.g. heat waves and severe droughts or floods), as well as increased climatic variability in certain regions of the world (Lean and Rind, 2008; Rahmstorf and Coumou, 2011). Theoretical and empirical approaches indicate that global warming impacts both the mean temperatures of local environments, and the magnitude of diel and seasonal variation in temperature (Easterling et al., 2000; Parmesan et al., 2000; Burroughs, 2007; Thompson et al., 2013). Indeed, in addition to increases in mean temperatures on Earth, the Intergovernmental Panel on Climate Change

Correspondence: F. Bozinovic, Departamento de Ecología, Universidad Católica de Chile, Santiago 6513677, Chile. e-mail: fbozinovic@bio.puc.cl

Consult the copyright statement on the inside front cover for non-commercial copying policies.

(IPCC, 2007) predicts increases in extreme events such as heat waves. The expected increased frequency of extreme environmental conditions is likely to have an even greater impact on biodiversity than the increase in average temperatures (Coumou and Rahmstorf, 2012). Although the potential impacts of warming are widely recognized by ecologists, studies in thermal ecology have focused on the impact of changes in average values. But the variability in these conditions (e.g. extreme temperature events) could be as important as average values, and could also be an important selective force on organisms in the wild (Jentsch et al., 2007; Lalouette et al., 2007; Terblanche et al., 2010; Bannerman et al., 2011; Bozinovic et al., 2011a; Thompson et al., 2013). For instance, during 2010 in Australia, a two-week heat wave led to the death of thousands of birds in the wild. This happened when environmental temperatures exceeded the maximum critical thermal limits of these birds (Miller and Stillman, 2012). Thus, observations and experiments that quantify not only the effect of increases in mean environmental variables but also the effect of environmental variation on physiological traits are important for inferring ecological and evolutionary responses to climate change and the mechanisms by which organisms cope with this variation (Chown and Terblanche, 2006; Terblanche et al., 2008; Bozinovic et al., 2011b; Catalán et al., 2012a, 2012b; DeVries and Appel, 2013).

Several studies have evaluated the effect of thermal variability on life-history traits, including developmental time (Ragland and Kingsolver, 2008), hatching success (Ji et al., 2007), and some phenotypic characteristics of the progeny (Pétavy et al., 2004; Folguera et al., 2009). Williams et al. (2012) reported that thermal variability increases the effect of fall (autumn) warming and drives metabolic depression of an overwintering butterfly (see also Paaijmans et al., 2010; Krams et al., 2011). Others have focused on the effect on parameters linked to population dynamics of the organisms (Orcutt and Porter, 1983; Estay et al., 2011). In the wild, ectotherms are continuously exposed to several short-term variations in environmental conditions. Organisms deal with this variability through thermal acclimation and/or acclimatization, affecting current and future survival of natural populations (Terblanche et al., 2010). In this context, the study of the impact of thermal conditions on the performance of individuals and their plastic responses to it, such as lethal and sub-lethal effects, are key to understanding the responses of biota to the environment and to climate change (Pörtner et al., 2006; Somero, 2011).

In this study, we experimentally tested the effect of acclimation to environmental thermal variability on the metabolic rate (SMR) performance curve in the mealworm beetle *Tenebrio molitor* (Coleoptera: Tenebrionidae). We hypothesized that the net effect of thermal variability on SMR would depend on the relative advantages (or disadvantages) of being exposed to low and high extreme temperatures. We address two main questions:

- Is thermal variability a modulator of the maximal SMR of those organisms acclimated to the same average temperature?
- How does thermal variability modify optimum temperature in those individuals acclimated to the same average temperature?

# **MATERIALS AND METHODS**

# **Experimental design**

We used larvae of *T. molitor* reared under laboratory conditions with a light/dark cycle of 12:12 h and fed a mixture of flour and oats *ad libitum*. Water was supplied *ad libitum* as wet sponges. We used larvae because they are extremely sensitive to environmental conditions

(Min et al., 2006; Morales-Ramos et al., 2010). We assigned individuals randomly to one of three thermal treatments in climatic chambers. In one of the treatments, we maintained the temperature at  $24^{\circ}$ C ( $\delta = 0$ , the  $\delta 0$  group, i.e. the constant temperature regime). In the other two treatments (i.e. the alternating temperature regimes), temperature varied during the day, reaching a maximum during daylight and a minimum during the night:  $28/12^{\circ}$ C ( $\delta = 8$ , the  $\delta 8$  group) and  $24/16^{\circ}$ C ( $\delta = 4$ , the  $\delta 4$  group). These thermal scenarios represent predictions of climate change for central Chile (see http://www.dgf.uchile.cl/PRECIS/#Escenarios Climaticos).

To test for the effect of our thermal treatments, we measured SMR as carbon dioxide production ( $\dot{V}\mathrm{CO}_2$ ) at the end of the acclimation period (3 weeks) and within a closed system as previously described (Lighton, 2008). Three weeks after the beginning of temperature acclimation, we placed larvae individually inside glass syringes (2 mL) fitted with three-way valves and completely sealed for 4 h at a controlled temperature of 12, 20, 24, 28, 34, 38, 40 or 42°C (n=5 individuals per treatment). Then air from the syringe (1 mL) was slowly injected into a short Tygon tube connected to a glass tube (10 cm long) to avoid any loss before taking the  $\mathrm{CO}_2$  readings (Lighton, 2008). The ambient air injected was passed through a Drierite column to scrub the  $\mathrm{CO}_2$  before injection. Samples passed directly to the  $\mathrm{CO}_2$  analyser (Fox-Box, Sable system) with a 350 mL·min<sup>-1</sup> flow rate. We transformed the data from percentages to volume per hour. Then, using the program EXPEDATA (Sable Systems), we calculated the total  $\mathrm{CO}_2$  production per animal as the integral of the curve.

### Statistical analyses

We began by evaluating the effect of variability on three specific parameters. We performed a parametric curve fitting procedure of the SMR/temperature relationship under each scenario of thermal variability (Fig. 1). For each acclimation/thermal variability treatment, we fitted the relationship using a Gaussian three-parameter curve:

$$SMR = SMR_{\text{max}} \times e^{-0.5 \left(\frac{T_{\text{a}} - T_{\text{o}}}{SMR_{\text{b}}}\right)^{2}}$$

where  $SMR_{\rm max}$  is the maximum SMR value,  $SMR_{\rm b}$  is a measure of thermal performance breadth,  $T_{\rm a}$  is average environmental temperature, and  $T_{\rm o}$  is the optimal temperature, i.e. the temperature at which  $SMR_{\rm max}$  is attained. Fitting this curve to each treatment allowed us to compare the parameter values and evaluate the effect of the variability of the thermal acclimation regime on SMR.

Next, we obtained a global picture of the effect of variability. To obtain a global response function of SMR to temperature and the variability of the thermal regime, we performed a non-parametric regression analysis using a generalized additive model (GAM) incorporating both variables. Generalized additive models can detect non-linear relationships because they are piecewise. We determined the form of the partial functions related to each variable by fitting cubic regression splines to the data. We determined the complexity of the curve (the number of degrees of freedom) and the smoothing terms using penalized regression splines and generalized cross-validation (GCV) to avoid over-fitting (Wood, 2006). To evaluate the importance of thermal variability in the acclimation regime, we used cubic regression splines with shrinkage (Wood, 2006). This technique allows for extra penalty to be added in the model, and if the penalty is high enough, it shrinks all smoothing coefficients to zero. In this

case, the effective degrees of freedom of the variable(s) are so small that they can be removed and the model fitted again without them (Wood, 2006).

#### **RESULTS**

Estimated parameters in the parametric regression show that  $SMR_{max}$  and  $SMR_{b}$  were significantly different between treatments, but  $T_{o}$  was not. These results suggest that increases in thermal variability in the acclimation treatments caused a reduction in maximum SMR (Table 1a, Fig. 1a).

In addition, this pattern is established by the generalized additive model, which shows clearly how thermal variability reduces SMR at any level of temperature (Table 1b, Fig. 1b).

#### **DISCUSSION**

Although simple laboratory experiments, such as the one we report here, certainly cannot capture the wide range of conditions in nature, even simple experiments should better position us to understand the mechanisms by which climate change impacts organisms. Indeed, we believe important lessons emerge from our results.

Variability has an obvious detrimental effect on  $SMR_{max}$ . It decreases  $SMR_{max}$  by almost 50% between the  $\delta 0$  and  $\delta 8$  treatments. If the pattern of thermal variability changes in any of the directions predicted by Burroughs (2007) and Solomon *et al.* (2008), the performance of ectotherms will be severely impacted, thus modifying geographical distributions, population fluctuations patterns or predator—prey relationships (Estay *et al.*, 2012).

The thermal performance curves of Fig. 1(a) are a suitable descriptor of how a change in temperature influences physiological sensitivity and fitness among ectothermic species and populations (Huey *et al.*, 2012). Low and high temperatures and hence body temperature  $(T_b)$  reduce an ectotherm's performance and can be lethal. As observed here, thermal

**Table 1.** Estimated parameters of parametric and non-parametric regression: (a) Parameter values of parametric regression. Maximal standard metabolic rate or maximum performance ( $SMR_{max}$ ), metabolic or performance breadth ( $SMR_b$ ), and thermal optimum ( $T_o$ ) are indicated. Confidence intervals are in parentheses. (b) Generalized additive model (GAM) for the joint effect of average temperature (Temp) and thermal variability (Var).  $s_i$  represents the cubic regression spline for these variables and df is the effective degrees of freedom for each term

		Acclimation treatment (°C	)
	δ0	δ4	δ8
(a) $SMR_{max}$	1.89 (1.52–2.28)	1.44 (1.15–1.74)	0.93 (0.68–1.20)
$SMR_{\rm b}$	4.99 (4.07–6.28)	4.68 (3.78–5.97)	8.28 (5.76–16.57)
$T_{o}$	32.55 (31.50–33.60)	32.87 (31.80–34.00)	33.12 (30.74–37.34)
$T_{o}$ $R^{2}$ model	0.66	0.66	0.37
(b) GAM model:	Ln(SMR) = -1.46 + s Explained deviance.	$S_1$ (Temp, df = 1.99) + $S_2$ (Variable 1.99) + $S_2$ (Variable 1.99)	ar, $df = 1.14$ )

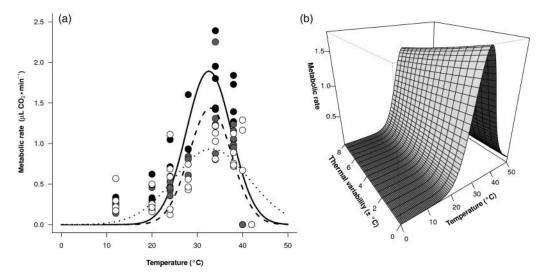


Fig. 1. Performance data fitted with both parametric and non-parametric models. (a) Parametric model fitted to each treatment for the relationship between temperature and metabolic rate. Lines represent fitted models; circles represent observed values. Symbols are:  $\delta 0$  treatment (solid line, black circles),  $\delta 4$  treatment (dashed line, grey circles), and  $\delta 8$  treatment (dotted line, white circles). (b) Generalized additive model fitted to performance data. We show the predicted response surface. Metabolic rate has the same units as (a).

performance curves shift to some extent depending on acclimation to thermal mean and variability.

Huey et al. (2012) note that the physiological impact of warming depends mainly on an ectotherm's  $T_{\rm b}$  (at the beginning of warming) relative to its optimal temperature. In fact, Huey and colleagues hypothesize that if pre-warming  $T_{\rm b}$  is not very close to optimal temperature ( $T_{\rm o}$ ), then warming-induced increases in  $T_{\rm b}$  will enhance performance. But if pre-warming  $T_{\rm b}$  is close to  $T_{\rm o}$ , a little warming will have a modest impact on performance. Nevertheless, if  $T_{\rm b}$  increases significantly above  $T_{\rm o}$ , warming should have lethal effects because performance quickly decreases when  $T_{\rm b} > T_{\rm o}$ . This kind of sensitivity to global warming may also change depending upon whether a species is a thermal generalist or specialist. In addition, recent models indicate that thermal variance could have as much (or more) of an impact on performance as mean temperature. Indeed, Bozinovic et al. (2011b) proposed that if the mean  $T_{\rm b}$  is below  $T_{\rm o}$  under variable thermal scenarios, brief warming will enhance physiological performance more than brief cooling will impair performance. In contrast, if the mean  $T_{\rm b}$  is close to  $T_{\rm o}$ , and thermal conditions are variable, both warming and cooling will impair performance. In general, the mean and variance of temperature should interact predictably to determine performance in a variable environment.

Although physiological and biophysical ecologists widely recognize the potential impacts of global warming, they have paid less attention to changes in thermal variation at a scale that pertains directly to organisms (Helmuth *et al.*, 2005; Lardies and Bozinovic, 2006; Folguera *et al.*, 2007). Also, the lack of a widely accepted theoretical basis to relate arthropod physiology to population processes makes it difficult to understand the role of thermal variability at higher ecological levels. Nevertheless, Lamb (1961) posits that, compared with constant temperatures, fluctuating temperatures have a distinct effect on insect development (see also

Dallwitz, 1984; Hilbeck and Kennedy, 1998; Hercus et al., 2003). For instance, Terblanche et al. (2010) and Folguera et al. (2011) showed that thermal variation enhanced the rate of population growth at a low mean temperature but depressed this rate at a high mean temperature. In addition, some species fail to acclimate their thermal sensitivity of performance during predictable or stochastic cooling (Schuler et al., 2011). Recently, Kjærsgaard et al. (2012) tested the effect of developmental temperature variability of wing traits on stressed locomotor performance in fruit flies, as well as its dependence on heterozygosity. They observed that locomotor performance declined in those flies reared under variable thermal conditions compared with those reared in thermal environments with the same mean. In addition, heterozygosity affected the flies' wing morphology. It would appear that environmental thermal mean and variability interact in a non-additive way to determine animal physiological performance. Thermal extremes – which can act as environmental cues – may also affect the performance curve parameters as observed here. Such changes in animal performance may also produce changes in activity patterns, and in the timing of breeding, and thus may ultimately alter synchronization between trophic levels and species community structure (Ashton et al., 2009).

Altogether, assessing the biodiversity consequences of climate change requires mechanistic and modelling approaches based on physiological and biophysical traits (Chown and Terblanche, 2006; Jentsch et al., 2007). In support of our belief, Huey et al. (2012) state that attempts to evaluate organismal vulnerability to global change and to develop management strategies should be based on relevant physiological foundations. Williams et al. (2008) developed a framework for assessing traits that promote vulnerability. In fact, the vulnerability of a species depends on its sensitivity to environmental change and variability, its exposure to change, its resilience to recover from ecological perturbations, and its potential to adapt to environmental changes and new scenarios. Physiological information at different hierarchical levels is fundamental here. Also, physiological plasticity may be important not only to deal with the increasing rate of environmental temperature increase, but also with the increasing unpredictability of climatic anomalies. In this vein, adaptive physiological acclimatization, in the context of global warming, may work as a buffer to minimize the coupling of environmental temperature and body temperature. It will be useful to know whether increased environmental variation has selected for increased physiological plasticity. Thus, new approaches linking physiological ecology and evolutionary ecology to climate change may produce important results at different scales. A major outcome will be to provide a solid, mechanistic foundation for predictions about the probability of a life form's success in the face of global change.

#### **ACKNOWLEDGEMENTS**

Funded by FONDECYT 1130015 to F.B. Thanks to S. Castillo and M. Barceló for technical assistance.

#### **REFERENCES**

Ashton, S., Gutierrez, D. and Wilson, R.J. 2009. Effects of temperature and elevation on habitat use by a rare mountain butterfly: implications for species responses to climate change. *Ecol. Entomol.*, **34**: 437–446.

Bannerman, J.A., Gillespie, D.R. and Roitber, B.D. 2011. The impacts of extreme and fluctuating temperatures on trait-mediated indirect aphid–parasitoid interactions. *Ecol. Entomol.*, **36**: 490–498.

- Bozinovic, F., Calosi, P. and Spicer, J.I. 2011a. Physiological correlates of geographic range in animals. *Annu. Rev. Ecol. Evol. Syst.*, **42**: 155–179.
- Bozinovic, F., Bastías, D.A., Boher, F., Clavijo-Baquet, S., Estay, S.A. and Angilletta, M.J. 2011b. The mean and variance of environmental temperature interact to determine physiological tolerance and fitness. *Physiol. Biochem. Zool.*, 84: 543–552.
- Burroughs, W.J. 2007. Climate Change: A Multidisciplinary Approach. Cambridge: Cambridge University Press.
- Catalán, T.P., Niemeyer, H.M., Kalergis, A.M. and Bozinovic, F. 2012a. Interplay between behavioural thermoregulation and immune response in mealworms. J. Insect Physiol., 58: 1450–1455
- Catalán, T.P., Wozniak, A., Niemeyer, H.M., Kalergis, A.M. and Bozinovic, F. 2012b. Interplay between thermal and immune ecology: effect of environmental temperature on insect immune response and energetic costs after an immune challenge. *J. Insect Physiol.*, **58**: 1450–1455.
- Chown, S.L. and Terblanche, J.S. 2006. Physiological diversity in insects: ecological and evolutionary context. *Adv. Insect Physiol.*, **33**: 50–152.
- Coumou, D. and Rahmstorf, S. 2012. A decade of weather extremes. *Nature Climate Change*, **2**: 491–496.
- Dallwitz, R. 1984. The influence of constant and fluctuating temperatures on development rate and survival of pupae of the Australian blowfly *Lucilia cuprina*. *Entomol. Exp. Appl.*, **36**: 89–95.
- Dawson, T.P., Jackson, S.T., House, J.I., Prentice, I.C. and Mace, G.M. 2011. Beyond predictions: biodiversity conservation in a changing climate. *Science*, **332**: 53–58.
- DeVries, Z.C. and Appel, A.G. 2013. Standard metabolic rates of *Lepsima saccharina* and *Thermobia domestica*: effects of temperature and mass. *J. Insect Physiol.*, **59**: 638–645.
- Easterling, D.R., Meehl, G.A., Parmesan, C., Changnon, S.A., Karl, T.R. and Mearns, L.O. 2000. Climate extremes: observations, modeling, and impacts. *Science*, **289**: 2068–2074.
- Estay, S.A., Clavijo-Baquet, S., Lima, M. and Bozinovic, F. 2011. Beyond average: an experimental test of temperature variability on the population dynamics of *Tribolium confusum. Pop. Ecol.*, **53**: 53–58.
- Estay, S.A., Lima, M., Labra, F. and Harrington, R. 2012. Increased outbreak frequency associated with changes in the dynamic behaviour of populations of two aphid species. *Oikos*, **121**: 614–622.
- Folguera, G., Muñoz, J.L.P., Ceballos, S., Bozinovic, F. and Hasson, E. 2007. Testing the interplay between physiological and life history traits: an experimental study in *Drosophila. Evol. Ecol. Res.*, 9: 1211–1222.
- Folguera, G., Bastías, D.A. and Bozinovic, F. 2009. Impact of experimental thermal amplitude on ectotherm performance: adaptation to climate change variability? *Comp. Biochem. Physiol. A: Molec. Integr. Physiol.*, **154**: 389–393.
- Folguera, G., Bastías, D.A., Caers, J., Rojas, J.M., Piulachs, M.D., Bellés, X. *et al.* 2011. An experimental test of the role of environmental temperature variability on molecular, physiological and life-history traits: implications for global warming. *Comp. Biochem. Physiol. A: Molec. Integr. Physiol.*, **159**: 242–246.
- Helmuth, B., Kingsolver, J.G. and Carrington, L. 2005. Biophysics, physiological ecology and climate change: does mechanism matter? *Annu. Rev. Physiol.*, **67**: 177–201.
- Hercus, M.J., Loeschcke, V. and Rattan, S.I.S. 2003. Lifespan extension of *Drosophila melanogaster* through hormesis by repeated mild heat stress. *Biogerontology*, **4**: 149–156.
- Hilbeck, A. and Kennedy, G.G. 1998. Effects of temperature on survival and preimaginal development rates of Colorado potato beetle on potato and horse-nettle: potential role in host range expansion. *Entomol. Exp. Appl.*, **89**: 261–269.
- Huey, R.B., Kearney, M.R. and Krockenberger, A. 2012. Predicting organismal vulnerability to climate warming: roles of behaviour, physiology and adaptation. *Phil. Trans. R. Soc. Lond. B*, **367**: 1665–1679.

- Intergovernmental Panel on Climate Change (IPCC). 2007. Climate Change 2007: The Physical Science Basis. Summary for Policymakers. Available at: http://www.ipcc.ch/SPM2feb07.pdf.
- Jentsch, A., Kreyling, J. and Beierkuhnlein, C. 2007. A new generation of climate change experiments: events, not trends. Front. Ecol. Environ., 5: 315–324.
- Ji, X., Gao, J. and Han, J. 2007. Phenotypic responses of hatchlings to constant versus fluctuate 7ng incubation temperatures in the multi-banded krait, *Bungarus multicintus* (Elapidae). *Zool. Sci.*, 24: 384–390.
- Kjærsgaard, A., Nguyet, L., Demontis, D., Novicic, Z.K., Loeschcke, V. and Pertoldi, C. 2012. The effect of developmental temperature fluctuation on wing traits and stressed locomotor performance in *Drosophila melanogaster*, and its dependence on heterozygosity. *Evol. Ecol. Res.*, 14: 803–819.
- Krams, I., Daukste, J., Kivleniece, I., Krama, T. and Rantala, M.J. 2011. Overwintering survival on immune defence and body length in male *Aquarius najas* water striders. *Entomol. Exp. Appl.*, **140**: 45–51.
- Lalouette, L., Kostál, V., Colinet, H., Gagneul, D. and Renault, D. 2007. Cold exposure and associated metabolic changes in adult tropical beetles exposed to fluctuating thermal regimes. *FEBS J.*, **274**: 1759–1767.
- Lamb, K.P. 1961. Some effects of fluctuating temperatures on metabolism, development and rate of population growth in the cabbage aphid, *Brevicoryne brassicae*. *Ecology*, **42**: 740–745.
- Lardies, M.A. and Bozinovic, F. 2006. Geographic covariation between metabolic rate and life history traits. Evol. Ecol. Res., 8: 455–470.
- Lean, J.L. and Rind, D.H. 2008. How natural and anthropogenic influences alter global and regional surface temperatures: 1889 to 2006. *Geophys. Res. Lett.*, **35**: 1–6.
- Lighton, J.R.B. 2008. Measuring Metabolic Rates. Oxford: Oxford University Press.
- Miller, N.A. and Stillman, J.H. 2012. Physiological optima and critical limits. *Nature Education Knowledge*, **3**(5): 1.
- Min, K.J., Hogan, M.F., Tatar, M. and O'Brian, D.M. 2006. Resource allocation to reproduction and soma in *Drosophila*: a stable isotope analysis of carbon from dietary sugar. *J. Insect Physiol.*, **52**: 63–70.
- Morales-Ramos, J.A., Rojas, M.G., Shapiro-Ilan, D.I. and Tedders, W.L. 2010. Developmental plasticity in *Tenebrio molitor* (Coleoptera: Tenebrionidae): analysis of instar variation in number and developmental time under different diets. *J. Entomol. Sci.*, **45**: 75–90.
- Orcutt, J.D. and Porter, K.G. 1983. Diel vertical migration by zooplankton: constant and fluctuating temperature effects on life history parameters of *Daphnia. Limnol. Oceanogr.*, **28**: 720–730.
- Paaijmans, K.P., Blanford, S., Bell, A.S., Blanford, J.I., Read, A.F. and Thomas, M.B. 2010. Influence of climate on malaria transmission depends on daily temperature variation. *Proc. Natl. Acad. Sci. USA*, **107**: 15135–15139.
- Parmesan, C., Root, T.L. and Willig, M.R. 2000. Impacts of extreme weather and climate on terrestrial biota. *Bull. Am. Met. Soc.*, **81**: 443–450.
- Pétavy, G., David, J.R., Debat, V., Gibert, P. and Moreteau, B. 2004. Specific effects of cycling stressful temperatures upon phenotypic and genetic variability of size traits in *Drosophila melanogaster. Evol. Ecol. Res.*, **6**: 873–890.
- Pörtner, H.O., Bennett, A.F., Bozinovic, F., Clarke, A., Lardies, M.A., Lenski, R.E. *et al.* 2006. Trade-offs in thermal adaptation: in need of a molecular to ecological integration. *Physiol. Biochem. Zool.*, **79**: 295–313.
- Ragland, G.J. and Kingsolver, J.G. 2008. The effect of fluctuating temperatures on ectotherm life-history traits: comparisons among geographic populations of *Wyeomyia smithii. Evol. Ecol. Res.*. **10**: 29–44.
- Rahmstorf, S. and Coumou, D. 2011. Increase of extreme events in a warming world. *Proc. Natl. Acad. Sci. USA*, **108**: 17905–17909.

- Schuler, M.S., Cooper, B.S., Storm, J.J., Sears, M.W. and Angilletta, M.J. 2011. Isopods failed to acclimate their thermal sensitivity of locomotor performance during predictable or stochastic cooling. *PLoS One*, **6**: e20905
- Solomon, S., Qin, D., Manning, M., Chen, Z., Marquis, M., Averyt, K.B. et al. 2008. Climate Change 2007: The Physical Science Basis. Cambridge: Cambridge University Press.
- Somero, G.N. 2011 Comparative physiology: a 'crystal ball' for predicting consequences of global change. *Am. J. Physiol.: Regul. Integr. Comp. Physiol.*, **302**: R1–R14.
- Terblanche, J.S., Clusella-Trullas, S., Deere, J.A. and Chown, S.L. 2008. Thermal tolerance in a south-east African population of tsetse fly *Glossina pallidipes* (Diptera, Glossinidae): implications for forecasting climate change impacts. *J. Insect Physiol.*, **54**: 114–127.
- Terblanche, J.S., Nyamukoniwa, C. and Elsje, K. 2010. Thermal variability alters climatic stress resistance and plastic response in a global invasive pest, the Mediterranean fruit fly (*Ceratitis capitata*). *Entomol. Exp. Appl.*, **137**: 304–315.
- Thompson, R.M., Beradall, J., Beringer, J., Grace, M. and Sardina, P. 2013. Means and extremes: building variability into community-level climate change experiments. *Ecol. Lett.*, **16**: 799–806.
- Williams, C.M., Marshall, K.E., MacMillan, H.A., Dzursin, J.D.K., Hellmann, J.J. *et al.* 2012. Thermal variability increases the impact of autumnal warming and drives metabolic depression in an overwintering butterfly. *PLoS One*, 7: e34470.
- Williams, S.E., Shoo, L.P., Isaac, J.L., Hoffmann, A.A. and Langham, G. 2008. Towards an integrated framework for assessing the vulnerability of species to climate change. *PLoS Biol.*, 6: 2621–2626.
- Wood, S.N. 2006. Generalized Additive Models: An Introduction with R. Boca Raton, FL: Chapman & Hall/CRC.