



UNIVERSIDAD DE CHILE-FACULTAD DE CIENCIAS-ESCUELA DE PREGRADO

“¿Qué está desaparecido, poblaciones o una especie? Antiguas grabaciones de llamadas revelan similitud entre las dos presuntas especies de Ranita de Darwin”

Seminario de Título entregado a la Universidad de Chile en cumplimiento parcial de los requisitos para optar al Título de: **Biólogo con Mención en Medio Ambiente**

Gabriel Ignacio Bidart Enriquez

Director del Seminario de Título: José Manuel Serrano

Codirector: Mario Penna

Patrocinante: Claudio Veloso

Fecha: 2 de marzo del 2021

Santiago – Chile



INFORME DE APROBACIÓN SEMINARIO DE TITULO

Se informa a la Escuela de Pregrado de la Facultad de Ciencias, de la Universidad de Chile
que el Seminario de Título, presentado por el Sr:

Gabriel Bidart Enriquez.

**“¿Qué está desaparecido, poblaciones o una especie? Antiguas grabaciones
de llamadas revelan similitud entre las dos presuntas especies de Ranita de
Darwin”**

Ha sido aprobado por la Comisión de Evaluación, en cumplimiento parcial de los requisitos
para optar al Título de Biologo con Mencion en Medio Ambiente

Director Seminario de Título: Dr. Jose Manuel Serrano

Cotutor Seminario de Titulo: Dr. Mario Penna

Comisión Revisora y Evaluadora

Presidente Comisión: Dr. Marco Mendez

Evaluador: Dr. Javier Simonetti

Santiago de Chile,

Agradecimientos

Agradezco en primer lugar al Dr. Jose Manuel Serrano por brindarme su compromiso, apoyo y sabiduría y por ayudar a reencontrarme con las ciencias biológicas e iniciarme en el estudio de los anfibios chilenos, dándome el privilegio de acompañarle en maravillosos terrenos por Chile para estudiar a las Ranitas de Darwin. Junto con él, agradezco también al Dr. Mario Penna por aceptarme en el laboratorio de Neuroetología y por su permanente disposición a ayudar y educar de manera humilde y desinteresada y al Dr. Nelson Velasquez por iniciarme en el camino de la Bioacústica. A todos y cada uno de los integrantes del laboratorio por ayudarme y aconsejarme durante todo el proceso de tesis, en especial a Claudio Reyes, cuyo constante apoyo tanto científico como de camaradería fue una luz al final del túnel siniestro de los análisis estadísticos en R.

A mi familia por quererme y aguantar mi difícil carácter, el cual se vio acentuado por la crisis sanitaria y social que enfrentamos este 2020, pero en especial a Patricia Enriquez Andueza, mi madre, la cual a pesar de ser madre viuda logró erigirse como un sólido pilar en el cual todos pudimos apoyarnos. A mi compañera de vida Camila Mondaca, cuya expertis en las letras y compañía fueron indispensables para terminar mi tesis y el respectivo artículo. A mis amigos de la vida y la universidad, con los cuales nos apoyamos mutuamente para no caer en la locura en tiempos de pandemia y crisis social, pero en especial a Dalila Briones por su infinita disposición a ayudarme con todos esos temas administrativos, burocráticos y de formatos de la universidad con los que sufrí tanto durante la carrera y la tesis.

A Stiftung Artenschutz 2018 por financiar la digitalización de las cintas magnéticas de las cuales se obtuvieron parte de las grabaciones analizadas en este estudio.

Y, por último pero no menos importante, un agradecimiento especial a mi difunto padre Álvaro Torrealba, cuya sabiduría y amor infinitos fueron los cimientos más fundamentales de mi carrera en las ciencias y en la vida.

Índice de contenidos

Agradecimientos.....	3
Resumen	7
Abstract	10
Introduction	11
Materials and Methods	16
Study site and general procedures	16
Acoustic recordings.....	16
Data analysis.....	17
Results	18
Acoustic differences between species	20
Distinctiveness analysis for all populations	21
Distinctiveness analysis for mainland populations.....	23
Discussion	24
Geographic variation of Rhinoderma advertisement calls and body size.....	24
Variation in other characters in Rhinoderma.....	27
Conservation implications	29
Acknowledgements	31
References	31
Supplementary information.....	47

Índice de figuras

Figure 1. Geographic distribution of the two <i>Rhinoderma</i> species according to the UICN red list.....	14
Figure 2. Spectrograms and oscillograms of representative advertisement calls of <i>Rhinoderma</i>	19
Figure 3. Proportions of number of notes in the calls of each population of <i>Rhinoderma</i>	21
Figure 4. Discriminant analysis of call characteristics of individuals of <i>R. darwinii</i> and <i>R. rufum</i>	23

Índice de tablas

Table 1. Average values and coefficients of variation (in cursive) of the acoustic variables from <i>Rhinoderma rufum</i> and <i>Rhinoderma darwinii</i> populations.....	20
Table 2. Loadings of the acoustic variables, eigenvalues and explained variances for each principal component (PC), resulting from the analysis including the four populations studied.....	22
Table 3. Loadings of the acoustic variables, eigenvalues and explained variances for each principal component (PC), resulting from the analysis excluding the insular population of Tantauco.. ..	24
Table S 1. Averages and coefficients of intra-individual variation (in cursive) of the acoustic variables analysed in the advertisement calls of <i>Rhinoderma rufum</i> from Las Carmelitas	47

1 **¿Qué está desaparecido, poblaciones o una especie? Antiguas grabaciones de llamadas**
2 **revelan similitud entre las dos presuntas especies de Ranita de Darwin**

3 **Resumen**

4 El género *Rhinoderma* tiene dos especies putativas, la Ranita de Darwin del norte y del sur
5 que tiene una simpatría histórica entre 36 y 27° S en Chile central. La identidad de las dos
6 taxones se basa en estudios que muestran variación en caracteres morfológicos, evolutivos,
7 cariológicos y moleculares. La Ranita de Darwin del norte no se ha avistado desde 1981. Las
8 señales acústicas de atracción de pareja están sujetas a la variación geográfica y son
9 herramientas importantes para aclarar estatus sistemáticos entre especies, dado que facilitan
10 barreras reproductivas entre especies. Para investigar la diferenciación acústica entre las
11 Ranitas de Darwin, hemos analizado grabaciones antiguas de llamadas de anuncio de Ranitas
12 de Darwin del norte, un taxón actualmente perdido, junto con más grabaciones recientes de
13 tres poblaciones de la Ranita de Darwin del sur. Combinamos pruebas GLM y análisis
14 discriminante para evaluar las diferencias de la Ranita de Darwin del norte y la formación de
15 conjuntos acústicos entre todas las poblaciones de las dos presuntas especies. El análisis
16 mostró diferencias limitadas en las llamadas de la especie del norte en comparación con las
17 del sur que se relacionan, probablemente, con las disimilitudes en las condiciones climáticas y
18 el tamaño del cuerpo entre las poblaciones. El análisis discriminante arrojó dos grupos
19 separados, uno para la población de Tantauco de la *R. darwinii* de la Isla de Chiloé y otro para
20 las poblaciones continentales que incluyeron Las Carmelitas, cerca de la ciudad de
21 Concepción (*R. rufum*), Huilo Huilo y Quelulat (*R. dariwini*). Luego de repetir este análisis
22 excluyendo a la población de la isla, las poblaciones continentales de la Ranita de Darwin del
23 norte y del sur permanecieron en un conjunto único. Proponemos que la divergencia entre
24 poblaciones de la Ranita de Darwin relacionada a los rasgos acústicos informados en este
25 estudio y otros caracteres informados en estudios previos se debe a una variación geográfica

26 que depende de regímenes climáticos a lo largo del rango histórico de *Rhinoderma*. Las
27 medidas de conservación recomendadas por la Estrategia Binacional de Conservación de las
28 Ranitas de Darwin debiesen considerar una falta potencial de diferenciación específica a lo
29 largo de la distribución geográfica de las poblaciones de la Ranita de Darwin en Chile.

30 **Research Article - Animal Conservation**

31 **Is it a species or populations that are missing? vintage call recordings reveal similarity**
32 **between the two alleged species of Darwin's frogs**

33 Gabriel Bidart-Enriquez^{1,2}, José M. Serrano^{3,4,*}, Mario Penna², Nelson Velásquez⁵

34 ¹Facultad de Ciencias, Universidad de Chile, Las Palmeras 3425, Ñuñoa, Santiago, Chile

35 ²Programa de Fisiología y Biofísica, Facultad de Medicina, Universidad de Chile,

36 Independencia 1027, 8380453, Santiago, Chile.

37 ³ONG Ranita de Darwin, Santiago, Chile.

38 ⁴Centro de Investigación de Estudios Avanzados del Maule, Universidad Católica del Maule,

39 San Miguel 3605, 3480112, Talca, Chile.

40 ⁵Laboratorio de Comunicación Animal, Facultad de Ciencias Básicas, Universidad Católica

41 del Maule, San Miguel 3605, 3480112, Talca, Chile.

42 *Corresponding author email: jose.rano@gmail.com

43 **Abstract**

44 The genus *Rhinoderma* comprises two putative species, the northern and southern Darwin's
45 frogs that had a historical sympatric area between 36 and 37 ° S in central Chile. The identity
46 of the two taxa is based in studies showing variation in morphological, developmental,
47 karyological and molecular characters. The northern Darwin's frog has not been seen since
48 1981. Mate attracting acoustic signals are subjected to geographical variation and are
49 important tools to resolve systematic status between species as these signals provide
50 reproductive barriers between species. To investigate acoustic differentiation in Darwin's
51 frogs we analysed vintage recordings of the advertisement calls of northern Darwin's frogs,
52 currently a missing taxon, together with more recent recordings of three populations of
53 southern Darwin's frogs. We combined GLM tests and discriminant analysis to assess the
54 differences of the northern Darwin's frog and the formation of acoustic clusters between all
55 populations of the two presumed species. The analysis showed limited differences in the calls
56 of the northern species compared to the southern species that are probably related to
57 dissimilarities in climate conditions and body size between populations. The discriminant
58 analysis yielded two separate clusters, one for the Tantauco population of *R. darwinii* on the
59 island of Chiloé and another for populations from the mainland that included Las Carmelitas,
60 near the city of Concepción (*R. rufum*) Huilo Huilo and Queulat (*R. darwinii*). After
61 repeating this analysis excluding the island population, the continental populations of the
62 northern and southern Darwin's frogs remained in a single cluster. We propose that the
63 divergence between populations of Darwin's frogs in acoustic traits reported in this study and
64 other characters reported in previous studies is due to geographical variation? dependent on
65 climate regimes along the historical range of *Rhinoderma*. The conservation measures
66 recommended by the Binational Conservation Strategy for Darwin's Frogs should account for

67 a potential lack of specific differentiation along the historical geographic distribution of
68 Darwin's frog populations in Chile.

69 **Keywords:** advertisement call; geographic variation; species delimitation; *Rhinoderma*;
70 integrative taxonomy.

71 **Introduction**

72 Acoustic signals of animals can reveal insights on their evolution, providing relevant tools for
73 promoting their conservation (reviewed in Laiolo, 2010). Species producing vocal signals
74 usually have particular characteristics that allow recognition by conspecifics (Tyack, 2000;
75 Feng et al., 2009; Araya-Salas et al., 2019). These signals are produced in different contexts
76 such as mate attraction, territorial defence or in response to stressful situations (Obrist et al.,
77 2010; Bradbury & Vehrencamp, 2011). In mate attraction contexts, the signal structure is
78 affected by coevolutionary processes between sender and receiver, promoting species-specific
79 communication (e.g. Tang et al., 2001; Wolley, 2011).

80 As such, different patterns of variation of acoustic signals can occur depending on the
81 geographic distance between populations and the identity of the species involved. For
82 instance, related species living in areas that have been apart for long time spans can show
83 divergence between populations that depends on the geographic distance among localities
84 (Nevo & Capranica, 1985; Wilczynski & Ryan, 1999; Castellano et al., 2000; Giacoma and
85 Castellano, 2001; Bernal et al., 2005; Prohl et al., 2006, 2007; Weaver et al., 2020). However,
86 a contrasting condition corresponds to allopatric populations having limited differentiation in
87 their acoustic signals, for which a specific status can be established by genetic evidence only
88 (Heyer et al., 2005; Greenbaum et al., 2013). Among anurans, morphologically cryptic
89 species occur frequently, but combined genetic and bioacoustic evidence has allowed to

90 detect unnoticed diversity among sympatric species (e.g., Funk et al., 2012) or to merge taxa
91 formerly considered as morphologically distinct species (e.g., Kuramoto, 1997).

92 Reproductive signals provide an isolation mechanism to prevent hybridization among
93 related sympatric species (Claridge, 1990; Freeman & Montgomery, 2017; Weaver et al.,
94 2020). As such, sexual signals contribute to evolutionary processes supporting species
95 recognition and delimitation (Mace, 2004; Padial et al., 2010; Cardoso et al., 2012;
96 Tishechkin & Vedenina, 2016; Bolaños-Sittler et al., 2020; Escalona et al., 2019). Therefore,
97 assessing how acoustic variables change along geographic distribution of a species is relevant
98 to understand the role of these traits in divergence and adaptation processes, and they are
99 useful as taxonomical tools (Köhler et al., 2017; Miller et al., 2020), and their pertinence for
100 conservation initiatives has been acknowledged (Laiolo, 2010).

101 To relate different acoustic variables with divergence processes of anurans, Gerhardt
102 (1991) proposed a classification along the continuum of variation of acoustic traits, using a
103 predetermined value of the coefficient of variation as a threshold to identify these variables as
104 static or dynamic. The two types of variables are likely to evolve under different pressures as
105 static traits (those in which the coefficient of variation is lower than 5%) are relevant for
106 species recognition whereas dynamic traits (those in which the coefficient of variation is
107 higher than 5%) are related to social interactions. Typically, dominant frequency and pulse
108 rate are static variables, whereas call rate and call duration are dynamic variables (Gerhardt,
109 1991; Castellano & Giacoma, 1998; Zimmitti, 1999; Castellano et al., 2000, 2002; Friedl,
110 2006; Reinhold, 2011, Köhler et al., 2017).

111 The number of notes in particular is considered a dynamic variable because it is
112 largely under behavioural and physiological control (Jehle & Arak, 1998; Gerhardt et al.,
113 2000). In some species, more or less notes in the repertoire may be associated with a
114 communication system that indicates territoriality (Toledo et al., 2015). But this diversity of

115 sequences differing in the number of notes may also reflect divergence in the call repertoires
116 (Höbel & Fellows, 2016; Serrano, 2018; Weaver et al., 2020). In contrast, dominant
117 frequency is a typical static acoustic variable because it is usually a species-specific trait that
118 can vary with body size both at the intra-population (Gerhardt & Huber, 2002; Gingras et al.,
119 2013) and inter-specific level (Penna & Veloso, 1990; Bosch and de la Riva, 2004; Serrano et
120 al., 2020).

121 *Rhinoderma* is an anuran genus endemic to the South American Patagonia,
122 comprising the species *Rhinoderma darwinii* and *Rhinoderma rufum*, commonly known as
123 southern and northern Darwin's frogs, respectively (Azat et al., 2020). The historical
124 distribution of *R. rufum* was restricted to small and isolated populations located mainly along
125 the coastal mountains of Chile, from Zapallar (32°33' S) to the south of the city of
126 Concepción (36°45' S) (Cuevas, 2014), while the distribution of *R. darwinii* ranges from
127 Concepción to Aysen (46°44' S) in Chile and also distributes in Argentina (Azat et al., 2020).
128 Interestingly, a sympatric zone of the two *Rhinoderma* species occurred in the region of
129 Concepción and Nahuelbuta mountains (37°47' S) (Bourke et al., 2012; Soto-Azat et al.,
130 2013; Fig. 1). Despite of its extended historical distribution, *R. darwinii* is currently in a high
131 risk of extinction due to its fragmented occurrence and populations decrease (Valenzuela-
132 Sánchez et al., 2017), while *R. rufum* is apparently extinct since 1981, as recent efforts to find
133 populations have been unsuccessful (Cuevas, 2014; Bourke, 2012; Soto-Azat et al., 2013;
134 IUCN SSC Amphibian Specialist Group, 2015).

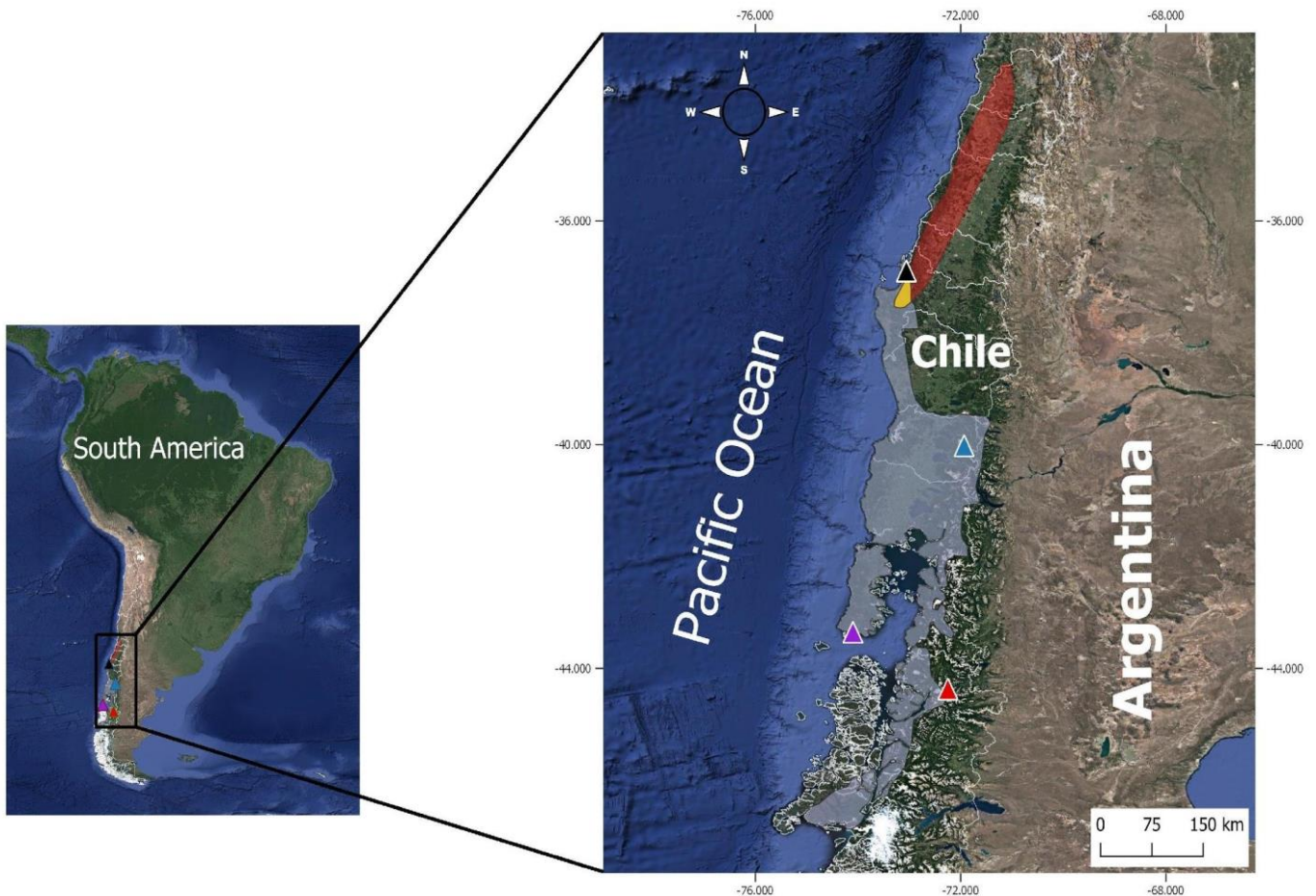


Figure 1. Geographic distribution of the two *Rhinoderma* species according to the UICN red list. The historical distribution of *R. rufum* and *R. darwinii* are depicted in red and grey, respectively, and the area of sympatry of the two species is in yellow. The population where the advertisement calls of *R. rufum* were recorded is indicated by a black triangle (Las Carmelitas) and the populations of *R. darwinii* are indicated by blue, red and purple triangles (Huilo Huilo, Queulat and Tantauco, respectively).

135 The two species of *Rhinoderma* differ in morphological features, including interdigital
 136 membranes of the hindlimbs, parental care phenology in adults and mouth and digestive tract
 137 structures in tadpoles (Formas et al., 1975; Jorquera et al., 1981, 1982; Formas, 2013).
 138 Furthermore, isoenzyme and chromosomal differences contribute to place the two species
 139 apart (Formas, 1976; Díaz & Veloso, 1979; Díaz, 1986). In addition, Formas et al (1975)
 140 describe qualitative call differences not supported by recordings between the two species, and
 141 Penna & Veloso (1990) contributed a preliminary analysis of the calls of *R. darwinii* and *R.*
 142 *rufum* indicating species distinctiveness. However, that description of *Rhinoderma*

143 advertisement calls was based on advertisement calls of just four and eight individuals of *R.*
144 *darwinii* and *R. rufum*, respectively. Differences in number of notes, notes and call duration
145 and dominant frequency were reported, but no statistical analyses were applied. One
146 additional evidence on vocal divergence between the two species is the production of release
147 calls by *R. rufum* males, a vocalization that is apparently absent in the congeneric species
148 (Serrano et al., accepted). In this context, to define the taxonomic delimitation of *Rhinoderma*
149 is priority for the current Binational Conservation Strategy of the Darwin's Frogs (Azat et al.,
150 2020) that considers *R. darwinii* and *R. rufum* as two units under different negative pressures,
151 and for which different conservation measures are planned.

152 The main purpose of this study is to compare the acoustic properties of the
153 advertisement calls of *R. rufum* and *R. darwinii* to assess potential divergence between both
154 species. We carried out a detailed description of the advertisement calls of *R. rufum* recorded
155 in 1980 (Penna & Veloso, 1990) and contrasted these data with large samples of
156 advertisement calls from three *R. darwinii* populations recorded recently (Serrano et al.,
157 2020). We analysed distinctiveness of the advertisement calls of the two species addressing
158 the hypothesis that they correspond to distinct taxonomic entities, in agreement with evidence
159 reported in comparative developmental, biochemical and karyological studies. Alternatively,
160 a lack of distinctiveness in the advertisement calls between the two species would contribute
161 to assess their similarity within *Rhinoderma*. Our study is also meant to provide relevant
162 information to support contemporary conservation efforts of these endangered or missing
163 taxa.

164 **Materials and Methods**

165 *Study site and general procedures*

166 The advertisement calls of *R. rufum* were recorded by Mario Penna during spring (October
167 1980) in a fragment of native forest located in the vicinity of Las Carmelitas convent,
168 approximately 10 km south from the city of Concepción, Chile (36°53'39.48'' S
169 73°3'37.349'' W; Fig 1). Advertisement calls of nine individuals (presumably males) were
170 recorded during day and night times. No individuals were captured and the air temperature
171 during recordings was 14-18 °C, but temperatures for recordings of individual recordings
172 were not registered. These recordings were partially reported in an early study on the
173 description of vocalizations of anurans from the temperate austral forest (Penna & Veloso,
174 1990). Advertisement calls of *R. darwinii* were recorded in three localities during spring
175 along the distribution of this frog (Huilo Huilo, n=12, Tantauco, n=13 and Queulat, n=12)
176 between 2011 and 2014 and their signal variation patterns have been reported recently
177 (Serrano et al., 2020; Fig. 1).

178 *Acoustic recordings*

179 Advertisement calls of *R. rufum* were recorded with an omnidirectional microphone (Uher
180 M517) and a reel tape recorder (Uher 4400 Report Stereo IC) at a tape speed of 19 cm/s. The
181 reel tapes were digitized in WAV format using the analogue-digital converter tools of
182 Sequoia 14 (Magix Software GmbH, Berlin, Germany) at a sampling rate of 44,100 Hz and
183 24-bit resolution. The calls were analysed with Raven PRO v1.5 software. Five acoustic
184 variables were analysed in oscillograms: call duration (s), note duration (ms), inter-note
185 interval (ms), notes per call and call rate (calls/min); and two variables were obtained from
186 spectrograms in which we measured the dominant frequency (Hz) and the proportion of note

187 duration segments containing the non-linear phenomenon of chaos. Figures of sound
188 spectrograms and oscillograms were obtained using the Seewave package (Sueur et al., 2019).

189 *Data analysis*

190 A data matrix containing the average values of calls of the *R. rufum* population and three
191 populations of *R. darwinii* was constructed. We calculated coefficients of variation (CV) of
192 advertisement calls within populations and within individuals of the four *Rhinoderma*
193 populations as a measure of the intra-population and intra-individual variation of the acoustic
194 characteristics.

195 A Generalized Linear Model (GLM) was constructed to assess differences between
196 the advertisement calls of *R. rufum* (at Las Carmelitas) and the three populations of *R.*
197 *darwinii* studied, using acoustic variables as dependent variables and populations as
198 predictors. To perform all the statistical analysis properly, the data matrix was analysed for
199 normality using the Mardia test for multivariate normality (Korkmaz, 2014). Call variability
200 corresponding to different numbers of notes per call, was analysed separately using a
201 Kolmogorov-Smirnov paired test (to maximize the test power as concluded by Böhm &
202 Hornik, 2012) to explore inter-population differences.

203 A linear discrimination analysis (LDA) was performed using the principal
204 components (PC) obtained from a principal component analysis (PCA) in order to study if the
205 acoustic variables discriminate the populations (Liang et al., 2010; Bolaños-Sittler et al.,
206 2020). The PCA was performed using a correlation matrix and a linear combination (z
207 transformation) to deal with non-normalities in some of the acoustic variables. The PCs
208 reaching above 80% of the cumulative explained variance (Jackson, 1993) were used as the
209 entry data into the LDA. This procedure has the advantage of reducing the effect of the great
210 variance of some variables that the LDA could mistake for observed variability, discard
211 collinearity effects and eliminate the “noise” that variables such as dominant frequency could

212 cause by having much higher values than the other analysed variables (Mardia et al, 1979;
213 Næs & Mevik, 2001) and has been employed in other evaluations of acoustic data (Funk et
214 al., 2012; Bolaños-Sittler et al., 2020). Finally, to validate the discriminant model, the same
215 individuals of all populations were reclassified using the linear model generated by the LDA.
216 For this, the probability of pertaining to a population was calculated for each individual
217 according to the model predictors; we a priori considered as correctly classified the
218 individuals that reached a belonging percentage higher than 50% relative to their original
219 locality. All the analyses were performed with the packages MVN (Korkmaz et al., 2014),
220 ade4 (Dray et al., 2016), ggplot2 (Wickham et al., 2016) and MASS (Ripley et al., 2013) to R
221 (R Core Team, 2019).

222 **Results**

223 The advertisement calls of *R. darwinii* and *R. rufum* are whistle-like sounds composed by two
224 to six notes.

225 Notes usually show a slight amplitude and frequency modulation, however in some
226 individuals these modulations can be pronounced. Non-linear phenomena, namely chaos
227 occurred preferentially at the beginning or end of the notes (Fig. 2).

228 In the continuum of variation of vocal traits proposed by Gerhardt (1991), we found
229 that in *R. rufum* dominant frequency had the lowest within population CV (below 5%) (Table
230 1), therefore this was considered as a static variable. Call duration, note duration, chaos
231 proportion, inter-note interval duration and call rate had higher CV's (from 17% to 59.1%)
232 and therefore were considered as dynamic variables (Table 1, for inter-individual CVs see
233 Table S1).

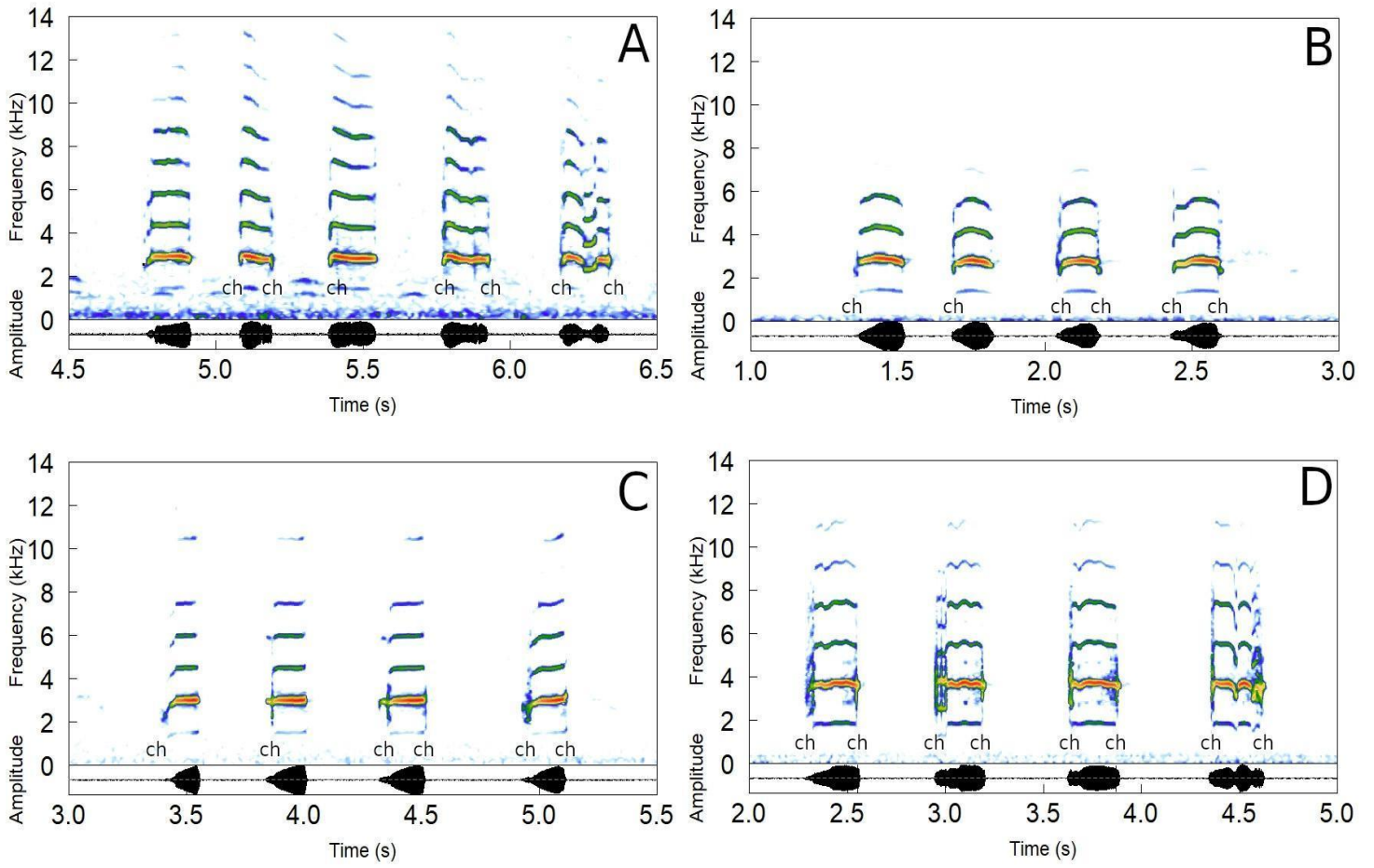


Figure 2. Spectrograms and oscillograms of representative advertisement calls of *Rhinoderma rufum* recorded in Las Carmelitas (A) and *Rhinoderma darwinii* recorded in Huilo Huilo (B), Queulat (C) and Tantauco (D). Chaos (ch) segments are indicated in spectrograms.

Table 1. Average values and coefficients of variation (in cursive) of the acoustic variables in *Rhinoderma rufum* and *Rhinoderma darwinii* populations. Asterisks indicate significance levels of GLMs resulting from comparisons between *R. rufum* population of Las Carmelitas versus each of the populations of *R. darwinii* (*: $p < 0.05$, ***: $p < 0.001$).

Species and Populations	Call rate (Calls / min)	Call duration (s)	Note duration (s)	Inter-note interval (s)	Chaos proportion (%)	Dominant frequency (Hz)
<i>R. rufum</i> Las Carmelitas (n= 9)	1.479± 0.64 <i>43.0</i>	1.5 ± 0.46 <i>30.6</i>	0.17 ± 0.03 <i>17.0</i>	0.19 ± 0.05 <i>24.5</i>	27.8 ± 16.45 <i>59.1</i>	2946 ± 113.53 <i>3.9</i>
<i>R. darwinii</i> Huilo Huilo (n= 12)	1.468 ± 0.45 <i>31.0</i>	1.8 ± 0.31 <i>39.6</i>	0.16 ± 0.02 <i>22.9</i>	0.32 ± 0.05*** <i>37.3</i>	58.11 ± 8.0 <i>16.0</i>	2806 ± 40.47 <i>5.4</i>
<i>R. darwinii</i> Queulat (n= 12)	1.522 ± 0.48 <i>30.0</i>	1.4 ± 0.17 <i>21.1</i>	0.16 ± 0.02 <i>18.4</i>	0.3 ± 0.04*** <i>24.7</i>	67.27 ± 21.7 <i>13.0</i>	3006 ± 45.62 <i>4.1</i>
<i>R. darwinii</i> Tantauco (n= 13)	1.752 ± 0.60 <i>34.0</i>	1.8 ± 0.36* <i>29.8</i>	0.18 ± 0.02 <i>25.7</i>	0.37 ± 0.07*** <i>23.2</i>	77.41 ± 12.6 <i>19.0</i>	3749 ± 96.58*** <i>6.0</i>

234 *Acoustic differences between species*

235 The GLM shows that advertisement calls of *R. rufum* from Las Carmelitas mainly differed
236 from *R. darwinii* from Tantauco population in their call duration ($t = 2.169$, $P = 0.036$), inter-
237 note interval ($t = 5.992$, $P < 0.001$) and dominant frequency ($t = 12.218$, $P < 0.001$). In
238 addition, inter-note interval of calls from Las Carmelitas show significant differences with the
239 calls of Huilo Huilo ($t = 3.996$, $P < 0.001$) and Queulat ($t = 3.797$, $P < 0.001$). No differences
240 between the *R. rufum* population and the three *R. darwinii* populations occurred in call rate,
241 note duration and proportion of chaos (Table 1). Although the most common call issued in the
242 *R. rufum* population is the five-note call and in the three *R. darwinii* populations it is the four-
243 note call (Fig. 3), the Kolmogorov–Smirnov test showed no differences in this feature among
244 the four populations ($D < 0.67$, $P > 0.13$ for all comparisons).

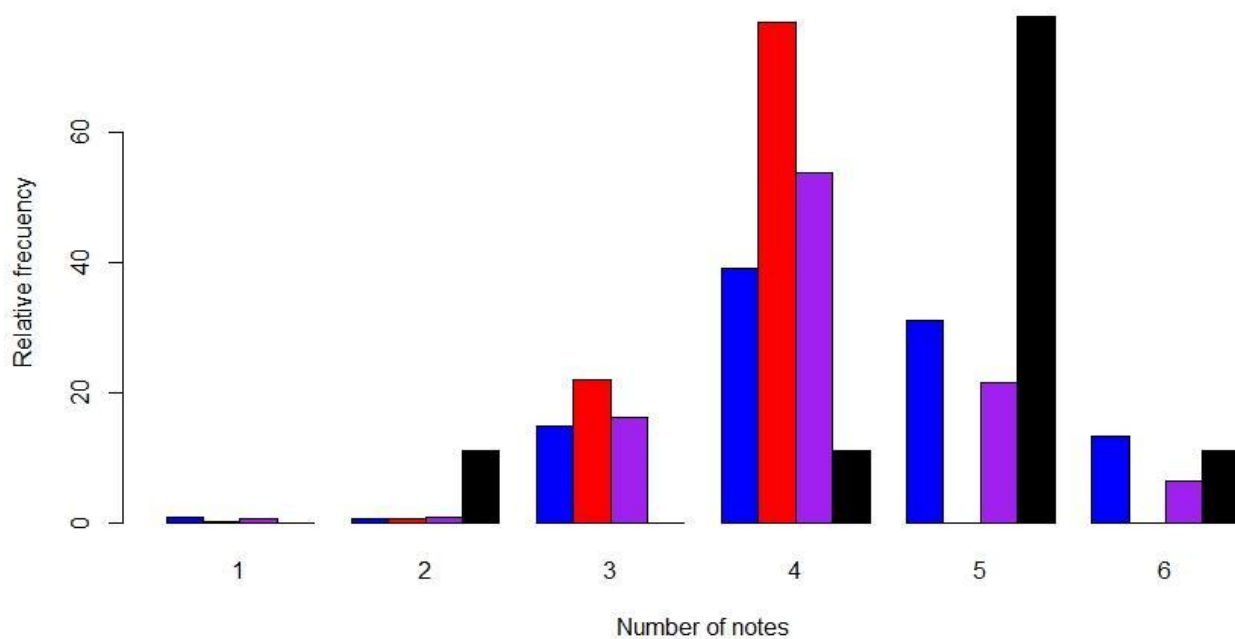


Figure 3. Proportions of number of notes in the calls of each population of *Rhinoderma*. Blue: Huilo Huilo, red: Queulat, purple: Tantauco, black: Las Carmelitas

245 *Distinctiveness analysis for all populations*

246 The PCA grouped the acoustic variables in six principal components from which the first four
 247 were considered, as they accumulated 86.7% of the explained variance (Table 2). The
 248 variables with higher weight (scores higher than 0.4) for PC1 after the z transformation were
 249 call duration, note duration, inter-note interval and dominant frequency (Table 2). Dominant
 250 frequency and chaos proportion had the highest weights for PC2, call rate and note duration
 251 for PC3 and note duration and inter-note interval for PC4 (Table 2).

Table 2. Loadings of the acoustic variables, eigenvalues and explained variances for each principal component (PC), resulting from the analysis including the four populations studied. Since the cumulative explained variance reaches an 86.77% with PC4, the first four PCs were considered for the discrimination function analysis.

Acoustic characteristics	PC1	PC2	PC3	PC4	PC5	PC6
Call rate	-0.142	0.269	-0.859	0.224	-0.293	0.184
Call duration	-0.542	0.236	0.114	-0.313	-0.454	-0.577
Note duration	-0.410	0.220	0.448	0.642	-0.191	0.366
Inter-note interval	-0.534	0.091	-0.038	-0.535	0.375	0.527
Chaos proportion	-0.149	-0.808	-0.027	-0.092	-0.503	0.251
Dominant frequency	-0.458	-0.403	-0.217	0.380	0.527	-0.399
Eigenvalues	2.196	1.223	1.053	0.733	0.502	0.290
Explained variance (%)	36.610	20.390	17.560	12.22	8.383	4.837
Cumulative explained variance (%)	36.610	57.000	74.560	86.780	95.163	100.000

252 The LDA correctly classified the individuals from Tantauco (*R. darwinii*), with 85%
253 of success, followed by 50% for Queulat (*R. darwinii*), 50% for Huilo Huilo (*R. darwinii*) and
254 33% of success for Las Carmelitas (*R. rufum*). The LDA indicated that the most important
255 components for discrimination among groups were PC1 and PC2 in the first linear
256 discriminant (LD1), which explained 93% of the variation, and PC4 was the most important
257 in the second linear discriminant (LD2), which explains 7% of the variation (Fig. 4A). As
258 such, the variables having larger relevance in the LDA because of their weights for principal
259 components were dominant frequency, call duration, inter-note interval and chaos proportion,
260 as shown in Table 2. As such, the LDA grouped the individuals in two clear clusters separated
261 mainly across LD1. These groupings did not show a separation between species. The left
262 cluster was completely formed by individuals from the *R. darwinii* insular population of

263 Tantauco and the right cluster was formed by the three mainland populations, comprising *R.*
264 *darwinii* and *R. rufum* (Fig. 4A).

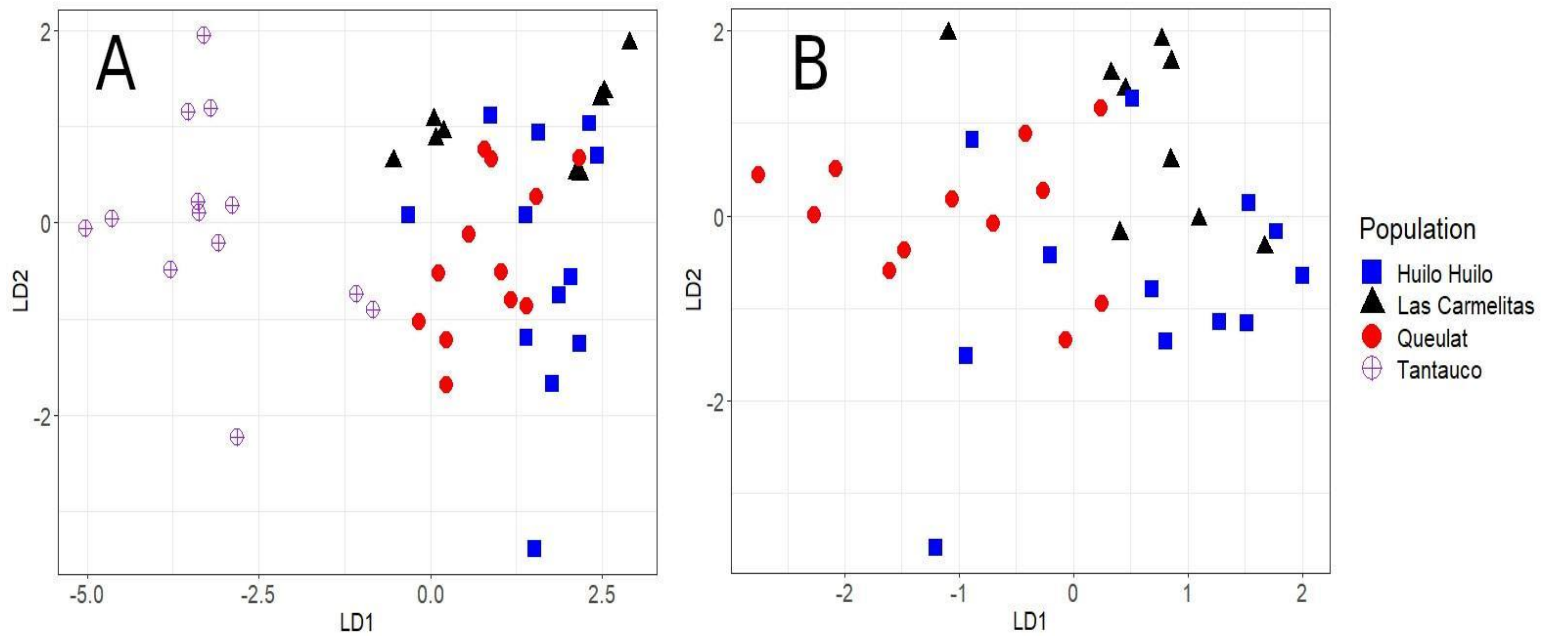


Figure 4. Discriminant analysis of call characteristics of individuals of *R. darwinii* and *R. rufum*: A) biplot of four localities showing that LD1 and LD2 account for 93 and 6.5 % of the total variance, respectively, and B) biplot excluding individuals from the insular population of Tantauco, showing that LD1 and LD2 account for 58% and 42% of the total variance, respectively.

265 *Distinctiveness analysis for mainland populations*

266 As populations were grouped indistinctly of the species to which they belong and because
267 discrimination showed separation among mainland and insular populations (Fig. 4A), we
268 performed a re-classification following the initial procedure, but excluding the individuals
269 from the insular population of Tantauco. This procedure intended to preclude the large
270 difference between mainland populations masked by the large variance of Tantauco. This
271 second classification increased the percentages of correct classification to 56%, 58% and 67%
272 for Las Carmelitas, Queulat and Huilco Huilo, respectively. Even though distinctiveness of the
273 mainland populations improved after excluding the insular population, the grouping did not
274 separate populations based on their species identity (Fig. 4B).

Table 3. Loadings of the acoustic variables, eigenvalues and explained variances for each principal component (PC), resulting from the analysis excluding the insular population of Tantauco. Since the cumulative explained variance reaches an 85.03% for PC4, the first four PCs were considered for the discrimination function analysis.

Acoustic characteristics	PC1	PC2	PC3	PC4	PC5	PC6
Call rate	-0.046	0.688	-0.067	0.506	0.361	-0.366
Call duration	-0.627	-0.021	-0.257	0.135	0.359	0.627
Note duration	-0.473	-0.041	0.308	0.480	-0.667	-0.069
Inter-note interval	-0.512	0.098	-0.459	-0.492	-0.157	-0.501
Chaos proportion	-0.055	-0.717	-0.170	0.399	0.336	-0.426
Dominant frequency	0.341	0.030	-0.772	0.307	-0.398	0.187
Eigenvalues	1.875	1.383	0.990	0.857	0.642	0.250
Explained variance (%)	31.260	23.060	16.510	14.290	10.710	4.182
Cumulative explained variance (%)	31.260	54.320	70.830	85.110	95.820	100.000

275 Discussion

276 *Geographic variation of Rhinoderma advertisement calls and body size*

277 Overall, advertisement calls of both *Rhinoderma* species were acoustically similar. Our
278 results indicate that the calls of *R. rufum* recorded in the sympatric zone did not differ
279 substantially from the calls of other mainland populations of *R. darwinii*. Among the
280 populations analysed, the insular population of *R. darwinii* showed larger divergence relative
281 to the population of *R. rufum*. Geographic variation of calls observed within mainland
282 populations of *Rhinoderma* (including *R. rufum* and *R. darwinii*) assimilates to the one
283 reported for *P. thaul*, a species having a wide distribution and environmental range in Chile
284 (Velásquez, 2014). Similar instances of geographic variation occur in *Hyla cinerea* in the
285 USA (Asquith et al., 1988) and *Allobates femoralis* in Brazil (Simões, 2008; Amézquita et al.,
286 2009). In these cases, geographic distance and geological barriers play a key role limiting
287 gene flow between populations, contributing to acoustic divergence. The lack of divergence

288 between the calls of *R. rufum* and continental *R. darwinii* populations contrasts with the
289 notable divergence of the island population of Tantauco, differing in several acoustic
290 properties from the mainland populations of *Rhinoderma*. This mode of geographic variation
291 in acoustic traits does not support the divergence process causing isolation between *R. rufum*
292 and *R. darwinii*, as suggested by morphological, karyological and molecular evidence in
293 earlier studies.

294 The calls of *R. rufum* differed from the three populations of *R. darwinii* in having shorter
295 inter-note interval duration, and from the Tantauco population in having shorter call duration.
296 Temporal variables in frog calls can result from environmental influences. For example, in
297 some anurans, relationships between body temperature and pulse rate (Gerhardt, 1978; Ryan,
298 1988; Giacoma & Castellano, 2001), and between air temperature and call duration (Gayou,
299 1984; Bee et al., 2013; Ziegler et al., 2016) have been reported. It is not possible to assess the
300 influence of temperature on the call differences between *Rhinoderma* species because of the
301 lack of precise temperature measurements for each individual of *R. rufum* recorded, however
302 it is unlikely that this factor is relevant in this particular case, as the temperature range of
303 recordings of *R. rufum* (14-18 °C) are within the ranges of those of the three populations of *R.*
304 *darwinii* recorded (Serrano et al. 2020).

305 Among the *Rhinoderma* populations analysed, the island population of *R. darwinii* in
306 Tantauco has the most divergent call characteristics relative to *R. rufum*, including inter-note
307 interval, call duration and dominant frequency. The Tantauco population of *R. darwinii* is
308 characterized by its smaller body size relative to mainland populations (Valenzuela-Sánchez
309 et al., 2015), and this morphometric feature could account for the variation of inter-note
310 interval duration and dominant frequency, as these traits are strongly related to body size of
311 this species (Serrano et al., 2020). In particular, the inverse relationship between body size

312 and dominant frequency of the advertisement call occurs extensively in anurans (Castellano &
313 Giacomini, 2000; Gerhardt & Huber, 2002).

314 The smaller size of the Tantauco population could be related to different geographical
315 and environmental factors. Populations of different vertebrates inhabiting insular territories
316 have a tendency to small size relative to continental counterparts, a tendency known as the
317 Island Rule (Van Valen, 1973), however this relationship does not occur regularly in anurans
318 (Rebouças et al., 2018). Another environmental determinant of anuran size is temperature
319 seasonality, which is inversely related to size of *R. darwinii* populations. The low temperature
320 seasonality in Tantauco is in correspondence with the relatively small size of animals of this
321 population (Valenzuela et al 2015).

322 Divergence in acoustic signals of populations not related to geographic distance has
323 been proposed to depend on genetic drift (Wilczynski & Ryan, 1999; Amézquita et al., 2009;
324 Velásquez et al., 2013) or multiple evolutionary forces such as inter and intra-sexual selection
325 (Asquith et al., 1988; Littlejohn, 1999; Petrusková et al., 2010; Velásquez, 2014). In
326 *Rhinoderma*, calls show a pattern of variation concordant with geographic isolation. Indeed,
327 after excluding the island population of Tantauco from distinctiveness analysis,
328 reclassification percentages of the mainland populations increased, but do not contribute to
329 improve the distinctiveness of the *R. rufum* population from Las Carmelitas. This result
330 indicates that the status of distinct species is unlikely for continental populations of
331 *Rhinoderma* (Funk et al., 2012). Such tenet is strengthened by considering that the population
332 of *R. rufum* analyzed is within the area of sympatry with *R. darwinii* (Bourke et al., 2012), as
333 in these zones, signal divergence between related species is in diverse cases reinforced,
334 promoting reproductive isolation and preventing deleterious hybridization (Littlejohn, 1999,
335 2001; Verrell, 1999; Höbel & Gerhardt, 2003; Rodríguez-Tejeda et al., 2014; Köhler et al.,
336 2017). As such, the lack of differentiation between the advertisement calls of *R. rufum* and *R.*

337 *darwinii* in the sympatric points to the absence of a main pre-mating isolation barrier
338 characteristic of anurans.

339 Our data indicate that the greatest acoustic divergence in *Rhinoderma* has taken place
340 in the island of Chiloé. Call differentiation related to geographic isolation, concurrent
341 morphological differentiation and exposure to extreme environments are likely to result in
342 speciation processes in anurans (Nevo & Capranica, 1985; Pröhl et al., 2006; Amézquita et
343 al., 2009; Funk et al., 2012; Rebouças et al., 2020). The historical geographic isolation of
344 Chiloé island dates from the late Pleistocene (Heusser, 1982, Villagrán et al., 1986; Denton et
345 al., 1999), and could account for the acoustic differentiation of the Tantauco population. The
346 distinctiveness analysis concurs to identify Tantauco as the only population that stands out
347 forming a cluster apart from the rest of populations. It is noteworthy that dominant frequency,
348 the only acoustic trait categorized as a static variable for all the populations analysed
349 (Gerhardt 1991) and likely having taxonomical relevance relative to dynamic variables, had a
350 high contribution to the separation between Tantauco and the mainland populations.
351 Similarly, Rebouças et al. (2020) found that call frequency is a static variable in *Boana*
352 *albomarginata* but having a lower frequency in island respect to mainland populations related
353 to larger body size. In contrast, the lack of distinctiveness observed between continental
354 populations of the two species of *Rhinoderma*, indicates an absence of reproductive barriers.

355 *Variation in other characters in Rhinoderma*

356 Studies of phenotypic and genetic characters have stressed the differences between *R.*
357 *darwinii* and *R. rufum* comprising morphological traits in adults and tadpoles and larval phase
358 and parental care phenology (Formas et al., 1975; Jorquera et al., 1981, 1982; Formas, 2013).
359 However, variation in such traits can result from phenotypic plasticity, as has been shown to
360 occur in other anurans some anuran species (Noble, 1925; Zippel, 1997; Relyea, 2003). The
361 expansion of interdigital membranes and the vascularization of folds in the integument can

362 vary according to the water level and other environmental conditions during development
363 (Cuello et al., 2008). The length of the larval stage can vary in response to environmental
364 conditions such as temperature, population density and food availability (Vences et al., 2001;
365 Cuello et al., 2014). Even the digestive tract development can vary in response to
366 temperature, as seen in tadpoles of *Calyptocephalella gayi* (Castañeda et al., 2006). Growth
367 rate of tadpoles and size at metamorphosis has shown to have effects on oral structures in
368 North American frogs (Venesky et al., 2010; Vences et al., 2001).

369 In particular, concerning the free aquatic living larval period that characterizes and
370 distinguishes *R. rufum* from *R. darwinii*, Wilhelm (1927) and Pflaumer (1935) successfully
371 raised adults from tadpoles taken from the vocal sac of *R. darwinii*'s specimens, showing the
372 plasticity of parental care, in contrast with the assertion of Goicoechea et al. (1986) that
373 tadpoles of *R. darwinii* are unable to develop in water. Parental care in vertebrates is a labile
374 behaviour and its variation in closely related species relates to the degree of parental
375 investment (Reynolds et al., 2002), as behavioural decisions to care for tadpoles are a trade-
376 off that compensate for higher costs of diverse selective pressures affecting reproduction
377 (Schulte et al., 2020). Reproductive site selection in the Brazilian treefrog *Aparasphenodon*
378 *arapapa* (Lantyer-Silva et al., 2018) and hatching age in glass frogs and tree frogs
379 (Warkentin, 1995; Delia et al. 2019) can be accounted for by plasticity.

380 The geographical distribution of the two *Rhinoderma* species suggests that differences
381 between the two taxa could relate to different environmental pressures as the boundary
382 between the distributions of *R. rufum* and *R. darwinii* coincides with the transition of
383 Mediterranean and temperate forests (Luebert & Plischoff, 2006).

384 The use of chromosomal karyotypes was widely used for taxonomic delimitation of
385 amphibians in the 60s (Hillis, 2019), and in Chilean amphibian as well, contributing to clarify
386 inter-genera and inter-family systematics, but having limited resolution between congeneric

387 species (Díaz & Veloso, 1979). The karyotypic structures of the two species of *Rhinoderma*
388 are basically similar, showing minor differences in chromosomal arms length and centromeric
389 fusions (Formas, 1976; Díaz & Veloso, 1979). This karyological technique as unique criteria
390 is not used in present days to set species relationships, as if not contrasted with banding
391 techniques to identify nucleolus organizer regions (Utsumi & Takehisa, 1974) it is not
392 appropriate to identify synapomorphies and can misidentify truly homologous characters
393 (Dobigny et al., 2002). Historically, chromosomal rearrangements (for example, between
394 haploid and diploid species) provide stronger evidence for reproductive isolation rather than
395 comparisons in chromosomal structures (Hillis, 2019). Instead, the use of one or more
396 combined DNA molecular markers (Funk et al., 2012; Hillis, 2019) would provide more
397 repeatable and comparable evidence to integrate genetics in delimiting species boundaries in
398 *Rhinoderma*.

399 In recent years, several other traits have been shown to vary widely within *R.*
400 *darwinii*, for example, climate-dependent colour variation (Bourke et al., 2011), geographic
401 variation of body size and nasal appendix extension (Valenzuela-Sánchez et al., 2015), and
402 population variation of the advertisement call (Serrano et al., 2020), including the occurrence
403 of release calls (Serrano et al., accepted). We propose that the morphological differences
404 observed in *Rhinoderma* could be related to specific environmental conditions influencing the
405 development of local morphotypes instead of speciation, as formulated in former studies.

406 *Conservation implications*

407 Our study is framed within the objectives of the Binational Conservation Strategy for
408 Darwin's Frogs, for which priorities are to define 1) the taxonomic status of *R. rufum* and 2)
409 relevant units for the conservation of *R. darwinii* (Azat et al., 2020). These definitions are
410 important to focus on prospective conservation actions, especially since *Rhinoderma*

411 populations are largely isolated among them (Bourke et al., 2018) and dispersal ability of the
412 species is likely limited (Uribe-Rivera et al., 2017). Such isolation restricts the genetic
413 diversity of the species and can jeopardize its survival while facing an alarming climate
414 change rate (Cummins et al., 2019), which has been identified as a main factor of Darwin's
415 frogs' decline (Bourke et al., 2018). In this scenario, a managed re-localization of populations
416 could be effective for in-situ conservation actions (Gascon, 2007; Olden et al., 2011). The
417 acoustic characterization that we have conducted for *Rhinoderma* can contribute to identify
418 populations and individuals sampled by means of automated acoustic monitoring (Gilbert et
419 al., 2002; Tripp and Otter, 2006; Vögeli et al., 2008), and also provide management tools in
420 wild sites using attracting or repelling signals for wildlife (Aubin, 1990; Buxton et al., 2018;
421 Santema et al., 2019).

422 The bioacoustic evidence contributed by our study and the critical review of
423 phenotypic and genetic traits studied formerly in *Rhinoderma* indicates that *R. rufum* may not
424 be a species different from *R. darwini*, suggesting instead that *Rhinoderma* should be
425 considered as a monospecific genus. Thus, the divergence between populations of
426 *Rhinoderma* more likely corresponds to an instance of geographic variation dependent on
427 climate regimes along the historical distribution range of the genus. Assessing the plasticity
428 supporting the behavioural and morphological differentiation between northern and southern
429 populations of *Rhinoderma* would be important to implement adaptive management measures
430 (Caro, 2007; Gascon, 2007), such as creating common gardens in protected areas where
431 *Rhinoderma* has disappeared. This management would also allow to confront the challenges
432 that climate change, habitat loss and chytridiomycosis impose on *Rhinoderma* in the near
433 future (Uribe-Rivera et al., 2017; Bourke et al., 2018) and hopefully will contribute new
434 measures for long-term conservation initiatives for this unique anuran entity.

435 **Acknowledgements**

436 This study was partially supported by Stiftung Artenschutz 2018.

437 **References**

- 438 Amézquita, A., Lima, A. P., Jehle, R., Castellanos, L., Ramos, O., Crawford, A. J. & Hödl,
439 W. (2009). Calls, colours, shape, and genes: a multi-trait approach to the study of
440 geographic variation in the Amazonian frog *Allobates femoralis*. *Biol J Linn Soc.* **98**, 826-
441 838. <https://doi.org/10.1111/j.1095-8312.2009.01324.x>
- 442 Asquith, A., Altig, R., & Zimba, P. (1988). Geographic variation in the mating call of the
443 green treefrog *Hyla cinerea*. *Am. Midl. Nat.* **119**, 101-110. <https://doi.org/10.2307/2426058>
- 444 Araya-Salas, M., Smith-Vidaurre, G., Mennill, D. J., González-Gómez, P. L., Cahill, J., &
445 Wright, T. F. (2019). Social group signatures in hummingbird displays provide evidence of
446 co-occurrence of vocal and visual learning. *Proc. R. Soc. B.* **286**, 20190666.
447 <https://doi.org/10.1098/rspb.2019.0666>
- 448 Aubin, T. (1990). Synthetic bird calls and their application to scaring methods. *Ibis* **132**, 290-
449 299. <https://doi.org/10.1111/j.1474-919X.1990.tb01046.x>
- 450 Azat, C., Valenzuela-Sánchez, A., Delgado, S., Cunningham, A., Alvarado-Rybak, M.,
451 Bourke, J. & Angulo, A. (2020). A flagship for Austral temperate forest conservation: an
452 action plan for Darwin's frogs bringing together key stakeholders. *Oryx*, 1-8.
453 <https://doi.org/10.1017/S0030605319001236>
- 454 Bee, M. A., Suyesh, R., & Biju, S. D. (2013). Vocal behavior of the Ponmudi Bush Frog
455 (*Raorchestes graminirupes*): repertoire and individual variation. *Herpetologica* **69**, 22-35.
456 <https://doi.org/10.1655/HERPETOLOGICA-D-11-00042>
- 457 Bernal, X. E., Guarnizo, C., & Lüddecke, H. (2005). Geographic variation in advertisement
458 call and genetic structure of *Colostethus palmatus* (Anura, Dendrobatidae) from the
459 Colombian Andes. *Herpetologica* **61**, 395-408.

460 Böhm, W., & Hornik, K. (2012). A Kolmogorov-Smirnov test for r samples. *Fundamenta*
461 *Informaticae*, **117**, 103-125.

462 Bolaños-Sittler, P., Sueur, J., Fuchs, J., & Aubin, T. (2020). Vocalisation of the rare and
463 flagship species *Pharomachrus mocinno* (Aves: Trogonidae): implications for its
464 taxonomy, evolution and conservation. *Bioacoustics* **29**, 654-669
465 <https://doi.org/10.1080/09524622.2019.1647877>

466 Bosch, J., & De la Riva, I. (2004). Are frog calls modulated by the environment? An analysis
467 with anuran species from Bolivia. *Can. J. Zool.* **82**, 880-888. <https://doi.org/10.1139/z04->
468 060

469 Bourke, J., Barrientos, C., Ortiz, J.C., Busse, K., Böhme, W., & Bakker, T.C. (2011) Colour
470 change in Darwin's frogs (*Rhinoderma darwinii*, Duméril and Bibron, 1841) (Anura:
471 Rhinodermatidae). *J. Nat. Hist.* **45**, 2661-2668.

472 Bourke, J., Busse, K. & Böhme, W. (2012) Searching for a lost frog (*Rhinoderma rufum*):
473 identification of the most promising areas for future surveys and possible reasons of its
474 enigmatic decline. *North-West. J. Zool.* **8**, 99-106.

475 Bourke, J., Busse, K., & Boehme, W. (2018). Potential effects of climate change on the
476 distribution of the endangered Darwin's frog. *North-West. J. Zool.* **14**, 165-170.
477 <http://biozoojournals.ro/nwjz/index.html>

478 Bradbury, J.W., & Vehrencamp S.L. 2011. *Principles of animal communication*. 2nd edn.
479 Sunderland, Massachusetts: Sinauer.

480 Buxton, V. L., Ward, M. P., & Sperry, J. H. (2018). Evaluation of conspecific attraction as a
481 management tool across several species of anurans. *Diversity* **10**, 6.

482 Butlin, R. K., & Ritchie, M. G. (1994). Behaviour and speciation. In *Behaviour and evolution*:
483 43–79. Slater, P. J. B., Halliday, T. R., & P. Barrett (Eds.). Cambridge University Press.

484 Cardoso, G. C., Hu, Y., & Mota, P. G. (2012). Birdsong, sexual selection, and the flawed
485 taxonomy of canaries, goldfinches and allies. *Anim. Behav.* **84**, 111-119.

486 Caro, T. (2007). Behavior and conservation: a bridge too far? *Trends. Ecol. Evol.* **22**, 394-400.
487 <https://doi.org/10.1016/j.tree.2007.06.003>

488 Castaneda, L. E., Sabat, P., Gonzalez, S. P., & Nespolo, R. F. (2006). Digestive plasticity in
489 tadpoles of the Chilean giant frog (*Caudiverbera caudiverbera*): factorial effects of diet and
490 temperature. *Physiol. Biochem. Zool.* **79**, 919-926. <https://doi.org/10.1086/506006>

491 Castellano, S., & Giacoma, C. (1998). Stabilizing and directional female choice for male calls
492 in the European green toad. *Anim. Behav.* **56**, 275-287.

493 Castellano, S., Giacoma, C., & Dujsebayaeva, T. (2000). Morphometric and advertisement call
494 geographic variation in polyploid green toads. *Biol. J. Linn. Soc.* **70**, 341-360.
495 <https://doi.org/10.1111/j.1095-8312.2000.tb00214.x>

496 CEPAL. (2012). La economía del cambio climático en Chile. Santiago: Naciones Unidas,
497 367. Download in
498 http://repositorio.cepal.org/bitstream/handle/11362/35372/S2012058_es.pdf?sequence=1.

499 Claridge, M. F. (1990). Acoustic recognition signals: barriers to hybridization in Homoptera
500 *Auchenorrhyncha*. *Can. J. Zool.* **68**, 1741-1746. <https://doi.org/10.1139/z90-255>

501 Cocroft, R.B., & Ryan, M.J. (1995) Patterns of advertisement call evolution in toads and chorus
502 frogs. *Anim. Behav.* **49**, 283–303

503 Crump, M. L., & Veloso, A (2005) El aporte de observaciones de terreno y del análisis genético
504 para la conservación de *Rhinoderma darwinii* en Chile. In *Historia, Biodiversidad y*
505 *Ecología de los Bosques Costeros de Chile*: 452–455. Smith-Ramirez, C., Armesto, J. J.,
506 Valdovinos, C. (Eds).. Santiago: Editorial Universitaria..

507 Cuello, M. E., Úbeda, C. A., & Bello, M. T. (2008). Relationship between morphotypes of
508 *Atelognathus patagonicus* (Anura, Neobatrachia) and environmental conditions: evidence

509 and possible explanation. *Phyllomedusa: J. Herpetol.* **7**, 35-44.
510 <https://doi.org/10.11606/issn.2316-9079.v7i1p35-44>

511 Cuello, M. E., Úbeda, C. A., Bello, M. T., & Perotti, M. G. (2014). Plastic patterns in larval
512 development of Endangered endemic *Atelognathus patagonicus*: Implications for
513 conservation strategies. *Endanger. Species Res.* **23**, 83-92. <https://doi.org/10.3354/esr00550>

514 Cuevas, C. C. (2014). Native forest loss impact's on anuran diversity: with focus on
515 *Rhinoderma rufum* (Philippi 1902) (Rhinodermatidae) in Coastal Range, South-Central
516 Chile. *Gestión Ambiental* **27**, 1-18.

517 Delia, J., Rivera-Ordóñez, J.M., Salazar-Nicholls, M.J. et al. (2019). Hatching plasticity and
518 the adaptive benefits of extended embryonic development in glassfrogs. *Evol. Ecol.* **33**, 37–
519 53. <https://doi.org/10.1007/s10682-018-9963-2>

520 Denton, G. H., Heusser, C. J., Lowell, T. V., Moreno, P. I., Andersen, B. G., Heusser, L. E. &
521 Marchant, D. R. (1999). Interhemispheric linkage of paleoclimate during the last glaciation.
522 *Geogr. Ann. A.* **81**, 107-153. <https://doi.org/10.1111/1468-0459.00055>

523 Díaz, N. F. (1986). Biosistemática de los Leptodactylidae Chilenos. *Ann. Mus. Hist. Nat.*
524 *Valparaíso* **17**, 65-85.

525 Díaz, N. F., Veloso, A. (1979). Sistemática y evolución de los anfibios de Chile. *Arch. Biol.*
526 *Med. Exp.* **12**, 59-70.

527 Dobigny, G., Ozouf-Costaz, C., Bonillo, C., & Volobouev, V. (2002). “Ag-NORs” are not
528 always true NORs: new evidence in mammals. *Cytogenet. Genome. Res.* **98**, 75-77
529 <https://doi.org/10.1159/000068541>

530 Dobigny, G., Ducroz, J. F., Robinson, T. J., & Volobouev, V. (2004). Cytogenetics and
531 cladistics. *Syst. Biol.* **53**, 470-484. <https://doi.org/10.1080/10635150490445698>

532 Escalona Sulbarán, M. D., Ivo Simões, P., Gonzalez-Voyer, A., & Castroviejo-Fisher, S.
533 (2019). Neotropical frogs and mating songs: The evolution of advertisement calls in glass
534 frogs. *J. Evolution. Biol.* **32**, 163-176. <https://doi.org/10.1111/jeb.13406>

535 Feng, A. S., Tobias, R., Arch, V. S., Yu, Z., Xu, Z. M., Yu, X. J., & Shen, J. X. (2009). Diversity
536 of the vocal signals of concave-eared torrent frogs (*Odorrana tormota*): evidence for
537 individual signatures. *Ethology* **115**, 1015–1028. [https://doi.org/10.1111/j.1439-](https://doi.org/10.1111/j.1439-0310.2009.01692.x)
538 [0310.2009.01692.x](https://doi.org/10.1111/j.1439-0310.2009.01692.x)

539 Formas, R., Pugin, E., & Jorquera, B. (1975). La identidad del batracio Chileno *Heminectes*
540 *rufus* Philippi, 1902. *Physis Sección C* **34**, 147–157.

541 Formas, J. R. (1976). New karyological data of *Rhinoderma*: The chromosomes of *Rhinoderma*
542 *rufum*. *Experientia* **32**, 1000-1002.

543 Formas, J. R. (2013). External morphology, chondrocranium, hyobranchial skeleton, and
544 external and internal oral features of *Rhinoderma rufum* (Anura, Rhinodermatidae). *Zootaxa*
545 **3641**, 395-400. <http://dx.doi.org/10.11646/zootaxa.3641.4.6>

546 Freeman, B. G., & Montgomery, G. A. (2017). Using song playback experiments to measure
547 species recognition between geographically isolated populations: A comparison with
548 acoustic trait analyses. *Auk* **134**, 857-870. <https://doi.org/10.1642/AUK-17-63.1>

549 Friedl, T. W. (2006). Individual male calling pattern and male mating success in the European
550 treefrog (*Hyla arborea*): is there evidence for directional or stabilizing selection on male
551 calling behaviour?. *Ethology* **112**, 116-126. [https://doi.org/10.1111/j.1439-](https://doi.org/10.1111/j.1439-0310.2005.01132.x)
552 [0310.2005.01132.x](https://doi.org/10.1111/j.1439-0310.2005.01132.x)

553 Funk, W. C., Caminer, M., & Ron, S. R. (2012). High levels of cryptic species diversity
554 uncovered in Amazonian frogs. *P. Roy. Soc. B-Biol. Sci.* **279**, 1806-1814.
555 <https://doi.org/10.1098/rspb.2011.1653>

556 Gascon, C. (2007). Amphibian conservation action plan: proceedings IUCN/SSC Amphibian
557 Conservation Summit 2005. IUCN.

558 Gayou, D. C. (1984). Effects of temperature on the mating call of *Hyla versicolor*. *Copeia* **1984**,
559 733-738. <https://doi.org/10.2307/1445157>

560 Gerhardt, H. C. (1978). Temperature coupling in the vocal communication system of the gray
561 tree frog, *Hyla versicolor*. *Science* **199**, 992-994.
562 <https://doi.org/10.1126/science.199.4332.992>

563 Gerhardt, H. C. (1991). Female mate choice in treefrogs: static and dynamic acoustic criteria.
564 *Anim. Behav.* **42**, 615-635. [https://doi.org/10.1016/S0003-3472\(05\)80245-3](https://doi.org/10.1016/S0003-3472(05)80245-3)

565 Gerhardt, H. C. (2001). Acoustic communication in two groups of closely related treefrogs.
566 *Adv. Stud. Behav.* **30**, 99-167. [https://doi.org/10.1016/S0065-3454\(01\)80006-1](https://doi.org/10.1016/S0065-3454(01)80006-1)

567 Gerhardt, H. C., & Huber, F. (2002). Acoustic communication in insects and anurans: common
568 problems and diverse solutions. University of Chicago Press.

569 Gerhardt, H. C., Roberts, J. D., Bee, M. A., & Schwartz, J. J. (2000). Call matching in the
570 quacking frog (*Crinia georgiana*). *Behav. Ecol. Sociobiol.* **48**, 243-251.
571 <https://doi.org/10.1007/s002650000226>

572 Giacoma, C., & Castellano, S. (2001). Advertisement call variation and speciation in the *Bufo*
573 *viridis* complex. In *Anuran Communication*: 205–219. Ryan, M. J. (Ed.) Smithsonian
574 Institution Press.

575 Gingras, B., Boeckle, M., Herbst, C. T., & Fitch, W. T. (2013). Call acoustics reflect body size
576 across four clades of anurans. *J. Zool.* **289**, 143-150. [https://doi.org/10.1111/j.1469-](https://doi.org/10.1111/j.1469-7998.2012.00973.x)
577 [7998.2012.00973.x](https://doi.org/10.1111/j.1469-7998.2012.00973.x)

578 Gilbert, G., Tyler, G. A., & Smith, K. W. (2002). Local annual survival of booming male Great
579 Bittern *Botaurus stellaris* in Britain, in the period 1990–1999. *Ibis* **144**, 51-61.

580 Goicoechea, O., Garrido, O., & Jorquera, B. (1986). Evidence for a trophic paternal-larval
581 relationship in the frog *Rhinoderma darwinii*. *J. Herpetol.* **20**, 168-178.

582 Greenbaum, E., Sinsch, U., Lehr, E., Valdez, F., & Kusamba, C. (2013). Phylogeography of
583 the reed frog *Hyperolius castaneus* (Anura: Hyperoliidae) from the Albertine Rift of Central
584 Africa: Implications for taxonomy, biogeography and conservation. *Zootaxa* **3731**, 473-494.

585 Heusser, Calvin J. (1982) Palynology of cushion bogs of the Cordillera Pelada, Province of
586 Valdivia, Chile. *Quaternary Res.* **17**, 71-92. [https://doi.org/10.1016/0033-5894\(82\)90046-1](https://doi.org/10.1016/0033-5894(82)90046-1)

587 Heyer, W. R., de Sá, R. O., & Rettig, A. (2005). Sibling Species, Advertisement Calls, and
588 Reproductive Isolation in Frogs of the *Leptodactylus pentadactylus* Species Cluster
589 (Amphibia, Leptodactylidae). *Russ. J. Herpetol.* **12**, 35-39.

590 Höbel, G., & Gerhardt, H. C. (2003). Reproductive character displacement in the acoustic
591 communication system of green tree frogs (*Hyla cinerea*). *Evolution* **57**, 894-904.
592 <https://doi.org/10.1111/j.0014-3820.2003.tb00300.x>

593 Höbel, G., & Fellows, S. (2016). Vocal repertoire and calling activity of a Dwarf Clawed Frog
594 (*Hymenochirus boettgeri*). *Herpetol. Rev.* **47**, 543-549.

595 IUCN SSC Amphibian Specialist Group. (2015). *Rhinoderma rufum*. The IUCN Red List of
596 Threatened Species 2015: e.T19514A79809567.
597 <https://dx.doi.org/10.2305/IUCN.UK.2015-4.RLTS.T19514A79809567.en>. Downloaded
598 on 7 March 2020.

599 Jackson, J.E. (2003). *A User's Guide to Principal Components*. New York: John Wiley &
600 Sons.

601 Jehle, R., & Arak, A. (1998). Graded call variation in the Asian cricket frog *Rana nicobariensis*.
602 *Bioacoustics* **9**, 35-48. <https://doi.org/10.1080/09524622.1998.9753378>

603 Jorquera, B., Pugin, E., Garrido, O., Goicoechea, O., & Formas, R. (1981). Procedimiento de
604 desarrollo en dos especies del género *Rhinoderma*. *Medio Ambiente (Chile)* **5**, 58-71.

605 Jorquera, B., Garrido, O., & Pugin, E. (1982) Comparative studies of the digestive tract
606 development between *Rhinoderma darwinii* and *R. rufum*. *J. Herpetol.* **16**, 205-214.

607 Köhler, J., Jansen, M., Rodríguez, A., Kok, P. J. R., Toledo, L. F., Emmrich, M., Glaw, F.,
608 Haddad, C. F. B., Rödel, M. O., Vences, M. (2017). The use of bioacoustics in anuran
609 taxonomy: theory, terminology, methods and recommendations for best practice. *Zootaxa*
610 **4251**, 1-124. <http://dx.doi.org/10.11646/zootaxa.4251.1.1>

611 Korkmaz, S., Goksuluk, D., & Zararsiz, G. (2014). MVN: An R package for assessing
612 multivariate normality. *The R Journal* **6**, 151-162.

613 Kuramoto, M. (1997). Relationships of the Palau Frog, *Platymantis pelewensis* (Anura:
614 Ranidae): Morphological, Karyological, and Acoustic Evidence. *Copeia* **1997**, 183-187.
615 <https://doi.org/10.2307/1447855>

616 Laiolo, P. (2010). The emerging significance of bioacoustics in animal species conservation.
617 *Biol. Conserv.* **143**,1635-1645. <https://doi.org/10.1016/j.biocon.2010.03.025>

618 Lantyer-Silva, A. S. F., Waldron, A., Zina, J., & Solé, M. (2018). Reproductive site selection
619 in a bromeliad breeding treefrog suggests complex evolutionary trade-offs. *PLOS One*, **13**,
620 e0207131. <https://doi.org/10.1371/journal.pone.0207131>

621 Liang, S., Singh, M., Dharmaraj, S., & Gam, L-H. (2010). The PCA and LDA analysis on the
622 differential expression of proteins in breast cancer. *Dis. Markers.* **29**, 231-242.
623 <https://doi.org/10.3233/DMA-2010-0753>

624 Littlejohn, M. J. (1999). Variation in advertisement calls of anurans across zonal interactions:
625 the evolution and breakdown of homogamy. In *Geographic variation in behavior:
626 Perspectives on evolutionary mechanisms*: 209-233. Foster, S. A., & Endler, J. A. (Eds.).
627 Oxford University Press.

628

629 Littlejohn, M. J. (2001). Patterns of differentiation in temporal properties of acoustic signals of
630 anurans. In *Anuran Communication*: 102–120. Ryan, M. J. (Ed.) Smithsonian Institution
631 Press.

632 Lomolino, M. V. (2005). Body size evolution in insular vertebrates: generality of the island
633 rule. *J. Biogeogr.* **32**, 1683-1699. <https://doi.org/10.1111/j.1365-2699.2005.01314.x>

634 López, C., Quispe, M., Villalón, A., Concha, M. L., Penna, M., & Velásquez, N. A. (2020).
635 Geographic variation in the laryngeal morphology of a widely distributed South-American
636 anuran: behavioural and evolutionary implications. *Zool. J. Linn. Soc-Lond.* **190**, 140-148.
637 <https://doi.org/10.1093/zoolinnean/zlz175>

638 Luebert, F., & Plischoff, P. (2006). *Sinopsis bioclimática y vegetal de Chile*. Santiago:
639 Editorial Universitaria.

640 Mace, G. M. (2004). The role of taxonomy in species conservation. *Philos. Trans. R. Soc. Lond.*
641 *B Biol. Sci.* **359**, 711-719. <https://doi.org/10.1098/rstb.2003.1454>

642 Marcellini, D. (1977). Acoustic and visual display behavior of gekkonid lizards. *Am. Zool.* **17**,
643 251-260. <https://doi.org/10.1093/icb/17.1.251>

644 Mardia, K.V., Kent, J.T. and Bibby, J.M. (1979). *Multivariate analysis*. London: Academic
645 Press.

646 Miller, E. H., Areta, J. I., Jaramillo, A., Imberti, S., & Matus, R. (2020). Snipe taxonomy based
647 on vocal and non-vocal sound displays: the South American Snipe is two species. *Ibis* **162**,
648 968-990. <https://doi.org/10.1111/ibi.12795>

649 Moreno, P. I., Villagrán, C., Marquet, P. A., & Marshall, L. G. (1994). Quaternary
650 paleobiogeography of northern and central Chile. *Rev. Chil. Hist. Nat.* **67**, 487-502.

651 Næs, T., & Mevik, B. H. (2001). Understanding the collinearity problem in regression and
652 discriminant analysis. *J. Chemometr.* **15**, 413-426. <https://doi.org/10.1002/cem.676>

653 Nevo, E., Capranica R.R. (1985). Evolutionary origin of ethological reproductive isolation in
654 cricket frogs, *Acris*. In *Evolutionary Biology*: 147-214. Hecht M.K., Wallace B., Prance
655 G.T. (Eds.). Boston: Springer.

656 Noble, G. K. 1925. The integumentary, pulmonary, and cardiac modifications correlated with
657 increased cutaneous respiration in the Amphibia: a solution of the “hairy frog” problem. *J.*
658 *Morphol. Physiol.* **40**, 341–416. <https://doi.org/10.1002/jmor.1050400206>

659 Nussey, D. H., Wilson, A. J., & Brommer, J. E. (2007). The evolutionary ecology of individual
660 phenotypic plasticity in wild populations. *J. Evolution. Biol.* **20**, 831-844.
661 <https://doi.org/10.1111/j.1420-9101.2007.01300.x>

662 Obrist, M. K., Pavan, G., Sueur, J., Riede, K., Llusia, D., & Márquez, R. (2010). Bioacoustics
663 approaches in biodiversity inventories. *Abc Taxa* **8**, 68-99.

664 Olden, J. D., Kennard, M. J., Lawler, J. J., & Poff, N. L. (2011). Challenges and opportunities
665 in implementing managed relocation for conservation of freshwater species. *Conserv. Biol.*
666 **25**, 40-47. <https://doi.org/10.1111/j.1523-1739.2010.01557.x>

667 Padial, J. M., Miralles, A., De la Riva, I., & Vences, M. (2010). The integrative future of
668 taxonomy. *Front. Zool.* **7**, 1-14.

669 Payne, R. B. (1986). Bird songs and avian systematics. In *Current ornithology*: 87-126.
670 Johnston, R. F. (Ed.). Boston: Springer. https://doi.org/10.1007/978-1-4615-6784-4_2

671 Penna, M., & Veloso, A. (1990). Vocal diversity in frogs of the South American temperate
672 forest. *J. Herpetol.* **24**, 23-33. <https://doi.org/10.2307/1564285>

673 Petrusková, T., Osiejuk, T. S., & Petrusek, A. (2010). Geographic variation in songs of the tree
674 pipit (*Anthus trivialis*) at two spatial scales. *Auk* **127**, 274-282.
675 <https://doi.org/10.1525/auk.2009.09077>

676 Pflaumer, C. (1935). Observaciones biológicas acerca de la *Rhinoderma darwini* D & B. *Rev.*
677 *Chil. Hist. Nat.* **39**, 28-30.

678 Pröhl, H., Koshy, R. A., Mueller, U., Rand, A. S., & Ryan, M. J. (2006). Geographic variation
679 of genetic and behavioral traits in northern and southern túngara frogs. *Evolution* **60**, 1669-
680 1679. <https://doi.org/10.1111/j.0014-3820.2006.tb00511.x>

681 Pröhl, H., Hagemann, S., Karsch, J., & Höbel, G. (2007). Geographic variation in male sexual
682 signals in strawberry poison frogs (*Dendrobates pumilio*). *Ethology* **113**, 825-837.
683 <https://doi.org/10.1111/j.1439-0310.2007.01396.x>

684 Rebouças, R., da Silva, H. R., & Solé, M. (2018). Frog size on continental islands of the coast
685 of Rio de Janeiro and the generality of the Island Rule. *PLOS One* **13**, e0190153.

686 Rebouças, R., Augusto-Alves, G. and Toledo, L.F. (2020), Evolution of treefrogs' calls in
687 tropical islands might be under directional selection. *J Zool.* **312**, 43-52
688 <https://doi.org/10.1111/jzo.12792>

689 Reinhold, K. (2011) Variation in acoustic signalling traits exhibits footprints of sexual
690 selection. *Evolution* **65**, 738–745. <https://doi.org/10.1111/j.1558-5646.2010.01130.x>

691 Relyea, R. A. 2003. Predators come and predators go: the reversibility of predator-induced
692 traits. *Ecology* **84**, 1840–1848.

693 Reynolds, J. D., Goodwin, N. B., & Freckleton, R. P. (2002). Evolutionary transitions in
694 parental care and live bearing in vertebrates. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* **357**,
695 269-281.

696 Ripley, B., Venables, B., Bates, D. M., Hornik, K., Gebhardt, A., Firth, D., & Ripley, M. B.
697 (2013). Package ‘mass’. *Cran R*, 538.

698 Rodríguez-Tejeda, R. E., Méndez-Cárdenas, M. G., Islas-Villanueva, V., & Garcia, C. M.
699 (2014). Geographic variation in the advertisement calls of *Hyla eximia* and its possible
700 explanations. *PeerJ* **2**, e420.

701 RStudio Team (2019). RStudio: Integrated Development for R. RStudio, Inc., Boston, MA
702 URL <http://www.rstudio.com/>.

703 Ryan, M. J. (1988). Constraints and patterns in the evolution of anuran acoustic
704 communication. In *The evolution of the amphibian auditory system: 637-677*. Fritzsche, B.
705 (Ed.). John Wiley & Sons

706 Santema, P., Valcu, M., Clinchy, M., Zanette, L., & Kempenaers, B. (2019). Playback of
707 predator calls inhibits and delays dawn singing in a songbird community. *Behav. Ecol.* **30**,
708 1283-1288. <https://doi.org/10.1093/beheco/arz075>

709 Schulte, L. M., Ringler, E., Rojas, B., & Stynoski, J. L. (2020). Developments in amphibian
710 parental care research: history, present advances, and future perspectives. *Herpetol. Monogr.*
711 **34**, 71-97. <https://doi.org/10.1655/HERPMONOGRAPHS-D-19-00002.1>

712 Serrano, J. M. (2018). The amplexant and aggressive calls and notes on the reproductive
713 behaviour of the Porthole Frog (*Charadrahyla taeniopus*). *Herpetol. Notes* **11**, 819-823.

714 Serrano, J. M., Penna, M., & Soto-Azat, C. (2020). Individual and population variation of linear
715 and non-linear components of the advertisement call of Darwin's frog (*Rhinoderma*
716 *darwinii*). *Bioacoustics* **29**, 572-589. <https://doi.org/10.1080/09524622.2019.1631214>

717 Simões, P. I., Lima, A. P., Magnusson, W. E., Hödl, W., & Amézquita, A. (2008). Acoustic
718 and morphological differentiation in the frog *Allobates femoralis*: relationships with the
719 upper Madeira River and other potential geological barriers. *Biotropica* **40**, 607-614.
720 <https://doi.org/10.1111/j.1744-7429.2008.00416.x>

721 Soto-Azat, C., Valenzuela-Sánchez, A., Collen, B., Rowcliffe, J. M., Veloso, A., &
722 Cunningham, A. A. (2013). The population decline and extinction of Darwin's frogs. *PLOS*
723 *One*, **8**. <https://doi.org/10.1371/journal.pone.0066957>

724 Sueur, J., Aubin, T., Simonis, C., Lellouch, L., Brown, E. C., Depraetere, M., ... & LaZerte, S.
725 (2020). Package 'seewave'.

726 Tang, Y. Z., Zhuang, L. Z., & Wang, Z. W. (2001). Advertisement calls and their relation to
727 reproductive cycles in *Gekko gecko* (Reptilia, Lacertilia). *Copeia*, 248-253.
728 [https://doi.org/10.1643/0045-8511\(2001\)001\[0248:ACATRT\]2.0.CO;2](https://doi.org/10.1643/0045-8511(2001)001[0248:ACATRT]2.0.CO;2)

729 Targino, M., Elias-Costa, A. J., Taboada, C., & Faivovich, J. (2019). Novel morphological
730 structures in frogs: vocal sac diversity and evolution in Microhylidae (Amphibia: Anura).
731 *Zool. J. Linn. Soc-Lond.* **187**, 479-493. [https://doi.org/10.1643/0045-](https://doi.org/10.1643/0045-8511(2001)001[0945:OCITMY]2.0.CO;2)
732 [8511\(2001\)001\[0945:OCITMY\]2.0.CO;2](https://doi.org/10.1643/0045-8511(2001)001[0945:OCITMY]2.0.CO;2)

733 Tishechkin, D.Y., Vedenina, V.Y. (2016). Acoustic signals in insects: A reproductive barrier
734 and a taxonomic character. *Entomol. Rev.* **96**, 1127–1164
735 <https://doi.org/10.1134/S0013873816090013>

736 Toledo, L.F., Martins, I.A., Bruschi, D.P., Passos, M.A., Alexandre, C. & Haddad, C.F.B.
737 (2014) The anuran calling repertoire in the light of social context. *Acta Ethol.* **18**, 87–99.
738 <https://doi.org/10.1007/s10211-014-0194-4>

739 Tripp, T. M., & Otter, K. A. (2006). Vocal individuality as a potential long-term monitoring
740 tool for Western Screech-owls, *Megascops kennicottii*. *Can. J. Zool.* **84**, 744-753.
741 <https://doi.org/10.1139/z06-055>

742 Tyack, P. L. (2000). Dolphins whistle a signature tune. *Science* **289**, 1310-1311.
743 <https://doi.org/10.1126/science.289.5483.1310>

744 Uribe-Rivera, D., Soto-Azat, C., Valenzuela-Sánchez, A., Bizama, G., Simonetti, J., &
745 Pliscoff, P. (2017). Dispersal and extrapolation on the accuracy of temporal predictions
746 from species distribution models for the Darwin's frog. *Ecol. Appl.* **27**, 1633-1645.
747 <https://doi.org/10.1002/eap.1556>

748 Utsumi, S., & Takehisa, S. (1974). Heterochromatin differentiation in *Trillium kamtschaticum*
749 by ammoniacal silver reaction. *Exp. Cell. Res.* **86**, 398-401.

750 Valenzuela-Sánchez, A., Cunningham, A. A., & Soto-Azat, C. (2015). Geographic body size
751 variation in ectotherms: effects of seasonality on an anuran from the southern temperate
752 forest. *Front. Zool.* **12**, 37. <https://doi.org/10.1186/s12983-015-0132-y>

753 Valenzuela-Sánchez, A., Schmidt, B. R., Uribe-Rivera, D. E., Costas, F., Cunningham, A. A.,
754 & Soto-Azat, C. (2017). Cryptic disease-induced mortality may cause host extinction in an
755 apparently stable host–parasite system. *P. Roy. Soc. B-Biol. Sci.* **284**, 20171176.
756 <https://doi.org/10.1098/rspb.2017.1176>

757 Velásquez, N. A., Marambio, J., Brunetti, E., Méndez, M. A., Vásquez, R. A., & Penna, M.
758 (2013). Bioacoustic and genetic divergence in a frog with a wide geographical distribution.
759 *Biol. J. Linn. Soc.* **110**, 142-155. <https://doi.org/10.1111/bij.12113>

760 Velásquez, N. A. (2014). Geographic variation in acoustic communication in anurans and its
761 neuroethological implications. *J. Physiol-Paris.* **108**, 167-173.
762 <https://doi.org/10.1016/j.jphysparis.2014.10.001>

763 Velásquez, N. A., Opazo, D., Díaz, J., & Penna, M. (2014). Divergence of acoustic signals in
764 a widely distributed frog: relevance of inter-male interactions. *PLOS One* **9**, e87732.
765 <https://doi.org/10.1371/journal.pone.0087732>

766 Vences, M., Puente, M., Nieto, S., & Vieites, D. R. (2002). Phenotypic plasticity of anuran
767 larvae: environmental variables influence body shape and oral morphology in *Rana*
768 *temporaria* tadpoles. *J. Zool.* **257**, 155-162. <https://doi.org/10.1017/S0952836902000754>

769 Venesky, M. D., Wassersug, R. J., & Parris, M. J. (2010). The impact of variation in labial
770 tooth number on the feeding kinematics of tadpoles of southern leopard frog (*Lithobates*
771 *sphenocephalus*). *Copeia* **2010**, 481-486. <https://doi.org/10.1643/CG-09-093>

772 Verrell, P. A. (1999). Geographic variation in sexual behavior: sex, signals and speciation. In
773 *Geographic variation in behavior: Perspectives on evolutionary mechanisms: 262-286.*
774 Foster, S. A., & Endler, J. A. (Eds.). Oxford University Press.

775 Vidal, M. A., Moreno, P. I., & Poulin, E. (2012). Genetic diversity and insular colonization of
776 *Liolaemus pictus* (Squamata, Liolaeminae) in north-western Patagonia. *Austral Ecol.* **37**,
777 67-77. <https://doi.org/10.1111/j.1442-9993.2011.02241.x>

778 Villagrán, C., Armesto, J. J., & Leiva, R. (1986). Recolonización postglacial de Chiloé
779 insular: evidencias basadas en la distribución geográfica y los modos de dispersión de la
780 flora. *Rev. Chil. Hist. Nat.* **59**, 19-28.

781 Vögeli, M., Laiolo, P., Serrano, D., & Tella, J. L. (2008). Who are we sampling? Apparent
782 survival differs between methods in a secretive species. *Oikos* **117**, 1816-1823.
783 <https://doi.org/10.1111/j.1600-0706.2008.17225.x>

784 Warkentin, K. M. (1995). Adaptive plasticity in hatching age: a response to predation risk
785 trade-offs. *P. Roy. Soc. B-Biol. Sci.* **92**, 3507-3510.

786 Weaver, S. J., Callaghan, C. T., & Rowley, J. J. (2020). Anuran accents: Continental-scale
787 citizen science data reveal spatial and temporal patterns of call variability. *Ecol. Evol.* **10**,
788 12115-12128. <https://doi.org/10.1002/ece3.6833>

789 Wells, K.D. (1977) The social behaviour of anuran amphibians. *Anim. Behav.* **25**, 666–693.
790 [https://doi.org/10.1016/0003-3472\(77\)90118-X](https://doi.org/10.1016/0003-3472(77)90118-X)

791 Wells, K.D. & Schwartz, J.J. (2007) The behavioral ecology of anuran communication. In
792 *Hearing and sound communication in amphibians*: 44–86. Narins, P. M., & Feng, A. S.
793 (Eds.), New York: Springer Verlag

794 Wickham, H., Chang, W., & Wickham, M. H. (2016). Package ‘ggplot2’. Create Elegant Data
795 Visualisations Using the Grammar of Graphics. Version, **2**, 1-189.

796 Wilczynski, W., & Ryan, M. J. (1999). Geographic variation in animal communication
797 system. In *Geographic variation in behavior: Perspectives on evolutionary mechanisms*:
798 234-261. Foster, S. A., & Endler, J. A. (Eds.). Oxford University Press.

- 799 Wilhelm, O. (1932). Nuevas observaciones acerca de la neomelia de la *Rhinoderma darwini*.
800 *Rev. Chil. Hist. Nat.* **36**, 166-170
- 801 Wolley, S. M., & Moore, J. M. (2011). Coevolution in communication senders and receivers:
802 vocal behavior and auditory processing in multiple songbird species. *Ann. Ny. Acad. Sci.*
803 **1225**, 155. <https://doi.org/10.1111/j.1749-6632.2011.05989.x>
- 804 Zippel, K.C. (1997). Hairy frogs (*Trichobatrachus robustus*) in the field. *Reptiles* **5**, 68–73.
- 805 Ziegler, L., Arim, M., & Bozinovic, F. (2016). Intraspecific scaling in frog calls: the interplay
806 of temperature, body size and metabolic condition. *Oecologia* **181**, 673-681.
807 <https://doi.org/10.1007/s00442-015-3499-8>

808 **Supplementary information**

Table S 1. Averages and coefficients of intra-individual variation (in cursive) of the acoustic variables analysed in the advertisement calls of *Rhinoderma rufum* from Las Carmelitas. Coefficient of variation was not computed for Call rate as a single recording of successive calls was carried out for most individuals. Call rate was not computed for one individual for which a single call was recorded.

Individuals	Number of calls	Notes per call	Call rate (Calls/min)	Call duration (s)	Note duration (ms)	Inter-note interval (ms)	Dominant frequency (Hz)	Chaos proportion (%)
1	4	5	1.805	1.23 2.9	0.14 7.8	0.13 26.8	2939.3 2.1	NR NR
2	1	5	–	1.22 NR	0.14 6.2	0.13 18.9	2894.1 2.7	35.2 51.1
3	4	2	1.116	0.64 0.41	0.18 6.7	0.26 22.27	2885.5 1.6	NR NR
4	4	5	1.846	1.57 20.7	0.17 8.1	0.18 18.6	3130.9 2.1	21.7 1.4
5	4	5	2.308	1.57 3.0	0.15 13.7	0.20 24.9	2851 1.9	4.1 0
6	3	5	1.488	1.67 1.5	0.16 11.9	0.21 18.0	2968.7 3.1	NR NR
7	3	6	1.978	2.08 1.5	0.20 7.5	0.18 10.4	2990.7 2.2	15.6 28.4
8	3	5	0.865	1.93 1.4	0.21 6.8	0.22 10.5	2876.8 2.5	38.1 47.2
9	2	5	1.667	1.92 0	0.20 6.8	0.23 12.2	2842.4 0	38.8 33.3